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Molecular identification of *Salmonella* Typhimurium from village chickens based on *invA* and *spvC* genes

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Abstract

Aim: This study aimed to identify *Salmonella enterica* serovars by polymerase chain reaction (PCR) based on virulence genes invasion A (*invA*) and *Salmonella* plasmid virulence C (*spvC*).

Materials and Methods: DNA extraction of eight bacteria isolates was done using the PowerSoil® DNA Isolation Kit. The amplification of *invA* and *spvC* genes was done using conventional PCR. The positive PCR products were purified using the GeneJET Purification Kit and then sequenced using ABI 3730 XL automated genetic analyzer. The sequences obtained were compared for similarities with other *Salmonella* serovars deposited on the NCBI GenBank using BLASTN.

Results: Four out of eight samples were amplified by primers FS139/RS141 that target *invA* gene with products of about 284 bp, and three out of four of the same *invA* positive samples were also amplified by primers FSPV-1/RSPV-2 targeting *spvC* with a product of about 571 bp. One sample was not amplified by primers FSPV-1/RSPV-2 as it lacked virulence plasmid. Analysis of sequences indicated 100% homology with closely related serovars of *S. enterica* subspecies enterica serovar Typhimurium.

Conclusion: *Salmonella* Typhimurium that contained *invA* and *spvC* genes are pathogenic and virulent strains.

Keywords: invasive gene A, polymerase chain reaction, *Salmonella* plasmid virulence gene, *Salmonella* Typhimurium, sequencing.

Introduction

Salmonella enterica serovar Typhimurium (*Salmonella* Typhimurium) is a motile Gram-negative bacterium in family *Enterobacteriaceae* [1]. The bacterium is one among serovars with broad host range responsible for gastroenteritis in chickens but rarely induce systemic infection [2]. In young chickens, *S. Typhimurium* results in severe inflammation and intestinal pathology [3]. However, in some cases, the pathogen does not cause noticeable clinical symptoms in older chickens [4,5]. Under these circumstances, the older chickens become carriers of *S. Typhimurium*, which colonizes the gut with persistent shedding of bacteria that contaminate different environments [6-9]. The carrier chickens with salmonellosis are among sources of contaminated chicken products in the abattoirs with the significance of forming biofilm layers in equipment involved in the value chain [10]. The biofilm contributes to the persistence of *Salmonella* in different biotic

and abiotic surfaces and protects the bacterium against antibiotics and disinfectants [11]. On the other side, salmonellosis is zoonotic; therefore, chicken products contaminated with *S. Typhimurium* increase number of gastroenteritis infections to humans who are the most affected than any other group worldwide [12].

There are different methods of identifying pathogenic microbes from clinical samples [13]. The best choices consider techniques which are cost-effective, accurate, rapid, and easier to use to improve diagnostic testing with the outcome of right treatments in a shorter time [14]. The conventional microbiological methods, including culture using selective medium, microscopy, Gram staining, and biochemical tests are labor-intensive, time-consuming with low specificity [15,16]. In contrary to these methods, the polymerase chain reaction (PCR) is the confident and rapid technique of detecting clinical samples with higher specificity and sensitivity [17,18]. Therefore, the PCR methods qualify for the identification of any number of pathogens using specific primers that amplify the minimal part of the genome. The invasion A (*invA*) is an invasion-related gene located in *Salmonella* pathogenicity island-1 [19]. The gene is responsible for the invasion of epithelial cells and the induction of macrophage apoptosis [20]. The *invA* gene is a conserved virulence gene in nearly all *Salmonella* species; therefore, the right candidate for

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the detection of *Salmonella* using different PCR methods [20]. The study also selected the *Salmonella* plasmid virulence C (*spvC*) gene, which is of virulence plasmid for detecting strains associated with non-typhoid bacteremia [21]. *Salmonella* has plasmids with a highly conserved locus known as *spv* operon. The operon consists of five genes *spvRABCD* essential for *Salmonella* virulence [22]. However, not all *Salmonella* serovars consist of *spv* genes [23].

Therefore, the present study used conventional PCR for the detection of virulence genes *invA* and *spvC* in *Salmonella* spp. isolated from fecal samples of chickens.

Materials and Methods

Ethical approval

This study got approval from Kibong'oto Infectious Diseases Hospital, Nelson Mandela African Institution of Science and Technology (NM AIST) and Centre for Educational Development in Health, Health Research Committee with approval number KNCHREC006.

Bacterial isolates

Eight bacterial isolates used in this study were among 54 bacteria isolates obtained from fecal samples of village chickens in Tengeru ward, Arusha, Tanzania in June 2019. Out of 54 isolates, 46 were biochemically identified to be *Salmonella* Gallinarum (in the other study). The remaining eight isolates were non-*S. Gallinarum* subjected to molecular identification using PCR.

DNA extraction

The DNA extraction was done using the PowerSoil® DNA Isolation kit (MO BIO Laboratories, Inc.) according to the manufacturer's instruction. The quantity and purity of DNA were assessed using Nanodrop, and quality of DNA was checked on a gel of 1.5% (w/v) agarose and visualized using ultraviolet (UV)-visible spectrophotometer (Cole-Parmer UV Transilluminator). The DNA was stored at -20°C until use.

PCR

The PCR amplification was carried out with a 25 µL amplification mixture consisting of 12.5 µL master mix (OneTaq® Quick-Load® 2×MM w/Std. buffer, BioLabs, New England), 0.5 µL of 10 µM each primer (Table-1), [24,25] 8.5 µL of nuclease-free water, and 3.0 µL DNA template. Amplification was conducted in a thermocycler (C-1000 Touch™ thermocycler). The cycle conditions consisted an initial

denaturation 94°C for 1 min followed by 35 cycles of denaturation at 94°C for 60 s, annealing at 64°C for 30 s, and elongation at 72°C for 30 s with 7 min final extension period at 72°C. The amplified products were separated by gel electrophoresis containing 1.5% w/v agarose (SRL, India) stained with ethidium bromide (0.5 µg/mL) and detected by gel documentation system (UVP, UK). The amplification cycles for *spvC* gene were similar to *invA* gene except the annealing temperature for *spvC* gene was 58°C instead of 64°C used for *invA*.

Nucleotide sequencing of *invA* and *spvC* genes

The positive PCR products were purified using the GeneJET Purification Kit (Thermo Fisher Scientific, MA, USA); then, the pure products were sequenced using both forward and reverse primers (Table-1).

A gene in an ABI 3730 XL automated sequencer (Applied Biosystems, MA, USA) was in a custom sequencing facility of Mbeya Referral Hospital, Tanzania. The sequences obtained were analyzed, and homology searches were conducted using the BLAST algorithm (www.ncbi.nlm.nih.gov/BLAST).

Results

Four samples were amplified by *invA* gene. Three out of four of the same *invA* positive samples were also amplified by *spvC* gene (Table-1). The amplified samples were 100% similar with *S. enterica* serovar Typhimurium strain number 14028 with accession number CP034479.1 and *S. enterica* subspecies *enterica* serovar Typhimurium strain number 14028 with accession number CP034480.1

The positive PCR reactions in this study were observed by agarose jelly with ~ 284 and ~571bp products for *invA* and *spvC* genes, respectively (Figure-1). The nucleotide sequence of the *invA* gene of *S. Typhimurium* strain obtained in this study was analyzed and 284 bp sequences deposited with NCBI under GenBank accession number MK204827.

Discussion

In the present study, the detection of *Salmonella* spp. by conventional PCR was done using pair of primers (FS139-RS141) specific for the *invA* gene and FSPV-1 and RSPV-2 for the *spvC* gene. The selection of these genes was made based on those associated with virulence in *Salmonella* spp. [26]. The *invA* gene of *Salmonella* contains sequences unique to this genus and has been approved as a suitable PCR target with

Table 1: Primer pair used to amplify *Salmonella* isolates.

Gene	Primers	Oligonucleotides (5'-3')	Annealing temperature	Length (bp)	Reference
<i>invA</i>	S139	GTG AAA TTA TCG CCA CGT TCG GGC AA	64°C	284	[24]
	S141	TCATCGCACCGTCAAAGGAACC			
<i>spvC</i>	SPV-1	ACTCCTTGACAACCAAATGCGGA	58°C	571	[25]
	SPV-2	TGCTTCTGCATTTCGCCACCATCA			

invA=Invasion A

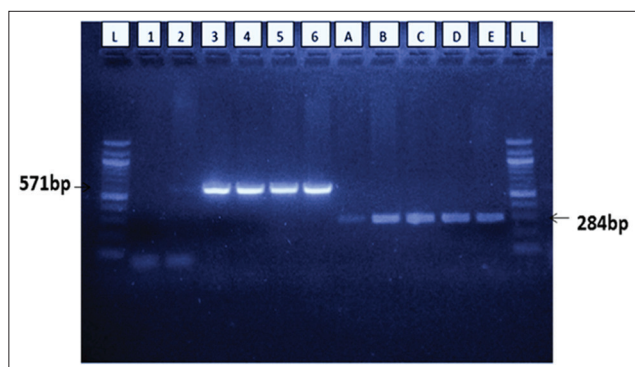


Figure-1: The polymerase chain reaction amplification for *invA* (284bp) and *spvC* (571bp) virulence gene of *Salmonella* spp. L is the molecular weight marker (1000 bp), 1 is a negative control, 2, 3, 4, and 5 are samples, while 6 is positive control. The B, C, D, and E are samples and A is a positive control.

the potential diagnostic application [20]. This gene encodes a protein in the inner membrane of bacteria responsible for the invasion of the epithelial cells of the host [19]. On the other side, *spvC* gene targets invasive non-typhoidal *Salmonella* spp. that induces systemic infection to chickens [27,28].

In this study, out of four samples amplified for *invA* gene, three were positive with *spvC*. Following blasting of the sequences for *invA* positive, *spvC* positive, and *spvC* negative, it was revealed that all samples were *S. enterica* serovar Typhimurium. The lack of *spvC* gene to one sample of *S. Typhimurium* is an indication that the species is non-invasive compared to invasive non-typhoidal *S. Typhimurium* which possess *spvC* gene. In support of these findings, Guiney and Fierer [29] observed that three genes represent virulence phenotype in *spv* locus designated as *spvRABCD* are positive transcriptional regulator *spvR* and two structural genes *spvB* and *spvC* [30]. The *spvBC* are sufficient to replace all of the *spv* genes, as well as the entire virulence plasmid to enable systemic infection in mice after subcutaneous inoculation [31]. However, in the absence of *spvC*, *spvB* does not have a detectable virulence phenotype [21]. From these observations, it is suggested that *spvC* gene is the primary determinant of virulence in *spv* locus. However, the exact mechanisms by which *spvB* and *spvC* manage to enhance virulence are still unclear [31].

It is becoming clear that the virulence of *S. Typhimurium* identified from this study is attributed by the possession of both *invA* and *spvC* genes [32,33]. *S. Typhimurium*, which lacks *spvC* gene, eliminates the ability of plasmid to confer virulence to chickens [21,34].

Conclusion

InvA and *spvC* genes identified pathogenic and virulent strains of *S. Typhimurium*. The *invA* is a factor in the outer membrane of *Salmonella* spp., thus enables *S. Typhimurium* to initiate infection on the epithelial cell of chicken and induction of macrophage apoptosis. *S. Typhimurium* containing *spvC*

gene is more virulent than those lacking this gene. This is because virulence plasmid with a structural gene *spvC* plays a role in the pathogenicity of *S. Typhimurium*. However, not all *Salmonella* serovars possess *spvC* gene responsible for virulence phenotype. Therefore, relying only on the *spvC* gene for the detection of *Salmonella* spp. may provide a high chance of false-positive results because some non-typhoidal *Salmonella* like *S. Typhimurium* possess *spvC* gene and others lack it.

Authors' Contributions

MM planned the study, collected samples, analyzed data, and developed the manuscript. ERM analyzed data, edited, and proofread the manuscript; MC analyzed data and wrote the manuscript. All authors read and approved the final manuscript.

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Competing Interests

The authors declare that they have no competing interests.

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References

1. Zeiner, S.A., Dwyer, B.E. and Clegg, S. (2012) FimA, FimF, and FimH are necessary for assembly of Type 1 fimbriae on *Salmonella enterica* serovar Typhimurium. *Infect. Immun.*, 80(9): 3289-3296.
2. Carattoli, A. (2009) Resistance plasmid families in *Enterobacteriaceae*. *Antimicrob. Agents Chemoth.*, 53(6): 2227-2238.
3. Eguale, T. (2018) Non-typhoidal *Salmonella* serovars in poultry farms in central Ethiopia: Prevalence and antimicrobial resistance. *BMC Vet. Res.*, 14(1): 217.
4. Foley, S.L., Johnson, T.J., Ricke, S.C., Nayak, R. and Danzeisen, J. (2013) *Salmonella* pathogenicity and host adaptation in chicken-associated serovars. *Microbiol. Mol. Biol. Rev.*, 77(4): 582-607.
5. Shehata, A.A., Basiouni, S., Elrazek, A.A., Sultan, H., Tarabees, R., Abd, M.S., Elsayed, E., Talat, S., Moharam, I., Said, A. and Mohsen, W.A. (2018) Characterization of *Salmonella enterica* isolated from poultry hatcheries and commercial broiler chickens. *J. Infect. Dev. Ctries.*, 11(4): 314-319.
6. Silva, C., Calva, E. and Maloy, S. (2014) One health and food-borne disease: *Salmonella* transmission between humans, animals, and plants. *One Health*, 2(5): 137-148.
7. Gopinath, S., Carden, S. and Monack, D. (2012) Shedding light on *Salmonella* carriers. *Trends Microbiol.*, 20(7): 320-327.
8. Cox, N.A., Richardson, L.J., Maurer, J.J., Berrang, M.E.,

- Fedoraka-Cray, P.J., Buhr, R.J., Byrd, J.A., Lee, M.D., Hofacre, C.L., O'kane, P.M. and Lammerding, A.M. (2012) Evidence for horizontal and vertical transmission in *Campylobacter* passage from hen to her progeny. *J. Food Prot.*, 75(10): 1896-1902.
9. Pande, V.V., Devon, R.L., Sharma, P., McWhorter, A.R. and Chousalkar, K.K. (2016) Study of *Salmonella* Typhimurium infection in laying hens. *Front. Microbiol.*, 7: 203.
 10. Paz-Méndez, A.M., Lamas, A., Vázquez, B., Miranda, J.M., Cepeda, A. and Franco, C.M. (2017) Effect of food residues in biofilm formation on stainless steel and polystyrene surfaces by *Salmonella enterica* strains isolated from poultry houses. *Foods J.*, 6(12): 106.
 11. Iñiguez-Moreno, M., Gutiérrez-Lomeli, M., Guerrero-Medina, P.J. and Avila-Novoa, M.G. (2018) Biofilm formation by *Staphylococcus aureus* and *Salmonella* spp. Under mono and dual-species conditions and their sensitivity to cetrimonium bromide, peracetic acid and sodium hypochlorite. *Braz. J. Microbiol.*, 49(2): 310-319.
 12. Dar, M.A., Ahmad, S.M., Bhat, S.A., Ahmed, R., Urwat, U., Mumtaz, P.T., Dar, T.A., Shah, R.A. and Ganai, N.A. (2017) *Salmonella* Typhimurium in poultry: A review. *Worlds Poult. Sci. J.*, 73(2): 345-354.
 13. Guardabassi, L., Damborg, P., Stamm, I., Kopp, P.A., Broens, E.M., Toutain, P.L. and ESCMID Study Group for Veterinary Microbiology. (2017) Diagnostic microbiology in veterinary dermatology: Present and future. *Vet. Dermatol.*, 28(1): 146-e30.
 14. Váradi, L., Luo, J.L., Hibbs, D.E., Perry, J.D., Anderson, R.J., Orenga, S. and Groundwater, P.W. (2017) Methods for the detection and identification of pathogenic bacteria: Past, present, and future. *Chem. Soc. Rev.*, 46(16): 4818-4832.
 15. Sousa, A.M. and Pereira, M.O. (2013) A prospect of current microbial diagnosis methods. In: *Microbial Pathogens and Strategies for Combating Them: Science, Technology and Education*. Vol. 3. Formatec, Badajoz. p1429-1438.
 16. Ahmed, O.B., Asghar, A.H., El-Rahim, I.H.A. and Hegazy, A.I. (2014) Detection of *Salmonella* in food samples by culture and polymerase chain reaction methods. *J. Bacteriol. Parasitol.*, 5(3): 1.
 17. Van Belkum, A., Durand, G., Peyret, M., Chatellier, S., Zambardi, G., Schrenzel, J., Shortridge, D., Engelhardt, A. and Dunne, W.M. (2013) Rapid clinical bacteriology and its future impact. *Ann. Lab. Med.*, 33(1): 14-27.
 18. Gawad, C., Koh, W. and Quake, S.R. (2016) Single-cell genome sequencing: Current state of the science. *Nat. Rev. Genet.*, 17(3): 175-188.
 19. Wang, Y.P., Li, L., Shen, J.Z., Yang, F.J. and Wu, Y.N. (2009) Quinolone-resistance in *Salmonella* is associated with decreased mRNA expression of virulence genes *invA* and *avrA*, growth and intracellular invasion and survival. *Vet. Microbiol.*, 133(4): 328-334.
 20. Shanmugasamy, M., Velayutham, T. and Rajeswar, J. (2011) *InvA* gene specific PCR for detection of *Salmonella* from broilers. *Vet. World*, 4(12): 562.
 21. Derakhshandeh, A., Firouzi, R. and Khoshbakht, R. (2013) Association of three plasmid-encoded *spv* genes among different *Salmonella* serotypes isolated from different origins. *Indian J. Microbiol.*, 53(1): 106-110.
 22. Mazurkiewicz, P., Thomas, J., Thompson, J.A., Liu, M., Arbibe, L., Sansonetti, P. and Holden, D.W. (2008) *SpvC* is a *Salmonella* effector with phosphothreonine lyase activity on host mitogen activated protein kinases. *Mol. Microbiol.*, 67(6): 1371-1383.
 23. Mkrtchyan, M.S., Zakharyan, M.K., Arakelova, K.A., Sedrakyan, A.M., Gevorgyan, Z.U. and Ktsoyan, Z.A. (2016) Molecular determinants of virulence genes of *Salmonella enteritidis* prevailing in Armenia. *Chem. Biol.*, 1(1): 55-60.
 24. Kumar, K., Saklaini, A.C., Singh, S. and Singh, V.P. (2008) Evaluation of Specificity for *invA* Gene PCR for Detection of *Salmonella* spp. Proceeding of 7th Annual Conference of Indian Association of Veterinary Public Health Specialists (IAVPHS).
 25. Chiu, C.H. and Ou, J.T. (1996) Rapid identification of *Salmonella* serovars in faeces by specific detection of virulence genes, *invA* and *spvC*, by an enrichment broth culture-multiplex PCR combination assay. *J. Clin. Microbiol.*, 34(10): 2619-2622.
 26. Pal, S., Dey, S., Batabyal, K., Banerjee, A., Joardar, S.N., Samanta, I. and Isore, D.P. (2017) Characterization of *Salmonella gallinarum* isolates from backyard poultry by polymerase chain reaction detection of invasion (*invA*) and *Salmonella* plasmid virulence (*spvC*) genes. *Vet. World*, 10(7): 814.
 27. Silva, C., Puente, J.L. and Calva, E. (2017) *Salmonella* virulence plasmid: Pathogenesis and ecology. *Pathog. Dis.*, 75(6): 70.
 28. Haneda, T., Ishii, Y., Shimizu, H., Ohshima, K., Iida, N., Danbara, H. and Okada, N. (2012) *Salmonella* Type III effector *spvC*, a phosphothreonine lyase, contributes to reduction in inflammatory response during intestinal phase of infection. *Cell. Microbiol.*, 14(4): 485-499.
 29. Guiney, D.G. and Fierer, J. (2011) The role of the *spv* genes in *Salmonella* pathogenesis. *Front. Microbiol.*, 2: 129.
 30. Guedda, I., Taminiau, B., Ferjani, A., Boukadida, J., Bertrand, S. and Daube, G. (2014) Antimicrobial and molecular analysis of *Salmonella* serovar Livingstone strains isolated from humans in Tunisia and Belgium. *J. Infect. Dev. Ctries.*, 8(8): 973-980.
 31. Matsui, H., Bacot, C.M., Garlington, W.A., Doyle, T.J., Roberts, S. and Gulig, P.A. (2001) Virulence plasmid-borne *spvB* and *spvC* genes can replace the 90-kilobase plasmid in conferring virulence to *Salmonella enterica* serovar Typhimurium in subcutaneously inoculated mice. *J. Bacteriol.*, 183(15): 4652-4658.
 32. Tasmin, R., Gulig, P.A. and Parveen, S. (2019) Detection of virulence plasmid-encoded genes in *Salmonella* Typhimurium and *Salmonella* Kentucky isolates recovered from commercially processed chicken carcasses. *J. Food Prot.*, 82(8): 1364-1368.
 33. García, P., Hopkins, K.L., García, V., Beutlich, J., Mendoza, M.C., Threlfall, J., Mevius, D., Helmuth, R., Rodicio, M.R. and Guerra, B. (2014) Diversity of plasmids encoding virulence and resistance functions in *Salmonella enterica* subsp. *Enterica* serovar Typhimurium monophasic variant 4, [5], 12: I--strains circulating in Europe. *PLoS One*, 9(2): 89635.
 34. Matsui, H., Isshiki, Y., Eguchi, M., Ogawa, Y. and Shimoji, Y. (2014) Evaluation of the live vaccine efficacy of virulence plasmid-cured, and *phoP*-or *aroA*-deficient *Salmonella enterica* serovar Typhimurium in mice. *J. Vet. Med. Sci.*, 77(2): 181-186.
