African nightshades (Solanum nigrum complex): The potential contribution to human nutrition and livelihoods in sub-Saharan Africa.

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African nightshades (Solanum nigrum complex): The potential contribution to human nutrition and livelihoods in sub-Saharan Africa

Frank Sangija | Haikael Martin | Athanasia Matemu

Abstract

Achieving zero hunger in sub-Saharan Africa (SSA) without minimizing postharvest losses of agricultural products is impossible. Therefore, a holistic approach is vital to end hunger, simultaneously improving food security, diversity, and livelihoods. This review focuses on the African nightshades (ANS) Solanum spp. contribution to improving food and nutrition security in SSA. Different parts of ANS are utilized as food and medicine; however, pests and diseases hinder ANS utilization. African nightshade is rich in micronutrients such as β-carotene, vitamins C and E, minerals (iron, calcium, and zinc), and dietary fiber. The leaves contain a high amount of nutrients than the berries. Proper utilization of ANS can contribute to ending hidden hunger, mainly in children and pregnant women. Literature shows that ANS contains antinutritional factors such as oxalate, phytate, nitrate, and alkaloids; however, their quantities are low to cause potential health effects. Several improved varieties with high yields, rich in nutrients, and low alkalooids have been developed in SSA. Various processing and preservation techniques such as cooking, drying, and fermentation are feasible techniques for value addition on ANS in SSA; moreover, most societies are yet to adopt them effectively. Furthermore, promoting value addition and commercialization of ANS is of importance and can create more jobs. Therefore, this review provides an overview of ANS production and challenges that hinder their utilization, possible solutions, and future research suggestions. This review concludes that ANS is an essential nutritious leafy vegetable for improving nutrition and livelihoods in SSA.

KEYWORDS
African nightshade, diversity, nutritional, postharvest losses, preservation, utilization

Abbreviations: AIV, African indigenous vegetable; ANS, African nightshade; BNS, Black nightshade; BW, body weight; CF, Controlled fermentation; DW, Dry weight basis; EFSA, European Food Safety Authority; FAO, Food and Agriculture Organization of the United Nations; FW, Fresh weight basis; HCN, hydrogen cyanide; KSH, Kenyan Shilling; LAB, Lactic acid bacteria; PaP, Processing and preservation; PHH, Postharvest handling; PHL, Postharvest losses; RDA, Recommended daily allowance; SF, Spontaneous fermentation; SGA, Steroidal glycoalkaloid; SSA, sub-Saharan Africa; TGA, Total glycoalkaloids; TM, Traditional medicine; TZS, Tanzanian shilling; UA, Urban agriculture; WHO, World Health Organization; WVC, World Vegetable Center.
1 | INTRODUCTION

African nightshades (ANSs) are among many underutilized and neglected African indigenous vegetables (AIVs) species; if adequately exploited, they could improve food, nutrition, and income among the rural population (Abukutsa, 2003; Abukutsa-Onyango, 2007; Dinssa et al., 2016). However, Weinberger and Msuya (2004) argued that AIVs are not underutilized as usually thought but somewhat are undervalued. ANS belongs to many species in the genus Solanum in the family Solanaceae found in temperate and tropical regions of the world, and it consists of about 90 genera and 2000 to 3000 species (Edmonds & Chweya, 1997; Yang & Ojiewo, 2013). Within this family, Solanum forms the largest and most complex genus composed of more than 1500 species (Edmonds & Chweya, 1997).

ANS is among the known AIVs rich in nutrients to promote food and nutrition security in sub-Saharan Africa (SSA) (Yang & Ojiewo, 2013). The AIVs are considered a new cash crop in most SSA regions because they contribute to income generation to individuals and households (Shackleton et al., 2009). AIVs have been part of SSA’s food systems for generations, and their leaves, young shoots, and flowers are consumed for various purposes (Abukutsa-Onyango, 2010; Ambrose-Oji, 2009; Yang & Ojiewo, 2013). ANSs are among high-priority AIVs with the potential for health, nutrition, and economic benefits (Edmonds & Chweya, 1997; Yang & Ojiewo, 2013). They are rich in macro- and micronutrients, including thiamine, ascorbic acid, iron, calcium, zinc, protein, and dietary fiber (Kirigia et al., 2019; Ontita et al., 2017; Ronoh et al., 2017; Yuan et al., 2018). Furthermore, they are rich in bioactive compounds, particularly lutein, zeaxanthin, polyphenol, flavonoids, and chlorophylls, which possess antioxidant activity, anti-genotoxicants, and anticancer properties (Odongo et al., 2018). In SSA, ANS is exploited for food, medicines, animal feed, and spiritual uses, but less exploited for socioeconomic benefits (Abukutsa-Onyango, 2007; Ontita et al., 2017). However, AIVs are wasted (Global Panel, 2018; Stevens et al., 2015) despite the favorable climatic conditions and high production; they account for 50% of postharvest losses (PHL) of all food produced in SSA (Global Panel, 2018; Weinberger & Msuya, 2004).

This review focuses on harnessing the ANS potential in SSA. It explores the ANS diversity, cultivation, nutritional and functional benefits, antinutritional factors, and safety. Further, it summarizes and discusses the impact of PHL, postharvest handling (PHH) and processing, and traditional recipes for ANS. Importantly, the ANS roles in health promotion, trends in utilization and possible constraints, and possible solutions are summarized.

2 | DIVERSITY OF ANS

The commonly available ANS species in SSA include S. americanum, S. scabrum, S. nigrum, and S. villosum (Table 1) (Edmonds & Chweya, 1997; Keller, 2004; Yang & Ojiewo, 2013; Yuan et al., 2018). Eight species belong to the S. nigrum complex or the black nightshades (BNSs) complex, namely, S. nigrum L., S. americanum, S. scabrum Mill., S. sarrachoides Sendtn., S. villosum Mill., S. grossedentatum A. Rich., S. florulentum Bitter, and S. tarderemotum Bitter, which are distinguished based on characteristic morphological traits (Dehmer & Hammer, 2004; Edmonds & Chweya, 1997; Ojiewo, Mwai et al., 2013; Ronoh et al., 2017). Solanum scabrum and S. villosum are most prevalent in East Africa (Table 1) (Ambrose-Oji, 2009) but are produced in other regions of Africa (Table 1 and Figure 1). Nonetheless, they are still considered a wild weedy crop in most SSA (Dinssa et al., 2016; Edmonds & Chweya, 1997). They can be grown in various places such as roadsides, hedgerows, around buildings and houses, under trees, as garden weeds, riverbanks and in gullies, on forest and grassland margins, quaysides and rubbish tips, on shingle beaches, on railway cuttings, and the edges of cultivated fields and plantations (Edmonds & Chweya, 1997). About 45% of farmers in Mbale, Uganda, collect ANSs from the wild for selling in the urban markets (Ambrose-Oji, 2009; Kasambula et al., 2007). Approximately 90% of vegetable supplies in big cities in SSA, including the Central Africa Republic, Guinea-Bissau, Madagascar, and Tanzania, are predicted to come from urban and peri-urban agriculture (UA) (Ambrose-Oji, 2009). Most of these vegetables are exotic and highly consumed in urban and peri-urban regions (Ambrose-Oji, 2009). However, many fresh AIVs leaves available in urban markets come from rural areas due to seasonal production (Gido et al., 2017).

Solanum nigrum, commonly known to be native to Eurasia, was introduced in the Americas, South Africa, and Australasia (Kuete et al., 2017). Whereby, S. americanum (American BNS) is native to the Americas (Dehmer & Hammer, 2004) and is the most unrelated species within the S. nigrum complex with only about 43% of genetic similarity (Gilbert, 2006). Within the S. nigrum complex, S. scabrum and S. nigrum are near related species (about 68%), with the S. villosum being close to them by 62% (Dehmer & Hammer, 2004; Gilbert, 2006). American BNSs are native to the Americas, particularly Cuba and South America (Dehmer & Hammer, 2004;
In the Americas, this species is found in California, Mexico, Central America, and South America. The nomenclature and taxonomy are associated with toxic BNS (*Atropa belladonna*) of temperate Eurasian origin; this species has many phenotypic similarities with many nightshades (Gilbert, 2006). ANS has been consumed for centuries by native peoples in Central America, Mexico, and Africa and is essential in these regions (Lotter et al., 2014). In SSA, Benin, Cameroon, Burkina Faso, Tanzania, and Kenya highly consume ANS (Keller, 2004; Weinberger & Msuya, 2004; World Vegetable Center [WVC], 2017), and it is also well promoted in the Southern African Development Community (SADC) region (Ojiewo, Mbwambo et al., 2013). Table 1 shows the diversity and distribution of ANS in SSA. In Tanzania, the northern (Arusha and Kilimanjaro), central (Dodoma), eastern (Morogoro and Tanga), and southern (Iringa and Mbeya) zones are well-known for ANS production (Keller, 2004; Weinberger & Msuya, 2004). Both traditional and exotic vegetables cover 9% of all cultivated land (Weinberger & Msuya, 2004; Dinssa et al., 2016). In East Africa, some improved cultivars of *S. scabrum* and *S. villosum* were released based on their superior yield and acceptability. Thirteen cultivars of *S. scabrum* (ACC.15B, ACC.16A, ACC.16B, ACC.18, ACC.20, ACC.21, ACC.3, ACC.4, ACC.6, ACC.7, and ACC.8B) and *S. villosum* (ACC.25 and ACC.29) were released in Kenya (Ronoh et al., 2019). In Tanzania, cultivars of *S. scabrum* (BG16-Nduruma and

<table>
<thead>
<tr>
<th>African nightshades</th>
<th>Description</th>
<th>Distribution</th>
<th>References</th>
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<tbody>
<tr>
<td><em>Solanum scabrum</em> Mil</td>
<td>Broad leaves Ripe berries (1000 to 2000 berries); 1000 seeds weight of 1.0 to 1.3 g Black or purple leaves of greater than 14 mm in size Differ in growth habits, bitterness, and leaf color Taller up to 1.5 m in height Low rainfall and temperature (18 to 30 °C) Good sources of carotenoid, vitamin C, E and A, C, calcium, iron, zinc, and protein Used as food and medicine</td>
<td>East Africa, West Africa, Central Africa</td>
<td>Abukutsa-Omyango, 2015; Edmonds and Chweya, 1997; Maundu et al., 1999; Wafula, 2017; Yang and Ojiewo, 2013</td>
</tr>
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<td><em>Solanum nigrum</em> Miller</td>
<td>Black nightshades, leafy green vegetables (<em>S. eldoretii</em>, <em>S. tarderemotum</em>, and <em>S. florilegium</em>) Berries about 3000 to 4000; 1000 seeds weight of 0.4-0.6 g Good sources of carotenoid, vitamin C, E and A, C, calcium, iron, zinc, and protein Used as food and medicine</td>
<td>North Africa, Kenyan Highlands, Northern Tanzania</td>
<td>Edmonds and Chweya, 1997; Jacoby et al., 2003; Maundu et al., 1999; Shackleton et al., 2009; Wafula, 2017; Yang and Ojiewo, 2013</td>
</tr>
<tr>
<td><em>Solanum americanum</em> Miller</td>
<td>Black nightshades; <em>Mnavu</em>, <em>Msogo</em>, and <em>Mhaki</em> (Tanzania) and <em>Wsuggaenzirugavu</em> (Uganda) Berries are shiny dark purple, and the barriers are small than 12 mm Small berries about 4500; 1000 seeds weigh 0.3 to 0.4 g Moderately thin branches than other species Good sources of carotenoid, vitamin C, E and A, C, calcium, iron, zinc, and protein Used as food and medicine</td>
<td>East Africa, West Africa, Central Africa, Southern Africa</td>
<td>Edmonds and Chweya, 1997; Maundu et al., 1999; Shackleton et al., 2009; Wafula, 2017; Yang and Ojiewo, 2013</td>
</tr>
<tr>
<td><em>Solanum villosum</em> Miller</td>
<td>Black nightshades or garden huckleberry; <em>Mnavu</em> (Tanzania and Kenya) Berries about 3000 to 4000 (orange to yellow); 1000 seeds weight of 0.4 to 0.6 g Wild and cultivated Used as food and medicine</td>
<td>East Africa, West Africa, Central Africa, Southern Africa, Egypt</td>
<td>Edmonds and Chweya, 1997; Maundu et al., 1999; Shackleton et al., 2009; Wafula, 2017; Yang and Ojiewo, 2013</td>
</tr>
</tbody>
</table>
FIGURE 1 Leaves, flowers, and berries of *S. nigrum*, *S. villosum*, and *S. scabrum*: a, e, and i, respectively. Unripe berries, ripe berries, and flower of *S. nigrum*: b, c, and d, respectively. Unripe berries, ripe berries, and flowers of *S. villosum*: f, g, and h, respectively. Unripe berries, ripe berries, and flower of *S. scabrum*: j, k, and l, respectively. Source: Authors.

SS49-Olevolosi), and RC18-ES13-3-Ambureni (*S. villosum*) and RC10-ES13-3-Malala (*S. scabrum*) were released in 2011 and 2018, respectively ([https://www.tosci.go.tz/publications/22](https://www.tosci.go.tz/publications/22); Ojiewo, Mbwambo, et al., 2013; Mbwambo et al., 2021). Also, about five advanced lines of *S. scabrum* (SS52, SS40, SS042, ABUK1, and ABUK2) and one of *S. villosum* (ABUK3) are available in Kenya and Tanzania (Ronoh et al., 2019).

3 | CULTIVATION OF ANS

ANS are a popular and cash-generating vegetable in Africa (Yang et al., 2013). The production of ANS requires a small portion of land, and it does not require extensive external inputs and production experts. Therefore, it encourages the majority of farmers to engage in its production (Abukutsa-Onyango, 2007). Demands for AIVs have surged in Africa due to rising market requirements for AIVs varieties (Yang et al., 2013). Small-scale production of ANS is common in Kenya; about 80% of farmers use less than 0.25 acres on production (Abukutsa-Onyango, 2007; Ambrose-Oji, 2009), characterized by low leaf yields of 1.5 to 3 tons/ha specifically for *S. scabrum*, *S. villosum*, and *S. americanum* (Ojiewo, Mbwambo, et al., 2013). A low level of production of ANS hinders the availability and consequently results in low consumption. Onyango et al. (2016) indicated that about 75% to 80% of ANS growers in Kenya are women cultivating small plots of less than 0.25 acres. About 59% of all farmers produce less than 50 kg of ANS per season (Abukutsa-Onyango, 2007; Ambrose-Oji, 2009; Gebru, 2015). AIVs are grown mainly in home gardens within the homestead (Abukutsa-Onyango, 2007), as a source of food and small income generation for their family; it also helps them become financially independent (Onyango et al., 2016).

3.1 | Cropping system of ANS

Mono-cropping, intercropping, and crop rotation are cropping systems used in the growing of ANS. However, most farmers practice mono-cropping, followed by intercropping, and few farmers practice crop rotations. There are several benefits of intercropping that include a diversity of crops in a given season and optimal utilization of resources such as nutrients, water, and light (Gebru, 2015; Onyango et al., 2016). ANS can be intercropped with maize, millet, sweet potatoes, kale, beans, avocado, cassava, groundnuts, and bananas (Abukutsa-Onyango, 2007). The current global trend is encouraging organic farming by avoiding using harmful chemicals to the environment and consumers (Abukutsa-Onyango, 2007). The planting of ANS is carried out twice in Kenya during the long rains (March to July) and the short rains (September to December) (Abukutsa-Onyango, 2007). About 69% of Kenya’s farmers depend on rainfall, and only 20% to 31% are practicing irrigation using watering cans (Abukutsa-Onyango, 2007; Gebru, 2015).

3.2 | Yields of ANS

ANS edible leaves give the highest yield between 12 and 50 tons/ha per season (Abukutsa-Onyango, 2015; Edmonds & Chweya, 1997; Ojiewo, Mbwambo, et al., 2013). However, Ojiewo, Mbwambo, et al. (2013) and Molina et al. (2020) reported an annual yield of 1.5–3.0 and 3.85 tons/ha, respectively. The average share of cultivated ANS varies across locations, and the most substantial proportion is the Arumeru district in Arusha, Tanzania, which covers 20% of all cultivated land (Dinssa et al., 2016; Weinberger & Msuya, 2004). Arumeru district produces about 71.8% of all cultivated ANS in Tanzania (Weinberger &
Msuya, 2004). The presence of national and international research centers, including the World Vegetable Center, Eastern and Southern Africa (WVC-ESA), International Institute of Tropical Agriculture (IITA), and Nelson Mandela African Institution of Science and Technology (NM-AIST), positively promotes cultivation and consumption of AIVs, including ANS around these regions (Dinssa et al., 2016).

ANS is tolerant to abiotic stress under low soil moisture and heat. Luoh et al. (2014) reported that ANS S. scabrum retains vitamins and minerals and undergoes less weight loss at water deficit. Thus, ANS suffers less in water deficiency and becomes a choice of crops under drought conditions in SSA. ANS can take 3 to 4 weeks for the first harvest; therefore, it is an essential crop to feed the world’s large population (Dinssa et al., 2016; Lobell & Gourdji, 2012).

Rapid urbanization has increased the demand for ANS in SSA (Ambrose-Oji, 2009; Shackleton et al., 2009). There is considerable scope to increase ANS yield; therefore, variation between yield obtained in farms and research has been reported (Dinssa et al., 2016). This variation further suggests the importance of technological application for the intense production of ANS in the farms.

3.3  Factors affecting cultivation of ANS

3.3.1  Pests and diseases

Pests and diseases are the main challenges during ANS production in SSA, resulting in low yields. Fungi, bacteria, and viruses are the main causative agents for ANS diseases. The major fungi diseases include damping-off (Rizoctonia spp.), early blight (Alternaria solani), late blight (Pyrchritium infestans), fusarium wilt (Fusarium oxysporum and Fusarium solanii), and verticillium wilt (Verticillium dahliae). The bacteria diseases are bacterial wilt (Ralstonia solanacearum), leaf mold (Cladosporium oxysporum), eyespot, and southern blight (Abukutsa-Onyango, 2007; Ambrose-Oji, 2009; Dinssa et al., 2016; Edmonds & Chweya, 1997; MoALF/SHEP PLUS, 2019; Mono-Womdim et al., 2012; Onyango et al., 2016; Shackleton et al., 2009; Yang & Ojewo, 2013).

Pests and diseases damage the leafy structure and reduce the quality of vegetables, leading to customers’ rejection (Abukutsa-Onyango, 2007). Enterobacter mori isolated from ANS pickle (Wafula, 2017) cause plant bacterial wilt (Zhu et al., 2011); perhaps they also cause wilt in ANS. The pest and diseases of ANS are the same as of other Solanaceae families. Pests and diseases are a significant problem in Tanzania; however, it becomes more intense when vegetables are grown for multiple harvests over a long time or used for seed production (Keller, 2004).

On the other hand, viruses control insects (aphids, whiteflies) and inadvertently bruising young leaves by touching them (Mono-Womdim et al., 2012; Ojewo, Mbwambo, et al. 2013). The viral diseases include leaf curl viruses, leaf mosaic viruses, yellow viruses, and tomato mosaic viruses (Mono-Womdim et al., 2012; Wafula et al., 2017). Tomato mosaic virus (ToMV) is a common virus found in ANS in SSA (Mono-Womdim et al., 2012). The control measures for pathogenic microorganisms include use of fungicides, disease free seeds, minimizing injury, avoiding dense planting, using furrow or drip irrigation, and crop rotation. Further, removing infected plants and destroying them immediately after harvest, avoiding overfertilization, adequate sanitation, and using healthy seedlings can control pests and diseases (Mono-Womdim et al., 2012).

The most common pests of ANS include aphids (Aphis sp.), spider mite (Tetranychus evansi Baker), red and black ants, cutworms, caterpillars (larvae), grasshoppers (Zonocerus variegaatus), whiteflies (Bemisia tabac, beetles (Epilachna hirta, Lagria spp., and Podagrica spp.), and Nematodes (Melodigyne spp.) (MoALF/SHEP PLUS, 2019; Mono-Womdim et al., 2012).

The short shelf life of ANS reduces the market supply and bargaining power of small-scale farmers and local open market sellers (Dinssa et al., 2016; Muhanji et al., 2011). Therefore, a multidisciplinary approach is needed between breeders and postharvest specialists to improve vegetables’ shelf life and storage conditions. Interventions through affordable preservation techniques can increase small-scale farmers’ income and the vegetables’ marketing (Dinssa et al., 2016; Muhanji et al., 2011).

3.3.2  Lack of improved cultivars

Lack of improved cultivars limits the high production of ANS in SSA, resulting in low-yielding cultivars (Dinssa et al., 2016). Seed manufacturing companies do not consider the production and marketing of ANS seeds as a profit-generating business because most AIVs are open-pollinated, forcing most farmers to produce their seeds (Dinssa et al., 2016). Nevertheless, the seeds of improved ANS cultivars are not available in seed stores of many regions in SSA, particularly in remote areas (Dinssa et al., 2016; Muhanji et al., 2011). As of recently, some improved cultivars of S. scabrum and S. villosum are available to farmers in Tanzania and Kenya for commercial use (Ronoh et al., 2019; https://www.tosci.go.tz/publications/22). Besides, several advanced lines of S. villosum and S. scabrum are available to advance the breeding research of the crop for improved characteristics such as yield, pests, and disease resistance (Ronoh et al., 2019).
3.3.3 Other factors

Other factors that hinder the production and consumption of ANS include inadequate rainfall, lack of knowledge, lack of appropriate storage facilities, low-quality seeds, drought, fragmented marketing channels, poor transport infrastructure, agronomic challenges, and lack of appropriate packaging (Abukutsa-Onyango, 2007; Ambrose-Oji, 2009). For better-improving ANS production, researchers should involve farmers as they are key producers. Improved cultivars are essential because they require fewer inputs, tolerate pests and diseases, and tolerate different climate conditions (Muhanji et al., 2011).

4 NUTRITIONAL AND FUNCTIONAL BENEFITS OF ANS

4.1 Macro and micronutrients

ANS' leaves and berries contain a high amount of protein, carbohydrates, minerals, and vitamins (A, C, E, and B complex) and could contribute to a healthy diet (Tables 2 and 3; Ambrose-Oji, 2009; Stoll et al., 2021; Wafula, et al., 2017). The high content of micronutrients is sufficient to contribute to the recommended daily allowance (RDA) (Table 4). According to Food and Agriculture Organization of the United Nations (FAO) and World Health Organization (WHO) (FAO & WHO, 2004) and Agudo & Joint FAO/WHO (2004), the RDA of vegetables and fruits is 400 g, indicating that ANS consumption alone can meet the RDA of vitamins C, E, and A, iron, and manganese. ANSs contain β-carotene (2.8 to 13.8 mg/100 g fresh weight basis [FW]) higher than the average level of various vegetables reported by the USDA National Nutrient Database (Table 2; Yuan et al., 2018). For instance, 100 g of leaves of ANS can provide over 100% of the vitamin A needs of pregnant women (Schreinemachers et al., 2018; WVC, 2017). ANS contains a significant amount of B vitamins including thiamine (0.08 to 0.35 mg/100 g FW), riboflavin (0.17 to 0.19 mg/100 g FW), and folate (12 to 56 μg/100 g FW) (Table 2). The B vitamins function as co-enzymes and help in energy production from carbohydrates, synthesis of neurotransmitters, fatty acids, and hormones (Blake, 2008). Also, ANS contains a high content of vitamin C (140 mg/100 g FW) and vitamin E (1.21 to 5.94 mg/100 g FW), which are natural antioxidants (FAO, 2004 (FAO & WHO, 2004; Blake, 2008). Young leaves of ANS have low vitamin C content than matured leaves (Cheptoo et al., 2019). Vitamin C prevents scurvy, reduces gastric cancer, and stabilizes folate in food and plasma. The dietary intake of 25 mg of vitamin C improves iron absorption and prevents anemia (FAO & WHO, 2004). The ANS leaves are rich in iron (1.3 to 7.2 mg/100 g FW), zinc (0.1 to 0.56 mg/100 g FW), and calcium (173 to 199 mg/100 g FW) (Table 2). Solanum retroflexum leaves contain 7.2 mg/100 g of iron (Sivakumar et al., 2020). Calcium content at a concentration of 300-600 mg inhibited 60% iron absorption (FAO & WHO, 2004). Increasing iron intake and avoiding foods rich in calcium and iron at the same time can improve iron absorption (FAO & WHO, 2004). Calcium contributes to bone development; the deficiency led to the development of osteoporosis. Zinc contributes to the improvement of the immune system and repairing of cell and organ integrity. Deficiency of zinc in humans results in growth retardation, bone maturation, delayed sexual maturity, diarrhea, skin lesions, reduced appetite, and increased vulnerability to infections (FAO & WHO, 2004).

Solanum nigrum leaves contain a high amount of protein, fiber, and total ash than the same species’ berries. However, the berries contain more fat and carbohydrates (Tables 2 and 3). In comparison, S. nigrum leaves contain a high amount of calcium, iron, magnesium, phosphorous, potassium, sodium, and zinc than the berries (Tables 2 and 3). Also, the leaves of S. nigrum contain a high amount of vitamins A, C, and B9 than the berries; besides, the berries contain a high amount of vitamin B1. The high contents of the macro- and micronutrients present in ANS leaves suggest that leaves are more nutritious than the berries despite some nutrients being high in the berries. Therefore, researches are needed to assess the nutrients contents of berries in other species of ANS. Notably, the ANSs’ dense and diverse nutrients can improve essential nutrients for better health if sufficiently eaten (Table 4).

4.2 Phytochemicals

Phytochemicals are secondary metabolites present in abundance in various parts of ANS. Total phenols, carotenoids, glycoalkaloids, and chlorophylls are phytochemicals in ANS (Table 5) (Neugart et al., 2017; Nyaga et al., 2019; Nyaga, 2020; Yuan et al., 2018). Solanum nigrum var. sarrachoides leaves contain flavonoids, alkaloids, tannins, saponin phenols, phytosterols, coumarins, and glycosides. In contrast, S. villosum leaves showed all the phytochemicals, except phytosterols and coumarins (Nyaga et al., 2019; Nyaga, 2020). These phytochemicals are also bioactive compounds in ANS with potential health benefits. Solanum scabrum contains many carotenoids such as β-caroteno, zeaxanthin, and lutein (Odongo et al., 2018). Carotenoid content in foods contributes to
### TABLE 2 Nutrient content of the raw and processed ANS species

<table>
<thead>
<tr>
<th>Nutrients</th>
<th>Raw&lt;sup&gt;e,f,g,j,m&lt;/sup&gt; S. nigrum Mill.</th>
<th>Solar drying&lt;sup&gt;l&lt;/sup&gt;</th>
<th>Blanched&lt;sup&gt;j&lt;/sup&gt;</th>
<th>Fermented</th>
<th>Raw&lt;sup&gt;e,j,k,l,m&lt;/sup&gt; S. scabrum Mill.</th>
<th>Dried&lt;sup&gt;h&lt;/sup&gt;</th>
<th>Blanched&lt;sup&gt;b&lt;/sup&gt;</th>
<th>Fermented&lt;sup&gt;b&lt;/sup&gt; S. villosum Mill.</th>
<th>Raw&lt;sup&gt;e,m&lt;/sup&gt;</th>
<th>Blanched</th>
<th>Dried</th>
<th>Fermented</th>
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<tr>
<td>Moisture (%)</td>
<td>87.7</td>
<td>6.5 to 12.3</td>
<td>98.5</td>
<td>–</td>
<td>858</td>
<td>–</td>
<td>89.3</td>
<td>–</td>
<td>–</td>
<td>–</td>
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<td>–</td>
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<tr>
<td>Protein (g)</td>
<td>39.8 DW/3.4 FW</td>
<td>36.1 to 38.2 DW</td>
<td>40.6 DW</td>
<td>–</td>
<td>3 to 6 FW</td>
<td>–</td>
<td>–</td>
<td>14 to 16 DM</td>
<td>4.2 FW</td>
<td>–</td>
<td>–</td>
<td>–</td>
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<tr>
<td>Energy (KJ)</td>
<td>162</td>
<td>–</td>
<td>162</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
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<tr>
<td>Fat (g)</td>
<td>7.6 DW</td>
<td>4.14 DW</td>
<td>7.28 DW</td>
<td>–</td>
<td>3.1 FW</td>
<td>–</td>
<td>–</td>
<td>1.9 FW</td>
<td>–</td>
<td>–</td>
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<td>–</td>
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<tr>
<td>Fiber (g)</td>
<td>2.5 FW/14.07 DW</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>12.8 DW</td>
<td>11.6 DW</td>
<td>12.05 DW</td>
<td>–</td>
<td>25.1 DW</td>
<td>–</td>
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</tr>
<tr>
<td>Dry matter (g)</td>
<td>8 FW</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>10.5 to 22 DW</td>
<td>–</td>
<td>–</td>
<td>21.7 to 22.4 DW</td>
<td>11.1</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Carbohydrate (g)</td>
<td>29.04 DW/3.78 FW</td>
<td>14 to 17.1 DW</td>
<td>60.7 DW</td>
<td>–</td>
<td>0.3 DW</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Total ash (%)</td>
<td>10.6 to 14.6 DW/1.38 to 1.9 FW</td>
<td>11.5 to 12.3 DW</td>
<td>8.8 DW</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Ca (mg)</td>
<td>173 to 199 FW</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>1460 to 2430 DW</td>
<td>–</td>
<td>–</td>
<td>3.3 FW</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Fe (mg)</td>
<td>1.3 to 7.2 FW</td>
<td>–</td>
<td>6.9 FW</td>
<td>–</td>
<td>70 to 130 DW</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Mg (mg)</td>
<td>25 to 92 FW</td>
<td>–</td>
<td>87 FW</td>
<td>–</td>
<td>330 to 410 DW</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>NA</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>P (mg)</td>
<td>36 to 75 FW</td>
<td>–</td>
<td>33 FW</td>
<td>–</td>
<td>270 to 320 DW</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>K (mg)</td>
<td>257 to 430 FW</td>
<td>–</td>
<td>232 FW</td>
<td>–</td>
<td>100 FW</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Na (mg)</td>
<td>3 to 8 FW</td>
<td>–</td>
<td>8 FW</td>
<td>–</td>
<td>7422 FW</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Zn (mg)</td>
<td>0.1 to 0.36 FW</td>
<td>–</td>
<td>0.53 FW</td>
<td>–</td>
<td>4.0 to 6.2 FW</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>0.8 FW</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Cu (mg)</td>
<td>0.16 FW</td>
<td>–</td>
<td>0.15 FW</td>
<td>–</td>
<td>2.6 to 4.2 DW</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Mn (μg)</td>
<td>2.1 FW</td>
<td>–</td>
<td>2.1 FW</td>
<td>–</td>
<td>257 to 28.8 DW</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>β-Carotene (mg)</td>
<td>2.8 to 14.2 FW/102 DW</td>
<td>1.7 to 69.4 DW</td>
<td>119 DW</td>
<td>–</td>
<td>5.3 to 10 FW/71.22 DW</td>
<td>47.64 DW</td>
<td>54.8 DW</td>
<td>11.8 FW</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Vitamin A (μg)</td>
<td>5 FW</td>
<td>–</td>
<td>3</td>
<td>–</td>
<td>8.8 FW</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Vitamin C (mg)</td>
<td>35 FW/62.9 to 75.2 DW</td>
<td>–</td>
<td>3</td>
<td>–</td>
<td>40 to 140 FW/63 to 177.97 DW</td>
<td>2.4 to 103.7 DW</td>
<td>10.89.4 DW</td>
<td>256.8 DW</td>
<td>2.9 FW</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Vitamin B1 (mg)</td>
<td>0.03 FW/0.71 DW</td>
<td>–</td>
<td>0.07 FW</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>0.21 to 1.5 DW</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Vitamin B2 (mg)</td>
<td>0.17 to 0.19 FW/15 DW</td>
<td>–</td>
<td>0.16 FW</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>0.98 to 1.1 DW</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Vitamin B9 (μg)</td>
<td>12 to 56 FW</td>
<td>–</td>
<td>0.07 FW</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Vitamin E (mg)</td>
<td>9.3 to 45.7 DW/1.21 to 5.94 FW</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>6.4 to 14.2 DW</td>
<td>–</td>
<td>–</td>
<td>44.5 to 48 DM</td>
<td>11.4 DW</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
</tbody>
</table>

**Note:** The nutrients are expressed per 100 g.

**Abbreviations:** DW, dry weight basis; FW, fresh weight basis.

**Sources:**<sup>e</sup>Gogo et al. (<2016>);<sup>g</sup>Yuan et al. (<2018>);<sup>a</sup>Ronoh et al. (<2017>);<sup>f</sup>Van Jaarsveld et al. (<2014>);<sup>j</sup>Nyambaka et al. (<2012>);<sup>h</sup>Cheptoo et al. (<2009>);<sup>i</sup>Mibe et al. (<2011>);<sup>b</sup>Traoré et al. (<2017>);<sup>l</sup>Habwe et al. (<2008>);<sup>k</sup>Kirigia et al. (<2019>);<sup>m</sup>Yang and Ojiewo (<2013>);<sup>n</sup>Wafula (<2017>).
TABLE 3  Nutrient content of ANS berry

<table>
<thead>
<tr>
<th>Nutrients</th>
<th>S. nigrum Mill.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Moisture (%)</td>
<td>76.86</td>
</tr>
<tr>
<td>Protein (%)</td>
<td>17.63 DW/4.08 FW</td>
</tr>
<tr>
<td>Fat (%)</td>
<td>12.18 DW/2.82 FW</td>
</tr>
<tr>
<td>Energy KJ</td>
<td>43.54 DW/10.08 FW</td>
</tr>
<tr>
<td>Fiber (%)</td>
<td>6.29 DW/1.46 FW</td>
</tr>
<tr>
<td>Carbohydrate (%)</td>
<td>55.85 DW/12.92 FW</td>
</tr>
<tr>
<td>Total ash (%)</td>
<td>8.05 DW/1.86 FW</td>
</tr>
<tr>
<td>Ca (mg)</td>
<td>11.82 DW/2.74 FW</td>
</tr>
<tr>
<td>Fe (mg)</td>
<td>12.91 DW/2.99 FW</td>
</tr>
<tr>
<td>Mg (mg)</td>
<td>201.36 DW/46.59 FW</td>
</tr>
<tr>
<td>P (mg)</td>
<td>62.50 DW/14.46 FW</td>
</tr>
<tr>
<td>K (mg)</td>
<td>37.19 DW/8.60 FW</td>
</tr>
<tr>
<td>Na (mg)</td>
<td>2.11 DW/0.49 FW</td>
</tr>
<tr>
<td>Zn (mg)</td>
<td>0.05 DW/0.01 FW</td>
</tr>
<tr>
<td>Sulfur (mg)</td>
<td>14.48 DW/3.35 FW</td>
</tr>
<tr>
<td>Vitamin A (mg)</td>
<td>1.71 DW/0.4 FW</td>
</tr>
<tr>
<td>Vitamin C (mg)</td>
<td>23.38 DW/5.4 FW</td>
</tr>
<tr>
<td>Vitamin B1 (mg)</td>
<td>10.91 DW/2.52 FW</td>
</tr>
<tr>
<td>Vitamin B9 (μg)</td>
<td>8.13 DW/1.88 FW</td>
</tr>
<tr>
<td>Vitamin E (mg)</td>
<td>5.71 DW/1.3 FW</td>
</tr>
</tbody>
</table>

Note: The nutrients are expressed per 100 g.
Abbreviations: DW, dry weight basis; FW, fresh weight basis.
Source: Akubugwo et al. (2007).

TABLE 4  The estimated quantity of fresh ANS to meet the RDA

<table>
<thead>
<tr>
<th>Micro-nutrients</th>
<th>RDA (mg/day)</th>
<th>EQ of ANS per day (g)</th>
<th>Quantity of ANS per 100 g</th>
</tr>
</thead>
<tbody>
<tr>
<td>Thiamine</td>
<td>1 to 1.7</td>
<td>1375 to 2125</td>
<td>0.08 mg</td>
</tr>
<tr>
<td>Riboflavin</td>
<td>1 to 1.5</td>
<td>647 to 882</td>
<td>0.17 mg</td>
</tr>
<tr>
<td>Folate</td>
<td>400 μgDFE/day</td>
<td>1739</td>
<td>23 μgDFE</td>
</tr>
<tr>
<td>Vitamin C</td>
<td>73 to 90</td>
<td>209 to 257</td>
<td>35 mg</td>
</tr>
<tr>
<td>Vitamin E</td>
<td>15</td>
<td>154</td>
<td>9.7 mg</td>
</tr>
<tr>
<td>Vitamin A</td>
<td>700 to 900 μRAE/day</td>
<td>165 to 213</td>
<td>422 μRAE</td>
</tr>
<tr>
<td>Calcium</td>
<td>1000</td>
<td>503</td>
<td>199 mg</td>
</tr>
<tr>
<td>Iron</td>
<td>18</td>
<td>250</td>
<td>7.2 mg</td>
</tr>
<tr>
<td>Magnesium</td>
<td>400</td>
<td>435</td>
<td>92 mg</td>
</tr>
<tr>
<td>Phosphorous</td>
<td>1000</td>
<td>1333</td>
<td>75 mg</td>
</tr>
<tr>
<td>Potassium</td>
<td>3500</td>
<td>814</td>
<td>430 mg</td>
</tr>
<tr>
<td>Sodium</td>
<td>2400</td>
<td>3243</td>
<td>74 mg</td>
</tr>
<tr>
<td>Zinc</td>
<td>15</td>
<td>2500</td>
<td>0.6 mg</td>
</tr>
<tr>
<td>Copper</td>
<td>2</td>
<td>1333</td>
<td>0.15 mg</td>
</tr>
<tr>
<td>Manganese</td>
<td>2</td>
<td>95</td>
<td>2.1 mg</td>
</tr>
</tbody>
</table>

Note: EQ represents estimated amount of ANS in grams of fresh weight required to be eaten to meet the recommended daily allowance. RDA represents recommended daily allowance. μgDFE represents microgram of dietary folate equivalent. μRAE retinal activity equivalency. The vitamins and minerals are recommended for a person above 17 and 4 years, respectively. However, this quantity can vary depending on health status, sex, age, pregnant, and lactating women.
Source: http://www.fda.gov/nutritioneducation; FAO & WHO (2004); Zempleni et al. (2007); Van Jaarsveld et al. (2014); Ronoh et al. (2017).

Health maintenance and risk reduction of various diseases (Neugart et al., 2017; Oluoch et al., 2012). The high content of β-carotene in ANS contributes to vitamin A production, with health benefits in reproduction, vision, immune function, tissue differentiation, and embryonic development (Blake, 2008; Zempleni et al., 2007). Younger leaves of ANS have low β-carotene than matured leaves; however, matured leaves are rich in carotenoids (Cheptoo et al., 2019). Carotenoids help to reduce reactive oxygen species and prevent some types of cancers. However, they display a pro-oxidative effect under high concentration, high oxygen tension (lung of smokers), low levels of endogenous enzymes, and higher levels of metal ions (Fe³⁺ and Cu²⁺) (Park et al., 2013; Shin et al., 2020). Usually, β-carotene acts as a pro-oxidant at higher oxygen partial pressure in cells and thermally oxidized bulk oil systems (Ha et al., 2012; Park et al., 2013; Shin et al., 2020). Lutein and zeaxanthin are carotenoids pigments imparting yellow or orange color to various foods such as carrots, peppers, fish, and eggs (Abdel et al., 2013). Carotenoids protect age-related eye disease and filter specific wavelengths of light, thus providing the visual performance and offering photoreceptors protection from light damage (Abdel et al., 2013; Eggersdorfer & Wyss, 2018; Raman et al., 2019). Similarly, they reduced the risk of cataracts and early age-related macular degeneration (Eggersdorfer & Wyss, 2018; Raman et al., 2019). Lutein and zeaxanthin can inactivate free radicals and oxygen triplicates caused by light-induced cellular activity (Raman et al., 2019). Lutein-rich diets improved learning performance in mice and memory in old men and women (Eggersdorfer & Wyss, 2018) and also helped in developing the infant brain (Perrone et al., 2016). Still, there are no established dietary guidelines for lutein required to reach optimal macular pigment density in healthy people’s eyes (Eggersdorfer & Wyss, 2018; Raman et al., 2019; Ranard et al., 2017).

Phenolic compounds concentration in ANS is about 16,387-16,677 μg/g dry weight basis (DW) whereas flavonoids, phenolic acids, saponins, and tannins are widely occurring phenolic compounds in ANS. They act as antioxidative compounds by scavenging free radicals that delay or inhibit the initiation step (Amalraj & Pius, 2015; Degrain et al., 2020; Shahidi & Ambigaipalan, 2015; Yang & Wyss, 2018; Raman et al., 2019).
TABLE 5  Phytochemicals and antinutritional factors of the leaves of ANS species

<table>
<thead>
<tr>
<th>Compounds</th>
<th>S. nigrum</th>
<th>S. scabrum</th>
<th>S. villosum</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>TPP per 100 g of GAE FW</td>
<td>725 to 1307 mg</td>
<td>775 to 1247 mg</td>
<td>1026 mg</td>
<td>Yang and Ojiewo, 2013; Yuan et al., 2018</td>
</tr>
<tr>
<td>Quercetin glycosides</td>
<td>NI</td>
<td>1400 to 3300 μg/g DW</td>
<td>NI</td>
<td>Neugart et al., 2017</td>
</tr>
<tr>
<td>(quercetin-3-rutinoside)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Carotenoids</td>
<td>7.9 to 20.0 mg/100 g FW</td>
<td>10,100 mg/100 g DW</td>
<td>1790 μg/g DW</td>
<td>Adebooye et al., 2008; Odongo et al., 2018</td>
</tr>
<tr>
<td>Flavonoids</td>
<td>4.7 mg/100 g FW</td>
<td>25.5 mg/100 g FW</td>
<td>19.9 mg/100 g FW</td>
<td>Yang and Ojiewo, 2013</td>
</tr>
<tr>
<td>Glycoalkaloids</td>
<td>1722 mg/100 g DW</td>
<td>NI</td>
<td>1448 mg/100 g DW</td>
<td>Mohy-Ud-Din et al., 2010</td>
</tr>
<tr>
<td>Chlorophyll</td>
<td>69.8 to 155.8 mg/100 g FW</td>
<td>25,000 to 60,000 mg/100 g DW</td>
<td>19,600 mg/100 g DW</td>
<td>Adebooye et al., 2008; Odongo et al., 2018</td>
</tr>
<tr>
<td>Tannins</td>
<td>355.5 mg/100 g DW</td>
<td>NI</td>
<td>NI</td>
<td>Amalraj and Pius, 2015</td>
</tr>
<tr>
<td>Phytate</td>
<td>58.8 mg/100 g DW/7.64 mg/100 g FW</td>
<td>NI</td>
<td>0.04 to 0.2 mg/100 g DW/0.0052 mg/100 g FW</td>
<td>Amalraj and Pius, 2015; Mwanri et al., 2018</td>
</tr>
<tr>
<td>Oxalate</td>
<td>776.2 mg/100 g DW</td>
<td>33 mg/100 g DW</td>
<td>28.7 to 68 mg/100 g DW</td>
<td>Amalraj and Pius, 2015; Mwanri et al., 2018; Yang and Ojiewo, 2013</td>
</tr>
<tr>
<td>Cyanogenic glycosidase</td>
<td>320 mg/100 g FW</td>
<td>NI</td>
<td>NI</td>
<td>Essack et al., 2017</td>
</tr>
<tr>
<td>Nitrates</td>
<td>NI</td>
<td>NI</td>
<td>63.0 to 85.6 mg/100 g DW</td>
<td>Mwanri et al., 2018</td>
</tr>
</tbody>
</table>

Note: The total glycoalkaloids content was calculated by the sum of (Solasonine, α-Solamargine, β-Solamargine, and α-Solanine).
Abbreviations: DW, dry weight basis; FW, fresh weight basis; NI, not indicated; TPP, total polyphenol; GAE, gallic acid equivalent.

Phenolic compounds possess a wide range of physiological properties, mainly antiallergenic, antiatherogenic, anti-inflammatory, antimicrobial, antioxidant, antiinflammatory, cardioprotective, and vasodilatory (Boudet, 2007; Manach et al., 2005; Shahidi & Ambigaipalan, 2015). The average daily intake of dietary polyphenols is approximately 1 g per person (Shahidi & Ambigaipalan, 2015). The total polyphenol content of ANS ranges from 725 to 1307 mg/100 g FW (Table 5). This amount is sufficient to meet the RDA for the consumption of 100 g of FW ANS. However, high consumption of polyphenols causes low iron absorption (Zijp et al., 2000). Blanching of ANS at 95 °C or steaming using water or lemon juice solution significantly increases the total phenolic content (Yuan, Dinssa, et al., 2020).

Chlorophylls are natural pigments present in ANS. They act as a natural antioxidant with the ability to scavenge free radical and prevent several oxidative stress-related diseases such as cancer, neurological disorders, inflammatory diseases, dermatitis, tissue damage, sepsis, cardiovascular disorders, decreased immune function, and aging (Lanfer-Marquez et al., 2005; Mishra et al., 2011; Sangija & Wu, 2020; Wang & Wink, 2016). Chlorophyll can inhibit calcium oxalate dihydrate formation, which are the primary sources of kidney stones (İnanç, 2011). Besides, it stimulates the immune system, helps in detoxification, combats foul odors, and helps combat anemia and eliminate molds and toxins in the body (İnanç, 2011). Despite health benefits, chlorophyll can act as a pro-oxidant for oil oxidation when subjected to light (İnanç, 2011; Wang & Wink, 2016). Steaming and water blanching of ANS significantly increase chlorophyll content of 2.82 to 8.87 and 1.19 to 5.54 mg/100 g FW, respectively (Managa et al., 2020).

Glycoalkaloids are plant-derived bioactive compounds capable of interacting with living tissue components with a wide range of likely effects (Huang et al., 2016). ANS berries are a rich source of alkaloids. Solamargine and solasonine are glycoalkaloids in S. scabrum and solasodine glycosides in S. americanum. Solanum scabrum and S. villosum methanol leaves extract lack glycoalkaloids but are present in S. villosum Grif 16939 and S. nigrum PI 381290 accessions in a very low concentration of 50 μg/g DW or 1 mg/100 g FW (Yuan et al., 2018). Alkaloids have therapeutic effects such as cytotoxic against human carcinoma cells and anti-inflammatory against psoriasis (Al-Ashaal, 2019; Kumar et al., 2012). It prevented cervical carcinoma and showed schistosomicidal effect against S. mansoni, and fascicidal effect against Fasciola hepatica. Additionally, it has an inhibitory effect against HSV-1 (Al-Ashaal, 2019). Steroidal alkaloid solanine A from S.
scabrum demonstrated anti-inflammatory activity on Institute of Cancer Research mice (ICR) or albino mice and suppressed the production of nitric oxide in lipopolysaccharide/interferon γ-activated RAW264.7 cells (Zhao et al., 2018).

For a long time, phytate is considered an antinutrient, but recent studies have proven its antioxidant properties (Bhowmik et al., 2017; Mora-Boza et al., 2019; Silva & Bracarense, 2016; Wang & Guo, 2021). Phytate exhibits therapeutic properties on various diseases such as Alzheimer’s (Abe & Taniguichi, 2014), Parkinson (Xu et al., 2011), and management of blood glucose for type 2 diabetes (Lee et al., 2006). Similarly, phytate exhibits anticancer properties against the prostate (Raina et al., 2008), hepatocarcinoma (Al-Fatlawi et al., 2014), colorectal (Navarro et al., 2016), rhabdomyosarcoma (Vucenik, 1998), skin (Wawszczyk, 2015), and breast (Hussein et al., 2006) cancers. Besides, it acts as an antibacterial against Enterococcus faecalis (Nassar & Nassar, 2016), anti-HIV (Tateishi et al., 2017), and hypolipidemic (Dilworth et al., 2005). Also, it inhibits lipid peroxidation due to its high affinity to multivalent cations (Mora-Boza et al., 2019). Phytate is used in food industries as a molecular binder and functional ingredient, that is, it aggregates proteins and increases precipitation or turbid (Wang & Guo, 2021). The daily intake of 1–2 and 8–12 g of phytate possess a prevention effect on cancer and antitumor therapies (Vucenik & Shamsuddin, 2006). Further studies should explore the benefits of phytate, such as health and functional benefits, instead of addressing the general concept of phytate as an anti-nutrient.

Tannins are a group of phytochemicals (polyphenols) with an astringent taste and are present in various concentrations in vegetables and herbs (Amalraj & Pius, 2015; Khanbabae & van Ree, 2001). Tannins are water-soluble polyphenols present in many plant foods. Tannins have various health benefits such as antioxidants, cardioprotective, anti-inflammatory, antiviral, antibacterial, anticarcinogenic, antimutagenic, and antidiabetic (Chung, 1998; Delimont et al., 2017; Khanbabae & van Ree, 2001; Sharma et al., 2019). They also help heal wounds, cures dysentery, and help in hardening arteries (Sharma et al., 2019); nevertheless, it is an antinutrient.

5 | ANTINUTRITIONAL FACTORS OF ANS

ANS contains antinutritional factors such as oxalates, tannins, cyanogenic glycosidase, phytate, glycoalkaloids, and nitrates (Table 5) (Amalraj & Pius, 2015; Essack et al., 2017; Mwanri et al., 2018; Wakanhu et al., 2015). Some of them, for example, tannin, phytate, and glycoalkaloids, exhibit functional properties, for example, anticancer, antibacterial, antiviral, and anti-inflammatory properties (Delimont et al., 2017; Silva & Bracarense, 2016), also elaborated in Section 4.2. The oxalate and phytate contents vary with the maturity stage in S. villosum (Silva & Bracarense, 2016). Solanum villosum (Nduruma BG 16 and Olevolosi SS 49) had the highest oxalate and phytates content, with a nonsignificant decrease in nitrate content at 35 days (Mwanri et al., 2018). Most of the Solanaceae family species are poisonous to humans and livestock (Jain et al., 2011). For instance, the deadly nightshade (Atropa belladonna L.) contains tropane alkaloids. Solanine glycoalkaloids in S. nigrum, S. villosum, S. americanum, and S. scabrum significantly cause toxicity (Jain et al., 2011). Notably, various ANS processing methods such as boiling, drying, and fermentation effectively remove antinutritional factors (Essack et al., 2017).

5.1 | Phytate in leaves

Phytate is an antinutrient when present in higher concentrations (Bhowmik et al., 2017). Phytate can chelate with divalent/trivalent metal ions such as zinc, copper, calcium, and iron and reduce bioavailability (Silva & Bracarense, 2016). Solanum nigrum contains the highest phytate content with the lowest in S. villosum (Table 5). The calculated molar ratio of phytate:iron was 0.09 in S. nigrum. An increase in ANS leaves’ phytate content from 0.04 mg/100 g DW in days 21 grown to 0.3 mg/100 g DW in 35 days was reported (Mwanri et al., 2018). Therefore, harvesting at the right maturity is crucial for ANS nutritional quality. The molar ratio above 0.4 has a significant effect on iron bioavailability (Dahdouh et al., 2019; FAO/IZInCG, 2018). Therefore, the molar ratio was meager to cause the chelating effect of iron in ANS. The phytate intakes reported in the United Kingdom were 692 to 948 mg/day in men and 538 to 807 mg/day for women. The daily phytate requirement for vegetarians is 1600 to 2500 mg/day (European Food Safety Authority [EFSA], 2014). Fermentation, germination, soaking, malting, boiling, solar, and pressure cooking are simple, inexpensive, and convenient techniques for the removal of phytates in ANS (Abdulwaliyu et al., 2019; Essack et al., 2017; Owade et al., 2019; Pasrija & Punia, 2010; Rasane et al., 2015; Wang & Guo, 2021). Water blanching at 95°C and steam blanching of ANS for 5 min reduce phytate concentration by 25% to 75% (Managa et al., 2020). Besides, enzyme degradation using phytase is the most effective and applicable way to remove phytate (Wang & Guo, 2021). Phytate in unprocessed foods does not reflect the actual quantity consumed; therefore, more emphasis should be
placed on assessing phytate in ready-to-eat foods rather than its content in raw forms (Abdulwaliyu et al., 2019).

5.2 Oxalate in leaves

As an antinutrient, oxalate chelates minerals such as potassium, sodium, and calcium, forming insoluble complexes, thus hindering their absorption. Insoluble calcium oxalate can cause joint pains, kidney stones, and kidney failure (Holmes & Kennedy, 2000; Judprasong et al., 2006; Soto-Blanco et al., 2009). The oxalate content of ANS can go up to 776.2 mg/100 g DM (Table 5); however, the fatal doses range from 1 to 30 g per person (Dassanayake & Gnanathasan, 2012; EMEA, 2004). The oxalate content in ANS ranges from 28.7 to 776.2 mg/100 g DW or 3.7 to 100.9 mg/100 g FW. Solanum nigrum contains the highest oxalate content and S. scabrum the lowest (Table 5). This amount is lower than the fatal dose, but it is still a concern regarding the risk of toxicity; therefore, monitoring ANS leaves’ oxalate levels is necessary. Nonetheless, the amount of oxalate in ANS leaves is significantly lower than in some exotic vegetables (Akhtar et al., 2011; Faudon & Savage, 2014). Fermentation and boiling reduce oxalate content in ANS and other AIVs (Essack et al., 2017; Muchoki et al., 2010; Owade et al., 2019; Wakhanu et al., 2015). Likewise, blanching at 95 °C and steam blanching for 5 min reduce oxalate in ANS by 42% to 75% (Managa et al., 2020). Moreover, the early harvesting stages (21 days) show lower oxalate content (42.9 mg/100 g DW) than the late stages of 35 days (60.9 mg/100 g DW) (Mwanri et al., 2018). Therefore, proper harvesting times and selection of ANS varieties should be the criteria for obtaining leaves with low oxalate content.

5.3 Tannins in leaves

Tannin, either nonhydrolyzable (condensed) or hydrolyzable (Sharma et al., 2019), forms a complex with proteins, digestive enzymes, starches, and minerals, thus reducing food’s nutritional value (Chung, 1998; Polycarp et al., 2012). Tannin is responsible for decreasing feed intake, feed efficiency, protein digestibility, and net metabolic energy. It also increases the excretion of protein and essential amino acids, damages the mucosa lining of the gastrointestinal tract, and alters cations’ excretion (Chung, 1998). Tannins are toxic when precipitating with heavy metals and alkaloids (Khanbabaee & van Ree, 2001; Sangijja & Wu, 2020). Tannin causes a browning reaction (darkening of food) due to polyphenol oxidase (Chung, 1998). Tannin at a concentration of 0.13 to 1 g/kg body weight (BW) decreases erythrocyte counts in pigs’ hemoglobin and hematocrit (Lee et al., 2010). Tannin content above 3 g/100 g caused mortality in test chicks (Chung, 1998). Tannin also causes liver, skin, oesophageal, stomach, lung, kidney, and nasal cancers in humans (Chung, 1998). The RDA of proanthocyanidin (tannin) is 53.6 to 450 mg/person/day and 1250 mg/person/day for hydrolyzable tannins in the Spanish population. Nevertheless, the total tannin content of S. nigrum of 360.1 mg/100 g DW or 46.81 mg/100 g FW is relatively low to harm the consumers. Therefore, consumption of ANS is safe with potential health effects. In case of sufficient tannin content to cause potential health problems, cooking is an effective removal method because it is heat-labile and facilitates its degradation (Essack et al., 2017; Kakati et al., 2010; Owade et al., 2019; Serrano et al., 2009). Further, lactic acid fermentation, drying, canning, boiling, soaking in water, and freezing can also remove tannins (Essack et al., 2017; Serrano et al., 2009). Managa et al. (2020) reported a decrease in tannin content by 7% to 14% through blanching at 95 °C and steam blanching for 5 min.

5.4 Nitrates/nitrites in leaves

Fruits and vegetables are the significant nitrate/nitrite sources and contribute 50% to 85% of overall dietary intake (EFSA, 2008; Nuñez et al., 2015). Several fruits and vegetables contain 200 to 2500 mg of nitrate per kilogram (WHO, 2003). Nitrates/nitrites are easily absorbed in the body; about 60% to 70% are excreted in the urine, whereas 3% appear in the urine as urea and ammonia (Karwowska & Kononiuk, 2020). In the stomach, blood, and tissue, nitrites are converted into bioactive reactive nitrogen oxide species (NO). The reactive NO contributes to the formation of carcinogenic nitrosamines of toxicological importance (Ding et al., 2018). Nitrates can contribute to carcinogenic, such as breast cancer, gastric cancer, renal cell carcinoma, adult glioma, colorectal cancer, esophageal cancer, and thyroid cancer (Karwowska & Kononiuk, 2020; Keszei et al., 2012; Yang et al., 2017). Besides, it contributes to genotoxicity, cytotoxicity, inhibition of enzymatic reactions and proteolysis, and altered immunogenicity (D’Ischia et al., 2011). Antioxidants, such as vitamins C and E, inhibit nitrosamines’ generation (Ding et al., 2018; Karwowska & Kononiuk, 2020). Mwanri et al. (2018) reported the nitrate content of 630 to 856 mg/kg DW in S. villosum; however, there is limited information on leaves of other ANS species. Some exotic vegetables such as spinach, rucola, celery, rhubarb, lettuce, beets, chard, and beetroot contain significantly higher nitrates than ANS (Karwowska & Kononiuk, 2020). Seasonality and the cultivation systems contribute to nitrites variation in ANS. Late harvested Nduruma BG 16 and Olevolosi SS
49 showed lower nitrate content than early harvested ones (Mwanri et al., 2018). Similarly, the nitrate content was higher in 21-day harvested leaves (856 mg/kg DW/111.3 mg/kg FW and significantly lower at day 35 (754 mg/kg DW/98 mg/kg FW) (Mwanri et al., 2018). Therefore, farmers should consider late harvest for lower nitrate content in ANS. Human generally consumes between 1.2 and 3.0 mg of nitrate daily (WHO, 2016). The RDA of nitrite and nitrates is 0.06 to 0.07 mg/kg BW/day and 7 mg/kg BW/day, respectively (Karwowska & Kononiuik, 2020).

Therefore, the nitrate content in ANS leaves is low comparing to the RDA (Table 5). Besides, the consumption of more than 600 to 700 g of ANS per day is beyond the RDA of nitrate and can cause health effects. The application of heat treatments, high-temperature storage conditions, and fermentation reduced nitrate content in vegetables (Ding et al., 2018; Prasad & Chetty, 2011), similarly to ANS leaves.

### 5.5 Glycoalkaloids in leaves

The solasonine, solanine, solamargine, and chaconine are major glycoalkaloids in the ANS, with steroidal glycoalkaloids (SGAs) as minor (Ronoh et al., 2017). Glycoalkaloids are toxic to humans and animals with symptoms such as constipation, dark-colored diarrhea, nausea, vomiting, and abdominal pain. These toxins affect the nervous system to cause drowsiness, apathy, weakness or paralysis, salivation, circulatory and respiratory depression, and unconsciousness. Toxic effects are primarily irritation of the digestive tract and sometimes neurological problems. (Abbas et al., 1998; Defelice, 2003; Mensinga et al., 2005).

Young leaves and unripe berries of ANS have a higher SGA concentration than matured ones (Ronoh et al., 2017). The SGA is associated with bitterness in ANS and causes toxic effects to animals when consumed above 5 mg/kg BW (Ronoh et al., 2017). The recommended upper limit of total glycoalkaloids (TGA) in plant foods is 200 mg/kg FW (1 g/kg DW) (Nono-Womdim et al., 2012). Alkaloid content in *S. nigrum* and *S. villosum* is 1722 and 1448 mg/100 g DW, respectively (Table 5). The quantity of solanine in *S. nigrum* and *S. villosum* is 470 and 150 mg/100 g DW, respectively (Mohy-Ud-Din et al., 2010). This content is significantly low to cause a potential toxic health effect on humans. Also, Yuan et al. (2018) reported a solasodine content of less than 5 mg/100 g DW and 1 mg/100 g FW in *S. nigrum* and *S. scabrum*, respectively. This amount is low compared to other vegetables such as eggplant (6.25 to 20.5 mg/100 g FW) (Yuan et al., 2018). The SGA solasodine is absent in *S. scabrum* (Yuan et al., 2018; Yuan, Dinssa, et al., 2020). According to Yuan, Lyu, et al. (2020), the levels of total alkaloids in *S. nigrum, S. villosum*, and *S. scabrum* are safe for human consumption. Alkaloids content in ANS leaves can be removed by boiling (Defelice, 2003; Edmonds & Chweya, 1997; Essack et al., 2017). The TGA content in *S. scabrum, S. villosum*, and *S. tarteremotum* leaves is 116.80, 100.82, and 112.97 mg/kg FW, respectively, with no maximum toxic levels of SGAs (Mwai, 2007). It is noteworthy that several factors such as the amount of TGA eaten, BW, and metabolism rate of SGA facilitate its toxicosis (Nono-Womdim et al., 2012).

Breeding and genetic improvement would also remove the toxicity commonly associated with ANS, that is, mainly attributed to SGA presence (Nono-Womdim et al., 2012). Although SGAs are harmless and enhance the flavor at low levels, they cause toxicity and even death in animals and humans at a high dose. TGA content above 140 mg/kg FW is associated with unpleasant flavor and bitter taste (Nono-Womdim et al., 2012). In Tanzania and Kenya, several improved cultivars of *S. scabrum* and *S. villosum* are available (https://www.tosci.go.tz/; Dinssa et al., 2016; Ojiewo, Mbwambo, et al., 2013; Ronoh et al., 2019). *Solanum scabrum* has a less bitter taste with large succulent and broad leaves (Figure 1), high seed yield, rapid new leaf sprouting after harvest, late flowering, and 40 days’ maturity, and picking can continue for 6 to 8 weeks. Besides, it is resistant to *Fusarium* wilt (Nono-Womdim et al., 2012; Ojiewo, Mbwambo, et al., 2013; https://www.tosci.go.tz/). *Solanum villosum* is bitter, with a narrow and low yield than *S. scabrum*; also, it has broad leaves and high yield than the original variety (Figure 1). People in rural areas, old peoples, and men prefer bitter variety; therefore, all varieties fetch the same demand in the market. Nduruma and Olevolosi leaves contain no glycoalkaloid aglycone; therefore, they are safe for consumption. Moreover, these varieties have high total phenols and high vitamin E and β-carotene (Yuan et al., 2018).

### 5.6 Glycoalkaloids in berries

ANS berries are commonly considered toxic and not consumed in SSA (Lyu et al., 2020). The young berries of *S. nigrum* contain the highest level of SGA compared with the whole plant or leafy and stem parts (Abbas et al., 1998; Defelice, 2003; Edmonds & Chweya, 1997; Mohy-Ud-Din et al., 2010; Ronoh et al., 2017; Yang & Ojiewo, 2013). The toxic level of matured berries is too low to harm but can harm children (Defelice, 2003; Edmonds & Chweya, 1997). *Solanum nigrum* accession (USDA PI 381239) contains trace levels of glycoalkaloids (Lyu et al., 2020). However, the glycoalkaloids (solasodine, solanine, and solamargine) are high in berries compared to shoots and leaves of *S. nigrum*, with solasonine being high in the shoots of *S. nigrum* (Al-Ashaal, 2019). Glycoalkaloids in unripe berries
of *S. nigrum* are equivalent to 356 mg/100 g DW. Environmental conditions such as frost contribute to a significant increase in glycoalkaloids by five to 15 times higher than mature berries. Yuan, Lyu, et al. (2020) reported a solasodine content of 380 to 930 mg total aglycone/100 g DW in unripe berries but was not detected in ANS leaves. This variation could be attributed to genetic diversity, environmental conditions, and cultural practices (Ojiewo, Mwai, et al., 2013). Glycoalkaloid concentration is low in mature berries. *Solanum scabrum* mature berries contain up to 1500 mg of glycoalkaloids (Yuan et al., 2019). Such high concentration of glycoalkaloids in berries could be essential for medicinal purposes (Al-Ashaal, 2019). Mature berries of some accessions contain low amounts of glycoalkaloids and could be potential for consumption in SSA. For such, the promotion of berry products for consumption is essential. Therefore, intensive research in breeding for selecting desirable lines and subsequent cultivars for release ensuring the quality of the final product and safety to consumers is mandatory (Yuan, Dinssa, et al., 2020; Yuan, Lyu, et al., 2020).

### 5.7 Cyanogenic glycosidase in leaves

When hydrolyzed, cyanogenic glycosidase produces the toxic hydrogen cyanide (HCN) (Selmar, 2018). Only *S. nigrum* contains cyanogenic glycosidase (Table 5), which is of safety concern. HCN inhibits cytochrome oxidase in the mitochondria. Also, it interacts with copper and iron ions to inhibit respiration and the inability to produce adenosine triphosphate (Sangija & Wu, 2020). Cyanide intoxication can cause mental confusion, diarrhea, vomiting, stomach pains, dizziness, diaphoresis, cardiac arrest, and rapid respiration (Gracia & Shepherd, 2004; Venketesh, 2014). A chronic low dose of cyanide may cause an elevation in the blood and induces a variety of symptoms such as kidney or mild liver damage (Manzano et al., 2007), miscarriage (Soto-Blanco et al., 2009), hypothyroidism (Soto-Blanco et al., 2009), paralysis, nervous lesions, and weakness (Soto-Blanco et al., 2002, 2008). The fatal dose of HCN can be as low as 0.5 to 3.5 mg/kg/BW (EFSA, 2019; Essack et al., 2017). In contrast, the cyanide content of *S. nigrum* is 320 mg/100 g FW (Essack et al., 2017); moreover, no information is available for other ANS species. HCN content in ANS is significantly low compared to that in raw cassava leaves (1905 mg/kg) (Umuhozariho et al., 2014). FAO (2005) suggested an acceptable limit of 500 mg/kg HCN, whereas the EFSA (2004) considers the maximum of 10 mg/kg HCN to be safe for human consumption. Fresh ANS contains low HCN content to cause potential health effects as per (FAO, 2005). Cassava leaves’ cooking reduces HCN similar to ANS (Umuhozariho et al., 2014).

### 6 SAFETY REGULATIONS FOR ANS

ANS safety is still a challenge, with a risk of toxicity to consumers if consumed at a high dose. Some ANS leaves accessions contain glycoalkaloids, potentially causing health concerns (Sivakumar et al., 2020). To date, there are no policy issues, regulations, or bay laws guiding the safety of consuming ANS in SSA or other parts of the world (Nondi, Womdim et al., 2012). Therefore, the SSA countries need to develop and implement policies and strategies to promote sustainable production and ANS safety (Abukutsa-Onyango, 2010). There is a need for policy-makers at the national, regional, and international levels to address the significant safety issues on ANS to attain maximum benefits. The EFSA, FAO, and WHO have set guidelines for minimal levels of toxic compounds such as GA, HCN, nitrate, and nitrates in foods. Therefore, these guidelines can be used as a benchmark in preparing safety regulations for ANS in SSA. Further, intensive research is needed to generate data on the toxic compounds and their levels in *Solanum* spp.

### 7 THE UTILIZATION OF ANS

#### 7.1 Leaves

In some societies, the consumption of ANS is influenced by the consumer’s local beliefs. For instance, most people in Kenya consume ANS believing that it is nutritious and healthy (Abukutsa-Onyango, 2015). ANS uses differ in SSA; however, it is used for leafy vegetables, medicinal, cosmetic, and spiritual purposes (Abukutsa-Onyango, 2007; Keller, 2004; Ontita et al., 2017; Yang & Ojiewo, 2013). Generally, ANS consumption is based on availability, taste, and affordability (Sato et al., 2002).

Consumption of ANS in some African societies is associated with hunger, shortage of food, and poverty; therefore, eating them is regarded as humiliating (Shackleton et al., 2009). Abukutsa-Onyango (2015) observed that most Western and Nyanza provinces in Kenya prefer ANS for its nutritional value. In Tanzania, moreover, ANS is highly consumed by the wealthiest households for their known nutritional and health benefits (Ambrose-Oji, 2009). ANS is generally said to be more palatable for adults, especially the old age group (Ontita et al., 2017; Onyango et al., 2016). Nevertheless, some parts of ANS are toxic to humans and livestock, including youngberries, which contain a high
concentration of alkaloids (Kuete et al., 2017; Ronoh et al., 2017).

In Kenya, ANS is used spiritually by older men to pronounce a curse on an offender (Ontita et al., 2017). ANS are consumed based on gender, whereby men prefer bitter dark variety. Some consume to recover the lost or poor appetite due to illness (Ontita et al., 2017; Shackleton et al., 2009). However, wrong beliefs, perishability, environmental degradation, limited storage, poor infrastructure, draughts, markets, arable land shortage, diseases, pests, and high production costs are factors hindering ANS utilization (Ayua & Omware, 2013; Ondieki et al., 2011). Henceforth, research on how to address the constraints to the utilization of ANS is mandatory.

7.2 | Berries/fruits

Green, orange, yellow, purple, red, black, and violet berries are edible; they can be consumed raw, at the same time, the juice of ripe berries is used as ink by children (Edmonds & Chweya, 1997). The berries are used as an ingredient in pies, food colorants, preservatives, and substitute for raisins in plum puddings (Edmonds & Chweya, 1997; Lotter et al., 2014; Schippers, 2002; Yang & Ojiewo, 2013). Berries can sometimes be mixed with colorful fruits such as apples. Also, berries can make a nice jam and preserve tea with bread and butter (Edmonds & Chweya, 1997; Yang & Ojiewo, 2013). Occasionally, the berries are used as a vegetable in soup, yam, and cocoyam porridges in some parts of Nigeria (Akubugwo et al., 2007). The berries are processed into value-added products, such as beverages, juices, jams, and nectars for international markets (Akinola et al., 2020).

8 | ROLES OF ANS IN HEALTH PROMOTION AND PROTECTION

8.1 | Leaves

Different ANS species and parts have various medicinal functions (Yang & Ojiewo, 2013). The leaf extracts of S. americanum relieve chronic conjunctivitis, treat sores, heart pain, skin problems, and treat worms in chicken. Simultaneously, the leaf extracts of S. scabrum treat diarrhea in children and cure jaundice and eye infections. Also, the leaf extracts of S. villosum treat stomach aches, tonsillitis, wounds, leucorrhoea, nappy rash, as well as an ointment for boils (Zahara et al., 2019). Fresh leaves or cooked or leaf juice can also be taken orally as a liver tonic or used to treat indigestion, stomach ache, and stomach ulcers (Jain et al., 2011). In Kenya, ANS is used to heal ailments relating to stomach complications (Abukutsa-Onyango, 2015). Leaves are eaten raw for heart pain and swellings; also, the leaf extract restores body skin pigment. Fermented leaves treat boils, ulcers, and swollen glands. ANS leaves also treat tonsillitis, swelling, and conjunctivitis. The Xhosa also use ANS for disinfecting anthrax-infected meat (Edmonds & Chweya, 1997).

In European traditional medicine (TM), nightshades are potent analgesic, sudorific, and sedative, containing powerful narcotic properties. In Indian tradition, the nightshades have been used as medicines to cure stomach complaints, dysentery, fever, and tuberculosis (Kaushik et al., 2009; Kuete et al., 2017; Lotter et al., 2014). TM is “the total of knowledge, skills, and practices based on the theories, beliefs, and experiences indigenous to different cultures that are used to maintain health, as well as to prevent, diagnose, improve, or treat physical and mental illnesses” (Che et al., 2017). TM has gained popularity in the last few decades and continued to be useful for the poor’s primary health care in SSA (Lezotre, 2014; Sivakrishnan, 2018).

The juice of the ANS leaves cures ulcers and other skin diseases. The BNS is used broadly as antitumorigenic, antioxidant, anti-inflammatory, hepatoprotective, diuretic, and antipyretic agent (Jain et al., 2011; Kuete et al., 2017). The S. nigrum leaves are used to treat fungal skin infections, reduce stress, relieve joint pain, treat malaria, and recover lost food appetite (Jain et al., 2011; Kuete et al., 2017; Ontita et al., 2017). The bitter dark green leaves are used for an alcohol-related hangover (Jain et al., 2011; Ontita et al., 2017). Also, ANS can treat the new circumcised patient and pregnant and lactating mothers (Ontita et al., 2017). Leaf paste is applied directly to rabies for healing wounds (Jain et al., 2011). Aqueous extracts of leaves from both S. villosum and S. nigrum var. sarrachoides possessed significant antidiabetic effects in the Streptozotocin-induced diabetes mice model (Nyaga et al., 2019; Nyaga, 2020).

8.2 | Berries and roots

The ripe berries of edible ANS are anticarcinogenic; other plant parts are also used as TM (Kuete et al., 2017). The raw berries of S. scabrum treat stomach aches and stomach ulcers. They are also used as a tonic, laxative, remedial for asthma, and restoring appetite, especially for people who recover from disease (Jain et al., 2011). Unripe berries relieve aching teeth and reduce the pain of babies’ gum during teething. The juice of berries treats sore eyes. Solanum nigrum berries and juices cure stomach ailments, blood impurities, and fevers. In East Africa, the raw berries treat stomach ulcers or general abdominal upsets (Edmonds & Chweya, 1997; Yang & Ojiewo, 2013). A mixture of ground ANS leaves and seeds is scrubbed onto the
gums of children with crooked teeth. In Tanzania, unripe berries treat ringworms and roots for stomach ache. The Zulus use an infusion as an enema for abdominal upsets in children. The burnt and powdered root is rubbed to scarification on the back for the relief of lumbago. Unripe berries paste is used to treat ringworm (Edmonds & Chweya, 1997; Yang & Ojiewo, 2013). The berries have been used as ancient Indian medicine to treat inflammation, diuretics, and tuberculosis (Kaushik et al., 2009). In Tanzania, the ripe berries in the edible form are given to kids to stop bed-wetting (Jain et al., 2011). However, in Algeria, berries’ diluted infusion treats blindness, conjunctivitis, glaucoma, trachoma, and cataract (Jain et al., 2011). The berries juice calms sore eyes, whereby the unripe berries soothe toothache (Yang & Ojiewo, 2013).

In contrast, the juice from roots treats cough and asthma. In many African cultures, various plant roots are used in traditional, complementary, and alternative medicine (Akinola et al., 2020). Berries and roots can control vomiting and treat tetanus after abortion and can also be used as sedatives (Ogwu et al., 2016). In India, consuming boiled roots with a little sugar increases fertility in women, whereas the juice of roots is extracted and used to cure asthma and whooping cough (Jain et al., 2011). Children take the tonic obtained after boiling roots in milk.

9 TRADITIONAL METHODS FOR PREPARATION OF ANS

Processing and preservation (PaP) of ANS can compensate for the low availability of vegetables during the off-season, hence improving the nutritional shortage during these seasons (Engle & Altoveros, 1999). Most of the ANSs are nutritious if well cooked, although the preparations and cooking methods significantly affect the nutritional contents (Keller, 2004; Ngegba, 2007; Oluoch et al., 2012). Therefore, it is necessary to develop recipes for optimizing nutritional value, increasing ANS intake, and reducing iron deficiency in children, pregnant women, children, and nursing mothers in SSA (Oluoch et al., 2012). A correlation between taste and appearance was observed in Kenya. Singly prepared ANS or combined with other vegetables did not influence sensory acceptability, although a mixture of ANS and cowpea increased carotenoids and vitamin C (Habwe, 2008; Habwe et al., 2008; Oluoch et al., 2012). Stir-frying can be the best method of retaining vitamin C, copper, and iron, although mixing slender leaves in ANS reduce iron content (Habwe et al., 2008). In Tanzania, different recipes for the preparation of ANS exist, making use of variable quantities of ingredients; hence, it is difficult to obtain a standard recipe to be adapted (Weinberger & Msuya, 2004). Traditional recipes for the preparation of ANS vary among different communities in SSA (Figure 2).

In Tanzania, ANS is cooked by stir-frying or steaming, with ingredients such as milk, coconut milk, peanut, carrots, sweet pepper, onion, tomatoes, and peppers (Keller, 2004; Ngegba, 2007). Similarly, ANS can be prepared by boiling and adding milk or cream for enrichment or boiling and frying with onions, tomatoes, spices, and salt (Figure 2). In Kenya, most local consumers boil the leaves and add salt and milk to improve taste (Ontita et al., 2017; Yang & Ojiewo, 2013). Furthermore, ANS can be mashed with potatoes or boiled with sweet potatoes, tuber, milk, pumpkin, and blood or immature gourd (Ontita et al., 2017). In Senegal, ANS is cooked with meat or fish (Figure 2) (Chagomoka et al., 2014; Keller, 2004; Shackleton et al., 2009). Besides, cooking of ANS with meat can increase cooking time and facilitate the degradation of vitamins B and C, β-carotene, and iron. Diabetic consumers prepare ANS by boiling and adding milk, but neither fry nor add salt for health reasons (Ontita et al., 2017).

Traditional cooking methods can retain carotenoids in a range between 16% and 70%, whereas long cooking and reheating of cooked ANS reduce carotenoids and iron (Oluoch et al., 2012). Various cooking methods can degrade some essential micronutrients (Table 2), such as iron, β-carotene, and vitamin C (Ngegba, 2007). In East Africa, ANS can be cooked with other vegetables such as spinach, kale, amaranth, and cabbages (Figure 2) and accompanied with staples such as stiff porridge (ugali), rice, yams, banana, and cassava (Abukutsa-Onyango, 2007; Schippers, 2002; Yang & Ojiewo, 2013).

Most studies have focused on ANS production aspects and forget different cooking methods, consequently altering the nutritional and functional properties. The absence of suitable recipes and information on proper use led to the low utilization of ANS and distortion of essential nutrients. Formulation of appropriate ANS recipes can maximize nutrient availability, eliminate seasonality, and guarantee their availability yearly. Additionally, it can reduce losses and improve dietary diversification, contributing to nutrition and food security.

10 ANS VALUE CHAIN

ANS is common in rural and urban markets in Africa, especially in Cameroon, Kenya, Ghana, Burkina Fasso, Madagascar, Nigeria, South Africa and Tanzania, Guatemala, New Guinea, and the Mediterranean (Edmonds & Chweya, 1997; Essack et al., 2017; Ojiewo, Mbwambo, et al., 2013). ANS is highly demanded and expensive than many vegetables, such as kale or spinach (Onyango et al., 2016). The AIVs value chains play a significant role in food...
security and poverty reduction. ANS supports a large number of microbusinesses along the supply chain (Ojewo, Mbwambo, et al., 2013). Nevertheless, it is yet to penetrate the international market (Edmons & Chweya, 1997). ANS faces typically stiff competition with other exotic vegetables such as cabbage, spinach, amaranths, kale, and lettuce (Abukutsa-Onyango, 2007); moreover, lack of formal reports on its value chain is the bottleneck. According to Gogo et al. (2017), the standard chain level of AIVs in SSA includes harvesting, transportation, and marketing. However, the lack of PaP in the chain limits AIVs utilization, ANS inclusive. Therefore, mapping the ANS value chain is vital to create well-functioning linkages between different actors for mutual benefits. In Eldoret, Kenya, consumers prefer to purchase ANS over exotic vegetables; however, the ANS price varies between towns. In the open market, the price for a bunch of ANS is Kenyan shilling (KSH) 18.8, whereas in the supermarket, the average price is KSH 34 (Chelang’a et al., 2013). In Tanzania, the wholesale price for an AIVs bunch of 40 to 50 kg is 20,000 Tanzanian shillings (TZS) (US$12.50) and can even fetch a low price of 10,000 TZS (US$6.25) during the bumper season (Lotter et al., 2014). The price of selling 1 kg of ANS is between 517–1065 TZS (Everaarts et al., 2017; Molina et al., 2020). The prices of 1 kg of ANS vary in different seasons and stages of the value chain: farmgate (400 to 1350 TZS), wholesalers (2200 TZS), and retailers (2400 TZS) (Molina et al., 2020). During the rainy season, a bed of AIVs of the same size can be sold at a wholesale price of up to 100,000 TZS (US$62.50) as rain disrupts production (Lotter et al., 2014). Generally, the value chain and distribution channels for ANS are not well established in SSA. A value chain analysis in Kenya earmarked supermarkets and local markets as key distribution channels for ANS. Smallholder farmers can directly supply ANS to retailers or wholesalers, and supermarkets with no specific entry point (Figure 3). Besides, the supply of AIVs is from the small farmers to different outlets (Figure 3). Lack of appropriate infrastructure, market information, extension services, acceptable products, and storage facilities hinders the proper distribution of AIVs (Senyolo et al., 2018; Temu & Temu, 2005).

In Tanzania, the ANS value chain is an informal sector where its marketing involves small-scale producers, retailers, and whole sellers. PHL is low in the market with a robust sales system, which keeps in touch producers, buyers, and retailers with their customers daily (Lotter et al., 2014; Shackleton et al., 2009). The use of mobile phones facilitates the selling of ANS because producers can keep in touch with the middlemen, retailers, and wholesalers (Edmons & Chweya, 1997). Most Tanzania retailers sell on an average of 1.5 kg of ANS, with 62% of all retailers store unsold produce for the next day (Lotter et al., 2014). Most Tanzania retailers sell on an average of 1.5 kg of ANS, with 62% of all retailers store unsold produce for the next day (Lotter et al., 2014). Men can sell 3 kg per day more than women, despite female dominance (96%) in ANS marketing. The sales volume has a significant effect on location, with an average of 2 kg per day in Dodoma and more than 5 kg in Arusha (Lotter et al., 2014).
Nightshade for Improving Human Nutrition...  

Figure 3: The traditional AIVs value chain in SSA. Dotted arrows indicate occasional pathway. Black arrows indicate the common pathway. Sources: Acedo and Katinka (2009); Sangija et al. (Unpublished).

Furthermore, sellers at district and ward markets sell 2 kg more than a seller at a street stall (Lotter et al., 2014).

Over 60% of farmers in Kenya and Tanzania were able to market the ANS and penetrate several markets. The majority of farmers in East Africa have changed their perception of production ANS for family use and shifted to business production to support their families and improve livelihood. Commercializing of ANS has been considered a profitable business by most farmers because it has many benefits, including the low use cost, less infestation by pests and diseases, high maturity, high consumer demand, and market opportunities (Muhangi et al., 2011). Economic returns from the use of ANS as vegetables have not yet been quantified, hence the low market price and a low economic value (Edmonds & Chweya, 1997).

On the other hand, the annual mean profit obtained by small-scale farmers from selling ANS in Kenya ranges from 5641 to 141,144 KSH (US$564.12 to US$1411.44) (Onyango et al., 2016). The profitability of ANS in Kenya is estimated to be US$754/month and US$766/ha (Mwangi & Kimathi, 2006). Production of ANS gives high yields and low fertilizer application than other exotic vegetables (Everaarts et al., 2017). The annual profit based on producing 1 ha of ANS in Tanzania was 8,129,280 TZS/ha/year (US$3505.51) (Everaarts et al., 2017). However, Molina et al. (2020) reported an annual net value of 4,612,042.25 TZS (US$2420.02), higher than exotic vegetables, Chinese cabbage, and spinach. The relatively high ANS price maintained yearly around in all markets, traders, and farmers signifies its preference and demand (Molina et al., 2020).

This further shows the potential of ANS in improving the livelihood of people in SSA; therefore, exploiting its value chain is necessary to maximize the benefits of all actors.

11 | Postharvest Handling

ANS needs proper handling after harvesting to maintain the quality before processing or being sent to the market (Arah et al., 2016). Harvesting is recommended in the morning with no dew or late in the evening (Abukutsa-Onyango, 2015). Because ANS is highly perishable, the chance of spoilage increases rapidly with time, mainly when poorly handled (Onyango & Imungi, 2007). Therefore, cooling is mandatory to remove field heat within a short period. The traditional cooling methods include putting the vegetable under the shade, sprinkling with water, and dipping in water (Figure 4) (Gogo et al., 2016). Water sprinkling has proven effective in quickly removing field heat and maintaining freshness for a longer time (Arah et al., 2016; Gogo et al., 2016). The chance of ANS deterioration increases within 24 hr if not immediately sold after harvest. Some farmers/sellers sprinkle water and leave the vegetable in the open space overnight; however, the problem of microbial contamination still hinders their effort (Gogo et al., 2016). Excessive field heat increases metabolic activity, favors microbial activity, and increases respiration rate and ethylene production (Akbudak et al., 2012).

After harvest, cleaning, sorting, de-stalking, grading, and packaging of ANS are performed. Cleaning of ANS is necessary to remove any dirt or residue particles and microbial contamination agents (Nyaura et al., 2014). However, most farmers forget to dry leaves after washing and consequently encounter a higher prevalence of microbial contamination, mainly fungi and bacteria (Arah et al., 2016; Gogo et al., 2016). Lotter et al. (2014) observed that 52% of growers do not wash the harvested ANS, increasing the chances of microbial infection and deterioration. To extend its shelf life, ANS is kept on cold shelves in the supermarkets at about 5 to 10 °C (Abukutsa-Onyango, 2007; Gogo et al., 2016).

Vegetables contain high water activity ($a_w$) of 0.970 to 0.996; this amount is sufficient to favor spoilage microorganisms (Barbosa-Cánovas et al., 2003). Pathogen bacteria cannot grow at $a_w < 0.86$; yeasts and molds can be tolerant to low $a_w$, and usually, no growth occurs at $a_w < 0.62$. Also, low $a_w$ of 0.3 prevents lipid oxidation, enzymatic browning, and nonenzymatic browning. Besides, lowering $a_w$ prevents the growth of vegetative microbial cells, germination of spores, and toxin production by molds and bacteria (Barbosa-Cánovas et al., 2003; Erkmen & Bozoglu, 2016). Heating, freeze-drying, dehydration, freeze...
Harvesting

- Reduce field heat
- Reduce microbial activity
- Reduce ethylene production
- Reduce respiration
- Reduce transpiration
- Reduce browning & texture loss

Storage

- Float in ventilated containers
- Put under shade
- Sprinkle water or dip in clean water
- Avoid over filling into the packaging

Packaging

- Use evaporative cooling storage e.g. ZECC
- Maintain optimum storage temperature & humidity
- Precool the vegetables before storage
- Prevent pest and insect damage
- Use ventilated storage containers
- Use cold storage or refrigeration
- Separate from climacteric produce
- Avoid over stacking

Receiving house

- Use perforated packaging
- Use cold packaging or icing box
- Avoid mechanical or physical damage
- Use clean packaging
- Use cold storage or refrigeration
- Use MAP or polymeric film

Selling

- Remove debris
- Washing and sanitizing
- Remove decayed leaves
- Use clean water
- Curing and cleaning
- Destalking/destemming
- Sorting, grading and trimming
- Use fungicide and pesticide preferred organic extract
- Packaging of the cleaned ANS

Transportation

- Avoid customers touching the ANS
- Put away from dust
- Avoid high temperature
- Put away from direct sunlight
- Use cold storage
- Put vegetables in perforated containers
- Keep away from insect and pests
- Use MAP
- Keep away from climacteric produces

Figure 4 Key points to consider during postharvest handling of ANS from farm to market. MAP, modified atmospheric packaging; ZECC, Zero-Energy Cooling Chamber.

Sources: Barbosa-Cánovas et al. (2003); Ekhuya et al. (2018); Gogo et al. (2018); Habwe et al. (2008); Nono-Womdim et al. (2012); Tournas (2005); Paltrinieri (2014).
concentration, crystallization, and osmotic dehydration reduce the $a_w$ of foods (Barbosa-Cánovas et al., 2003; Erkmen & Bozoglu, 2016).

PHH of vegetables has led to safe and nutritious food for the consumers (Shiundu & Oniang’o, 2007; Smith & Eyzaguirre, 2007). Lack of scientific and economic knowledge among farmers and food handlers to develop, use, implement, and sustain PHH systems in SSA results in PHL of ANS. Despite having several effective postharvesting technologies, especially in developed countries (Acedo, 2010), some are difficult to implement in SSA (Arah et al., 2016; Atanda et al., 2011; Kitinoja et al., 2011). Most small-scale farmers and sellers in SSA do not afford to use standard cold rooms due to insufficient power supply and high investment and maintenance costs (Ambuko et al., 2017). Furthermore, farmers have limited knowledge of the benefits of using low-temperature storage (Gogo et al., 2016). Lotter et al. (2014) observed that none of the sellers had access to refrigeration storage in public markets in Tanzania. Lack of low-temperature storage facilities accelerates PHL of ANS and reduces its utilization. Vegetable deterioration affects the qualitative and quantitative loss, including appearance, flavor, texture, shape, and nutritional composition (Affognon et al., 2015; Global Panel, 2018; Wakholi et al., 2015). Also, it causes food and nutrition insecurity in individuals and households in SSA (FAO et al., 2019). Therefore, farmers and food handlers must follow proper PHH techniques from the farm to the fork (Figure 4).

12 | POSTHARVEST LOSSES OF ANS

PHL of AIVs are a significant problem facing SSA (Onyango & Imungi, 2007). The leaves’ quality deteriorates within 4 days of harvest if stored at ambient temperatures (Edmonds & Chweya, 1997). Loss of vegetables in developing countries accounts for about 20% to 50% of total production, reflecting the ANS. Both quantitative and qualitative losses of ANS occur at various stages along the value chain, that is, from harvesting, handling, packing, storage, processing, and transportation to the consumers. According to Gogo et al. (2017), significant losses of nutritional, quantitative, and economic values of ANS in Kenya occur at harvest, transportation, and market. Therefore, proper planning is needed to solve the problem in SSA (Kitinoja & Kader, 2015; Kitinoja et al., 2018). Due to the high demand for nutritious food from the growing populations in SSA, PHL management is unavoidable.

Lack of appropriate PHH technologies results in a high loss of ANS, especially during the peak season. Consequently, limited supply during the off-season is accompanied by high prices, as most of the AIVs are seasonal (Abukutsa-Onyango, 2007; Habwe et al., 2008). Deterioration of vegetables results in partial or total loss of produce; it can happen during preharvest, harvest, and PHH (Acedo, 2010), although factors affecting PHL vary widely from place to place (Munhuweyi, 2012; Yang & Ojiewo, 2013).

Vegetables undergo physiological deterioration and pathological and mechanical/physical damage if not adequately handled (Acedo, 2010; Castro et al., 2005; Munhuweyi, 2012). The incidence of the losses affects the physical and nutritional quality (Munhuweyi, 2012). Physiological deterioration reduces 5% to 10% FW of vegetables, which renders vegetables unsuitable for consumption (Yang & Ojiewo, 2013). Ethylene production results in oxidation of vitamin C, destruction of chlorophyll, yellowing, and increased phenylpropanoid metabolism (Munhuweyi, 2012; Yang & Ojiewo, 2013). A low concentration of ethylene, about 0.01 ppm, accelerates vegetable loss; however, about 0.02 to 0.06 ppm of ethylene resulted in a 10% to 30% loss of product quality (Yang & Ojiewo, 2013). Therefore, the leafy vegetables should be stored away from high ethylene-producing fruits or vegetables (Munhuweyi, 2012; Yang & Ojiewo, 2013).

Furthermore, low crop variety, inappropriate cultural practice, lack of harvesting techniques, unfavorable climate, improper handling, poor storage conditions, lack of competent human resources, lack of technological knowledge, ineffective commercialization and functioning value chain, lack of logistical support, and lack of enabling policy are primary causes of PHL (Acedo, 2010).

Mechanical damage of vegetables causes leafy tearing and crushing, midrib breakage, head cracking, or bruising. It increases oxidation to phenolic compounds and increases susceptibility to decay. Also, damaged leaves increase water loss by about three to four times of undamaged vegetables (Acedo, 2010; Kanlayanarat, 2007). High-temperature storage and high relative humidity cause wilting to the vegetables, which results in loss of freshness of vegetables (Acedo, 2010; Edmonds & Chweya, 1997; Yang & Ojiewo, 2013). Onyango and Imungi (2007) reported 3.5% of *S. scabrum* losses due to excessive wilting. Thus, wilting is a challenge that increases ANS deterioration rate during marketing, especially when vegetables are stored at the ambient temperature (Abukutsa-Onyango, 2007; Onyango & Imungi, 2007). Failure to sell ANS harvested on the same day can result in deterioration due to lack of proper storage facilities (Shiundu & Oniang’o, 2007).

Unfinished day supply of ANS from local markets and supermarkets in Kenya is thrown as waste, thus increasing vegetable loss in SSA (Figure 5), consequently contributing to high PHL (Onyango & Imungi, 2007). Therefore, due to high PHL loss in SSA, there is a need to promote the use of appropriate PHH and PaP technologies to extend the shelf life of AIVs and ensure regular supply from farm to the table in rural, peri-urban, urban all year round (Habwe et al., 2008; Smith & Eyzaguirre,
Reducing PHL of vegetables using appropriate technologies can further increase food availability to the exponentially growing world population and conserve natural resources by reducing the area needed for production (Acedo, 2010) and reducing inputs. It will also reduce fungi and bacteria growths and insect infestation (Acedo, 2010; Bradford et al., 2018) and improve food quality, food safety, and financial opportunities. For that reason, there is a need to develop strategies that could improve the PHH of ANS in SSA. Solar drying and fermentation technologies play a significant role in ensuring the long-term preservation of foods for sustaining food security in SSA (Sivakumar et al., 2020). Therefore, training on feasible and affordable preservation techniques can add value to the ANS in SSA.

Microorganisms are the primary causative agents of PHL of fresh vegetables if they are not well handled (Acedo, 2010; Gil et al., 2015). They contribute about 55% of the total production losses in developing countries (Sanzani et al., 2016). Vegetables can also harbor higher numbers of microorganisms during harvesting due to pre-harvest and postharvest contamination (Tournas, 2005). Bacteria, yeasts, and molds can contaminate the ANS in the value chain. The microbial number of fresh vegetables differs from location, PHH, and storage conditions. Not all microorganisms can grow on fresh vegetables, but some microorganisms can grow and cause spoilage (Tournas, 2005). Cutting and mincing can increase microbial contamination of vegetables (Tournas, 2005). Washing, blanching, sanitary handling, and clean packaging containers and processing environment substantially decrease microbial contamination. Microbial numbers in fresh vegetables per gram differ from species: total bacteria count, \(1.0 \times 10^{10}\); coliforms, \(3.1 \times 10^7\); fecal coliforms, \(5.5 \times 10^6\); and lactic acid bacteria (LAB), \(1.0 \times 10^6\) (Tournas, 2005). The most common spoilage bacteria of vegetables are LAB, pseudomonads, and \textit{Xanthomonas campestris} (Moss, 2008; Tournas, 2005; Wafula, 2017). The fungi \textit{Aspergillus}, \textit{Penicillium}, \textit{Fusarium}, \textit{Alternaria}, and \textit{Mucor} were isolated in spinach and tomatoes (Sanzani et al., 2016; Suleiman et al., 2017). These fungi can also cause spoilage of ANS and production of their toxins (mycotoxins). Sanzani et al. (2016) reported different mycotoxins ( aflatoxin, ochratoxins, and patulin) on fruits and vegetables and fumonisins in tomatoes fruits but the absence of Alternaria toxin in vegetables. Suleiman et al. (2017) observed the presence of aflatoxin (B1, B2, G1, and G2) contamination in fresh spinach, fresh and dried tomatoes, and bitter leaves (\textit{Vernonia amygdalina}).

### 13. Drying techniques

Drying preserves leafy vegetables to make them accessible during the off-season (Maseko et al., 2017; Smith &
Eyzaguirre, 2007). Drying has been used for a century and has been a critical method of preserving vegetables in SSA (Kiringia et al., 2017; Smith & Eyzaguirre, 2007; Vorster et al., 2007). Drying results in a minimal change in texture, flavor, and color (Gogo et al., 2016) and reduced weight, which improves handling, storability, lower transport cost, and microbial degradation of the product (James & Kuiper, 2003; Kiringia et al., 2017; Maseko et al., 2017). Drying is an economically feasible way to preserve AIVs at the local level in most SSA societies (Gogo et al., 2016). There are several drying techniques used for vegetables; however, the standard technique in SSA is open sun-drying, with less use of solar drying (Abukutsa, 2003; Gogo et al., 2016).

13.1.1 Open sun-drying

Sun-drying is the oldest, simplest, and most commonly used method for the preservation of AIVs. It is considered the least expensive method (Atanda et al., 2011; Gogo et al., 2016; Oniang’o et al., 2008). Low-quality sun-dried AIVs are a consequence of physical and biological hazards contamination (Habwe et al., 2008; Keller, 2004). Open sun-drying was reported as a standard technology in areas with low production of vegetables, such as Singida and Kongwa, Tanzania (Keller, 2004), to preserve the vegetables during the off-season. Performing sun-drying under shade for preserving vitamins is highly recommended Match Maker Associates Limited (MMA, 2008; Traoré et al., 2017). About 45% and 55.6% of farmers sun-dry their vegetables in Siaya county Kenya and Bahi district Tanzania (Ayua & Omware, 2013; Kandonga et al., 2019). Sun-drying under shade increased carbohydrate, ash, and lipid content of S. nigrum but reduced protein and β-carotene to 4.03 and 1.76, respectively (Traoré et al., 2017). Besides, sun-drying concentrated mineral elements and reduced antinutritional factors in other AIVs (Kandonga et al., 2019).

13.1.2 Solar drying

The solar dryer is a simple device that reduces the adverse effects of sun-drying. Solar drying technology is more effective in utilizing solar energy (Gogo et al., 2016; James & Kuiper, 2003; MMA, 2008). The indirect solar dryer is a more complicated and expensive solar drying method (MMA, 2008); however, the technology gives high-quality products in terms of nutritional and other physicochemical properties compared to the direct and mixed solar dryers. Solar drying reduced antioxidant activity, flavonoid, vitamin C, and β-carotene in the S. nigrum (Cheptoo et al., 2019); however, it increased carbohydrate and lipids content of ANS (Traoré et al., 2017). Solar drying of S. scabrum reduced vitamin C and β-carotene by 96.2% and 92.7 %, respectively, but it did not affect mineral content (Mibe et al., 2011). Additionally, it retained β-carotene about 693.55 µg/g (68%) of S. nigrum (Nyambaka et al., 2012). Likewise, solar drying of ANS leaves increases sucrose and glucose, with no significant effect on protein and vitamins B1 and B2 and E, and a significant decrease in Vitamin C and ash contents (Wafula, 2017). On the other hand, blanching and solar drying of ANS improved β-carotene and vitamin C content (Kiringia et al., 2017; Nyambaka et al., 2012; Traoré et al., 2017). Solar drying is yet to be fully exploited in SSA countries despite its economic feasibility and quality benefits. Nonetheless, the cost for installing a full mixed solar drier is US$75 (Ayua & Omware, 2013). Therefore, solar drying is a relatively cheap and feasible technique for many small farmers and processors.

13.2 Low-temperature storage

Many studies recommended that most vegetables, including ANS, should be stored at about 10 to 15 °C and 85% to 95% relative humidity to avoid chilling injuries (Castro et al., 2005; Gogo et al., 2016). The technologies to be used in developing countries should suit their needs and be comfortable and less costly than the technology to be applied in developed countries (Arah et al., 2016).

Alternative of inaccessible convection cold rooms have been developed; evaporative cooling (zero-energy cooling chamber) is among these technologies. This technology uses the principle of latent heat of evaporation, and heat exchange occurs when water evaporates; moreover, the chamber does not require electricity to operate. It is appropriate for smallholder farmers and sellers in rural areas with limited electricity supply (Ambuko et al., 2017; Workneh & Woldetsadik, 2004). Additionally, the chamber’s construction is inexpensive, and it uses locally available materials and unskilled labor, making the cost affordable for the resource-poor smallholder farmers (Acedo, 2010; Edmonds & Chweya, 1997; Yang & Ojiewo, 2013). A zero-energy cooling chamber can store the vegetable for 3 to 5 days or more inside the chamber (Yang & Ojiewo, 2013).

13.3 Blanching

Blanching is a primary, intermediate heat treatment process aiming to improve food preservation and quality by inactivating enzymes that can facilitate vegetable spoilage (Habwe et al., 2008; Huang et al., 2016). It also enhances the drying rate, kills plant tissues, improves the peeling of products, removes foreign matters, and reduces oil uptake.
Also, it reduces microbial loads before further processing or storing and removes pesticide and toxic residues. However, timing and temperature of blanching are crucial, as too little temperature is ineffective and too much temperature damages the vegetables and leads to loss of essential nutrients (Huang et al., 2016; Traoré et al., 2017). Blanching is performed using hot water, steam, microwave, radiofrequency, ohmic, and infrared (Liu et al., 2014; Xiao et al., 2017).

Blanching with water at 80 °C for 10 min, followed by solar drying, was shown to retain essential nutrients in AIVs (Acedo, 2010; Halim et al., 2017; Njoroge et al., 2015). On the other hand, blanching, combined with the drying at about 100 °C for 30 min, can degrade essential nutrients in AIVs (Njoroge et al., 2015). Blanching of ANS, followed by lyophilization, can extend the shelf life (Habwe et al., 2008). Also, hot water blanching of ANS at 90 to 92 °C decreases ash content by 1.73% with a high loss of β-carotene but did not affect protein and high carbohydrate content (Traoré et al., 2017). *Solanum scabrum* leaves' phenolic content was reduced by 85% after blanching (Odongo et al., 2018). Blanching of *S. nigrum* reduced antioxidant activity, flavonoid, vitamin C, and β-carotene (Cheptoo et al., 2019) (Table 2). Also, it decreases phytate and tannin contents (Amalraj & Pius, 2015). Besides, blanching improves texture and flavor and retains color (Reis, 2017). Blanching of *S. nigrum* at 100 °C for 30 and 60 min increased carbohydrate but decreased ash content, protein, and β-carotene (Traoré et al., 2017). Therefore, adopting blanching as an intermediate treatment should be recommended as a preservation method for ANS.

### 13.4 Fermentation

Fermentation is a sustainable postharvest strategy for improving quality and product safety in Africa (Wafula et al., 2017). Vegetable fermentation is an inexpensive and attractive preservation technique for small-scale farmers and food processors in SSA. Fermentation is often combined with boiling, drying, salting, steaming, and frying (Adam & Moss, 2008; Adams & Nout, 2001; Wafula et al., 2016). About 32% of people in Kenya use fermentation to preserve ANS and other AIVs (Ayua & Omware, 2013). Fermentation of AIVs such as ANS and cowpeas leaves has been reported in Africa (Kasangi et al., 2010; Oguntoyinbo et al., 2016; Owade et al., 2019; Stoll et al., 2021; Wafula, 2017). Despite high PHL in SSA, fermentation of AIVs has not been given enough priority and is highly underutilized in Africa (Oguntoyinbo et al., 2016; Wafula et al., 2016). Therefore, tapping the potential of ANS fermentation is essential in reducing PHL and improving nutrition and food security.

LAB are convenient starter cultures for fermented plant products, such as fermented pickles. *Lactobacillus plantarum*, *Lactobacillus fermentum*, *Leuconostoc mesenteroides*, *Lactobacillus acidophilus*, and *Lactococcus lactis* are some LAB used (Behera et al., 2020; Wafula et al., 2016). Relishes and other pickled products are prepared from fermented ANS pickles (Hutkins, 2006).

Lactic acid fermentation can preserve vegetables for a long time and retain their original freshness and nutritive value (Madson & Coleman, 2007; Ray & Didier, 2014). LAB improves the bioavailability of vitamins and minerals and production of vitamins such as B vitamins; also, they contribute to the improvement of flavor, texture, and appearance (Madson & Coleman, 2007; Ray & Didier, 2014; Rodríguez et al., 2009; Soccol et al., 2010). *Lactobacillus plantarum* 75 strain did not affect the color of fermented ANS leaves but increases total phenol content and antioxidant activity (Degrain et al., 2020).

During fermentation, LAB produces antimicrobial compounds such as lactic acid, acetic acids, bacteriocins, alcohol, carbon dioxide, diacetyl, hydrogen peroxide, aldehyde, and esters. They inhibit spoilage microorganisms in fermented food, hence ensure food quality (Adam & Moss, 2008; Adams & Nout, 2001; Leroy & De Vuyst, 2004; Ray & Didier, 2014; Soccol et al., 2010; Wafula, 2017). LAB survive and stabilize in fermented vegetables, then dominate other microbial populations. LAB produces acetic acid, lactic, carbon dioxide, bacteriocins, and biosurfactants that inhibit undesirable microorganisms. Also, the acids contribute to lowering the product’s pH (Adam & Moss, 2008; Adams & Nout, 2001). Owade et al. (2021) reported 13 LAB isolates from spontaneous fermentation (SF) of cowpea leaves with *Lactobacillus brevis* and *Lactococcus lactis* dominating the onset stage, with only *Lactobacillus brevis* dominating the onset stage of optimized fermentation.

Lactic acid fermentation can eliminate natural toxins such as phytic acid, oxalate, cyanogen, tannin, and alkaloids and therefore ensure food safety (Fang et al., 2017; Indrastuti et al., 2018; Leblanc et al., 2011; Leroy & De Vuyst, 2004; Muchoki et al., 2010). *Lactobacillus plantarum* 75 strain ANS extract exhibited a 60% reduction of aflatoxin B1-induced DNA damage and a 38% reduction in FeSO₄-induced oxidative stress (Odongo et al., 2018).

Furthermore, it improves digestion in the human gastrointestinal tract and hence increases the absorption of nutrients. Additionally, it improves antioxidant activity by increasing the number of phenolic and flavonoids compounds (Hur et al., 2014). Consumption of fermented food contributes to the lowering of cholesterol, inhibiting infectious diseases and food allergies. Also, it reduces diarrhea, reduces inflammation, and improves digestion of various food in the body, such as the lactose in intolerant people (Deshpande et al., 2011; Hur et al., 2014; Kaprasob et al., 2018; Leroy & De Vuyst, 2004; Soccol et al., 2010).
Vegetable fermentation can be achieved either by spontaneous or controlled fermentation (CF) (Figure 6). In SF, natural flora is the fermenting agent. This technique is simple and affordable even to small-scale processors. Also, back-slopping techniques can be used, where a small quantity of materials from the previous batch of fermented products is used to inoculate the next batch (Kim et al., 2018). In back-slopping, the initial phase of fermentation is short and reduces the risk of fermentation failure (Ghnimi & Guizani, 2018). Furthermore, it reduces the risk of proliferation of contaminant bacteria and acid-resistant bacteria (Adams & Nout, 2001). During SF of cabbage, cucumber, and ANS, heterofermenters, mainly Leuconostoc mesenteroides and L. brevis, initiate the fermentation process, whereas L. plantarum and Pediococcus cerevisiae occur later (Belitz et al., 2009; Hutkins, 2006; Wafuła, 2017).

A commercial starter culture is used in CF, which requires high initial investment and maintenance costs, pausing a challenge to small-scale farmers and processors. Two commercial starter cultures, Lactobacillus plantarum BFE 5092 and Lactobacillus fermentum BFE 6620, were used to ferment nightshade leaves (Wafuła, 2017). In another study by Stoll et al. (2021), Lactiplantibacillus plantarum BFE5092 and Limosilactobacillus fermentum BFE 6620 were two starter culture strains in the fermentation of S. scabrum leaves. Lactobacillus fermentum BFE 6620, isolated from fermented cassava, was reported as a potential starter culture for AIVs (Wafuła et al., 2017). SF and CF significantly affected the nutritional and sensory quality of ANS leaves (Wafuła, 2017). Thus, proper selection of commercial starter cultures is essential in the CF of ANS leaves for value addition (Stoll et al., 2021).

Fermentation of S. scabrum leaves increases ash content and reduces protein and vitamin C by 36–38.3% and 66–81%, respectively (Wafuła et al., 2017). In contrast, S. scabrum leaves’ fermentation decreases vitamins E, B1, and B2 by 10–34%, 68–76%, and 30–47.3%, respectively (Wafuła et al., 2017). Furthermore, fermentation of S. scabrum
reduces chlorophyll and total polyphenol but increases carotenoid content by 67\% (Odongo et al., 2018). Fermentation is a suitable technique for preserving ANS, thus improving nutrition and food security and enhancing the livelihoods of millions of people in SSA (Behera et al., 2020; Stoll et al., 2021; Wafula et al., 2016).

14 GENERAL CHALLENGES FACING ANS

ANS production lacks good extension services from the government to indicate neglect of the crop, despite its significance in improving health and rural livelihoods (Weinberger & Msuya, 2004). Information on ANS yield in most SSA is not available, showing that these vegetables have not been given priority as other vegetables or crops (Edmonds & Chweya, 1997; Weinberger & Msuya, 2004). Furthermore, there is little information on indigenous knowledge on the utilization of ANS; this information could facilitate the consumption, value addition, and commercialization of ANS (Onyango et al., 2016).

Seasonal availability is another challenge as the ANS becomes scarce and expensive during the rainy season or off-season (Lotter et al., 2014). The lack of proper management consequently results in a decline in ANS supply, especially during the off-season (Edmonds & Chweya, 1997; Weinberger & Msuya, 2004). Heavy rainfall reduces the yields, especially for cultivated ANS (Lotter et al., 2014), therefore, screen houses can be used.

Pests and diseases are the key concern that hinders ANS production in SSA (Abukutsa-Onyango, 2007; Sangija et al., unpublished). Lack of appropriate technical skills in farmers such as PHH, processing, and preservation hinders the availability of ANS, especially during the off-season. Some preservation technologies require a high initial cost for the implementation, which becomes difficult for small farmers. Indeed, lack of proper infrastructure hinders the supply of ANS, especially from the production areas to the markets. Production is solely on a small scale by smallholder farmers and local households; therefore, it facilitates the low availability and utilization of ANS. Unsold raw ANS get spoiled due to a lack of storage facilities along the value chain. Of importance, very minimal value addition is done to ANS, resulting in short shelf life, low market price, and low utilization.

Other challenges that hinder ANS utilization are foodborne diseases and their toxin. Common pathogens causing foodborne diseases in vegetables are Listeria monocytogenes, Clostridium spp., Bacillus cereus, Campylobacter jejuni, Salmonella spp., Shigella and Vibrio cholera, and Yersinia spp. and pathogenic strains of Escherichia coli (Balali et al., 2020; Hernández-Cortez et al., 2017; Micali, 2016; Sant’Ana et al., 2011; Waturangi et al., 2019; Zhu et al., 2017). Pathogens can cause gastroenteritis, abortion, and central nervous system infection (Kilonzo-Nthenga & Makuna, 2018; Zhu et al., 2017). Wafula (2017) isolated pathogenic bacteria, that is, Providencia rettgeri, Enterobacter cloacae, E. coli, Klebsiella oxytoca, Enterobacter asburiae, Klebsiella spp., and Enterobacter ludwigi, from fermented ANS leaves. These pathogenic bacteria may cause pneumonia, urinary tract and respiratory infection, and postsurgical peritonitis (Singh et al., 2016; Wie, 2015; Zhu et al., 2017). Also, they produce hemolytic enterotoxins and pore-forming toxins (Davin-Regli & PagÁls, 2015). Simultaneously, E. coli causes diarrhea and produces Shiga toxin, causing severe diarrhea or hemorrhagic colitis (Smith & Fratamico, 2017). However, the presence of pathogenic bacteria differs from sources of ANS (Wafula, 2017).

Also, fungi can contaminate vegetables, but the risk of mycotoxin production is low compared to cereal foods. However, Sanzani et al. (2016) and Suleiman et al. (2017) reported mycotoxin in pepper, fresh spinach, tomatoes and onion; perhaps they can also contaminate ANS. Pathogens can contaminate vegetables at any point along the vegetable value chain (Hernández-Cortez et al., 2017; Kilonzo-Nthenga & Makuna, 2018; Waturangi et al., 2019; Zhu et al., 2017). The contamination sources include animals, insects, water, soil, dirty equipment, human, manure, sewage water, and feces (Kilonzo-Nthenga & Makuna, 2018; Zhu et al., 2017). Therefore, farmers must follow good agricultural practices to avoid contamination. Contaminated vegetables can affect human health and result in several health problems. Mycotoxins are carcinogenic, mutagenic, and teratogenic (Sanzani et al., 2016). The consumption of fresh vegetables should be encouraged, but significant measures that ensure safety before consumption should be in place (Balali et al., 2020).

UV radiation, washing, low-temperature storage, and washing with diluted acetic acid, salted water, and vinegar can control PHH for vegetables (Balali et al., 2020; Kilonzo-Nthenga & Makuna, 2018; Verbikova et al., 2018; Zhu et al., 2017). Additionally, another essential factor in ensuring food safety and quality is the adequate control of time and temperature during processing, cooling, and storing food (Hernández-Cortez et al., 2017). Additionally, fermented pickles should be well pasteurized to ensure safety.

15 POSSIBLE SOLUTIONS AND SUGGESTIONS FOR FUTURE RESEARCH

Through feasible PaP technologies, value addition can improve ANS nutritional and sensory quality, reduce losses, and improve its utilization. Awareness on PaP
techniques and nutritional and health benefits of ANS is necessary for dietary diversity and also for improving nutritional status in communities where malnutrition is prevalent and ANS is highly underutilized and wasted. Furthermore, PaP can ensure the supply of ANS yearly. Also, greenhouse and screen houses should be encouraged because they use low input levels, improve yields, and protect against pests and diseases. Organic pesticides and fertilizer should be promoted over synthesized ones to reduce health effects on consumers and environmental protection.

More emphasis should be on improving ANS production, providing extension services to the farmers, and supporting agricultural inputs, including quality seeds, fertilizer, and organic pesticides. Besides, ANS data documentation for enhancing traceability and control should be encouraged. Specifically, data on production, postharvest handling, value addition, utilization, marketing, cultivars diversity, and improved varieties are missing in most SSA countries due to lack of precise documentation.

Research and development should focus on ANS breeding to improve local cultivars for improved yield, tolerance to climatic change such as drought, resistance to pests and disease, and low-toxicity varieties. Sensitization on the adoption of recently improved ANS varieties such as Nduruma, Ovelolosi, Amburenii, and Malala to improve yield and returns is mandatory. Furthermore, technologies for PaP should be feasible and affordable. Presently, most consumers utilize the leaves despite the nutraceutical potential and commercial value of other ANS parts. Therefore, research focusing on the nutritional, functional, and safety of ANS berries is inevitable for their further utilization.

The promotion of urban farming (UA) in SSA can boost and ensure environmentally friendly food production, that is, increasing green spaces in urban areas, beautifying, and enhancing bioavailability (Stewart et al., 2013). UA is also thought to increase income, prevent hunger and malnutrition, and improve health (Beach, 2013; Stewart et al., 2013). In Uganda, UA has been linked to improving child nutrition, especially for mothers engaging in UA (Beach, 2013). Likewise, in Kampala, about 70% of heads of farming households earn more than the national annual income per capita (US$330) (Prain & Lee-Smith, 2010). Therefore, urban cities can benefit from UA through easy access to fresh and nutritious vegetables, reduced transportation costs and pollution, shortening of the ANS value chain, reducing PHL, easily accessible and available markets, and employment opportunities. Nevertheless, UA is given little recognition, regulation, and support, with only 40% of urban farmers in most SSA countries (FAO, 2012). Therefore, more support is required to improve the efficiency of UA for sustainable livelihood in SSA.

16 | CONCLUSION

ANSs’ contribution to human nutrition is never to be ignored; regardless of the huge PHL, it remains an essential source of livelihood to local SSA communities. Appropriate PHH techniques such as drying and fermentation can further improve ANSs’ distribution and sustainability along the value chain if well adopted. The use of improved ANS varieties can address issues of pests and diseases and safety. Furthermore, good agriculture practices and appropriate PHH are vital in improving ANS production and reducing PHL. The safety of ANS should be taken as a priority as it contains some antinutrients and toxic compounds. Great market opportunities should be tapped by mapping the ANS value chain for income generation. Therefore, good agriculture practices and proper PHH and processing, if well adopted, can further improve ANS supply and hence its contribution to human nutrition and livelihoods in SSA.

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AUTHOR CONTRIBUTIONS

Frank Sangija planned, designed, drafted, and finalized the manuscript. Haikael Martin revised the manuscript. Athanasia Matemu revised the drafted manuscript and proofread and approved the final manuscript.

CONFLICTS OF INTEREST

The authors declare no conflicts of interest.

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