

2022-04-30

Survival Mechanisms and Management Challenges Associated with Silver Leaf Whitefly on Tomato in Africa: A Review

Mrosso, Secilia

Friends Science Publishers

<https://dspace.nm-aist.ac.tz/handle/20.500.12479/2508>

Provided with love from The Nelson Mandela African Institution of Science and Technology



Review Article

Survival Mechanisms and Management Challenges Associated with Silver Leaf Whitefly on Tomato in Africa: A Review

Secilia E. Mrosso^{1,2*}, Patrick A. Ndakidemi^{1,2} and Ernest R. Mbega^{1,2}

¹School of Life Science and Bioengineering, Nelson Mandela African Institution of Science and Technology, 447, Arusha Tanzania

²Centre for Research, Agricultural Advancement, Teaching Excellence and Sustainability in Food and Nutritional Security (CREATES), Nelson Mandela African Institution of Science and Technology, 447, Arusha, Tanzania

*For correspondence: mrossos@nm-aist.ac.tz; smrosso@yahoo.com

Received 25 January 2022; Accepted 15 March 2022; Published 30 April 2022

Abstract

Silver leaf whitefly (*Bemisia tabaci* Gennadius) (Hemiptera: Aleyrodidae) is a polyphagous winged insect pest that causes high yield losses in tomatoes and other vegetable crops globally. To combat the infestation by the silver leaf whitefly and other insects, tomato growers use cultural and synthetic chemical-based methods. However, the silver leaf whitefly continues to dominate the tomato production systems. Some of the reasons for such continued dominion by the silver leaf whitefly in tomato include among other reasons; little understanding of the mechanisms for survival of the insect pest by tomato production stakeholders which consequently results in difficulties in making appropriate pest management decisions, presence of diverse hosts, the ability of the silver leaf whitefly to develop resistance to synthetic pesticides and ineffective techniques used by tomato grower in combating the insect. Of these challenges, this review discusses the mechanisms for survival of the insect, current pest management options and recommendations for a way forward concerning the silver leaf whitefly management in Africa. © 2022 Friends Science Publishers

Keywords: *Bemisia tabaci*; Survival mechanism; Control techniques; Continued dominion; Joint action

Introduction

Silver leaf whitefly (*Bemisia tabaci* Gennadius) is an invasive insect pest that threatens tomato (*Lycopersicon esculentum* L.) and other various cultivated and weed plant species worldwide (Sri and Jha 2018). In Africa, silver leaf whitefly like in other areas of the world has been reported to affect the crops both directly and indirectly. The direct effect occurs when the silver leaf whitefly nymph or its adult pierces and sucks the plant phloem using the stylets to exploit nutrients, consequently reducing its growth vigour (McKenzie *et al.* 2014). The damaged tomato develops chlorosis, stunting and stopping its growth and making the plant wilt (McKenzie *et al.* 2014). The direct effects also include secretion of honeydew by the insect leading to the development of black sooty mould on plant vegetative parts consequently impairing plant's photosynthetic efficiency, leading to low plant productivity (Mugerwa *et al.* 2021). The indirect effects of silver leaf whitefly on a tomato plant are from its ability to vector > 350 pathogenic plant viruses, which cause diseases of economic importance in tomatoes and other crops in the tropical and subtropical regions (Zhang *et al.* 2014; Ochilo *et al.* 2019). The combined effects of the silver whitefly (whether direct or indirect) causes significant yield losses of up to 100% in tomatoes

which are equated to worth more than one hundred million dollars each year (Moodley *et al.* 2019).

Efforts for managing silver leaf whitefly are always going on; however, the following challenges affect the effectiveness of the management efforts. The main challenges are; the majority of tomato growers in Africa have little understanding of the mechanisms for survival of the insect pest making then ineffectively managing the pest (Laizer *et al.* 2019), presence of diverse host range (Simmons and Riley 2021) and ability of this pest to develop resistance to synthetic pesticides (Legg *et al.* 2014). Thus, this review discusses the survival mechanisms of silver leaf whitefly, its management options and recommendations for a way forward to control silver leaf whitefly in Africa.

Silver leaf whitefly survival mechanisms

Silver leaf whitefly uses different strategies to colonize and survive in different environments (Jiu *et al.* 2017). The most common mechanisms are discussed below:

Life cycle and small body size

The life cycle of silver leaf whitefly goes through three main stages: egg, four larval stages and the adult (Walker *et al.*

2009). The average time between laying eggs to the first larval stage is seven (7) days, and that from first to second and second to third, third to fourth and fourth to adult is 3–4, 2–3, 2–3 and 3–5 respectively (Ghelani *et al.* 2020). This style allows the silver leaf whitefly to produce 11–15 generations per year in tropical climate areas, leading to an increase of its population within a short period (Liu *et al.* 2015; Jiu *et al.* 2017).

The body size of the silver leaf whitefly also gives it an easy survival favor. For instance, at emergence, the size is 0.5 mm long and attains a maximum of 2 mm at the adult. This makes the insect feed on a small amount of food. In addition, males are smaller than females making matting easy and the small sizes of the insects allow them to move from place to place unnoticed through wind and human activities (Kliot *et al.* 2016).

Haplodiploid mode of reproduction

The reproduction systems of silver leaf whitefly are in such a way that females are diploid as they develop from the fertilized eggs, while males are haploid as they grow from unfertilized eggs. This mechanism allows females to control the males and or female ratio making their growth difficult to predict and manage (Gill *et al.* 2015). The males lack one of the genome copies, impairing their fitness as they develops less resistance to environmental pressure, including pesticides, than is the case with the diploid insects (Kliot *et al.* 2016). In the case of positive selection of resistant mutations in silver leaf whitefly males, only individuals with resistant alleles will survive the selection pressure when pesticides are applied, whether they are recessive or dominant. Such individuals will pass the resistant allele to their progeny, resulting in a generation resistant to pesticides that becomes irreversible within a few generations in case of equal male: female ratio resistance and increases the difficulties in controlling the pest (Kliot *et al.* 2016).

Hiding and cuticle waxy materials

The female silver leaf whitefly lays eggs in a circular group under the leaf surface (Sri and Jha 2018) and inserts them in the leaf tissues to hide and protect them from enemies (Vashisth *et al.* 2013). Additionally, all other silver leaf whitefly developmental stages continue under the leaf surface to defend themselves against enemies, sunburn and heat stress, rainwater during heavy rains, and pesticides from overhead spraying (Kumar *et al.* 2017). In addition, the waxy cuticle material covering silver leaf whiteflies' bodies protect them from dehydration, natural enemies, mechanical damage and toxic molecules such as pesticides (Schoeller *et al.* 2018).

Adaptation

Silver leaf whiteflies have a high ability to cope with

different environmental (abiotic and biotic) stresses such as pesticides molecules and high temperatures (Firdaus *et al.* 2013). The pests survive up to the environmental temperatures of 20–30°C (Xiao *et al.* 2016). The symbiotic association between silver leaf whitefly and some primary and secondary bacteria is the tentative reason for silver leaf whitefly survival under these harsh environments (Lv *et al.* 2018).

The primary symbionts provide silver leaf whitefly with essential nutritional elements, especially in a poor diet, for example, *Portiera aleyrodidarum* that synthesize some amino acids and carotenoids, which silver leaf whiteflies cannot synthesize (Skaljic *et al.* 2017). In addition, some secondary symbionts give silver leaf whitefly immunity and affect their development and reproduction (Ferrari and Vavre 2011), while others especially those localized in the salivary glands and the midgut facilitate virus transmission (Rana *et al.* 2012).

Wide host range

Silver leaf whitefly feeds on more than 900 plant species of different families, both cultivated and wild (Gill *et al.* 2015; Alam *et al.* 2016). Such silver leaf whitefly host plants are present in Africa, where the tropical climatic condition favors massive biodiversity (Primack and Corlett 2011). As such, during the offseason, silver leaf whitefly relies on these alternative host plants while waiting for their favorite hosts in the cropping season, thereby increasing their survival chance and enabling them to colonize a wide range of distribution. Additionally, the global trade of silver leaf whitefly host plant materials widened the spread of silver leaf whitefly (Hadjistyli *et al.* 2016).

High genetic diversity of silver leaf whiteflies

The silver leaf whitefly has high genetic diversity seen from a complex of biotypes called cryptic species (Boykin and Barro 2014). Biotypes of the silver leaf whitefly are differentiated based on molecular polymorphism. Such biotype or species differences are expressed in the ability of a particular biotype to cause plant disorders, attract natural enemies, susceptibility to insecticides and resistance expression, host range and capabilities to transmit plant virus (Hadjistyli *et al.* 2016). Also, different biotypes of silver leaf whitefly respond differently to control measures applied which necessitate the knowledge on the silver leaf whitefly biotypes present in a particular place (Naveen *et al.* 2017).

Based on these characteristics, more than 44 silver leaf whitefly cryptic species are reported globally (Acharya *et al.* 2020). However, a recent study found the addition of three silver leaf whitefly species in Uganda, where they were named Sub Saharan Africa 14–16 (Mugerwa *et al.* 2021). In Africa, the Sub Saharan Africa 1–5, Sub Saharan Africa 6 or Uganda 3, Sub Saharan Africa 7, Indian Ocean and East

African 1, the Mediterranean species (MEM) and the Middle East Asia Minor I (MEAM I) are present (Mugerwa *et al.* 2018). Studies show silver leaf whitefly of Sub Saharan Africa species to be the most widely distributed occurring in West, East and South Africa, on contrary, Sub Saharan Africa 4 and 5 occasionally occur in Cameroon and South Africa, respectively (Legg *et al.* 2014). The Middle East Asia Minor I (MEAM I) and Mediterranean silver leaf whitefly species (MEM) are the most invasive and globally distributed, while the Mediterranean species is the most spread (Shadmamy *et al.* 2019; Kriticos *et al.* 2020).

Silver leaf whitefly management options and their associated advantages and challenges

Some control methods and challenges associated with silver leaf whitefly are summarized in Table 1. Nevertheless, the insect is very difficult to control since besides its role as a pest, it carries and transmits viruses that cause economically critical viral diseases (Satar *et al.* 2018). Therefore, the management of silver leaf whitefly as a pest and as a vector for various diseases is essential. Tomato growers apply different Pest Management options globally to reduce infestation by silver leaf whitefly as discussed hereunder:

Integrated pest management (IPM)

IPM is a long term, economical and eco-friendly silver leaf whitefly control strategy of mitigating the adverse effects of pesticides resulting from extensive use of synthetic pesticides during the green revolution (Lamichhane *et al.* 2016; Wilson and Daane 2017; Horowitz *et al.* 2018). The method employs a combination of control methods of silver leaf whitefly such as biological control, modification of cultural practices, the use of resistant varieties and, when needed, judicious and timely use of chemical pesticides (Flint and Bosch 2012).

IPM focuses mainly on monitoring, pest avoidance and practical chemical usage to ultimately develop the most effective and cost-effective solution to silver leaf whitefly management (Legg *et al.* 2014). As such IPM reduced pesticides usage and sustained the reduction of the silver leaf whitefly population with economic benefits of more than \$US 200 million. In Burkina Faso, pyriproxyfen as a growth regulator was used and proved to be effective in controlling silver leaf whitefly and conserving the natural enemies in cotton (Horowitz *et al.* 2018). Planting the tomato associated with aromatic plants reduced silver leaf whiteflies, unlike the case with tomatoes grown as a mono-crop because the volatiles disrupts the silver leaf whitefly development while promoting the development of the host plants in Burkina Faso (Son *et al.* 2018). In South Africa, a mixture of fermented plant extract from neem leaves and wild garlic reduced Silver leaf whiteflies and aphids on tomatoes (Nzanza and Mashela 2012).

Biological control

Biological pest control involves the use of other living organisms such as predators, insect pathogens and parasitoids to reduce the population of another organism such as the silver leaf whitefly (Lenteren *et al.* 2018). The method has been in use for over 2000 years when the augmentative release of *Encarsia* species (Hymenoptera: Aphelinidae) in 1027 successfully controlled greenhouse silver leaf whitefly *Trialeurodes vaporariorum* (Westwood) (Speyer 1927). Biological silver leaf whitefly control can be conservation, natural, augmentative or classical (Cock *et al.* 2010).

Conservation of biological silver leaf whitefly control includes human actions aiming at protecting and stimulating the functioning of naturally occurring natural enemies in the environment, and it is currently receiving much attention (Mendes *et al.* 2011). Natural biological silver leaf whitefly control is an ecosystem service where naturally occurring beneficial organisms reduce the pest population with no human intervention. In contrast, classical biological silver leaf whitefly control through humans collecting the natural enemies from the area of origin and releasing them to the places where the pest is invasive to permanently reduce the problem (Cock *et al.* 2010).

Finally, augmentative biological silver leaf whitefly control can either be inundative or inoculative. In the inundative control, the natural enemies are mass-reared and released in large numbers to have immediate pest control on crops with a short production cycle. In contrast, in inoculative control, the natural enemies are mass-reared and released in large numbers to control pests in several generations in crops with a long production cycle (Lenteren 2012). Below is the review of some biological agents used in controlling silver leaf whitefly.

The use of natural enemies (Parasitoids and Predators)

Parasitoids are insects whose larvae live as parasites that eventually kill their hosts. About 115 species of silver leaf whitefly parasitoids from 23 genera in the family Anatidae, Aphelinidae, Signiforidae, Platygasteridae, Pteromalidae, Encyrtidae, Eupelmidae and Eulophidae, are widely used to control silver leaf whitefly in the tropics (Lahey and Stansly 2015; Ramos *et al.* 2018).

For example, the use of aphelinid (tiny parasitic wasp) from the genus *Eretmocerus*, especially *E. melanoscutus* and *E. Mundus*, proved successful in controlling silver leaf whitefly in Southern America (Navas-Castillo *et al.* 2011). Chalcidoid wasp (*Encarsia Formosa*), a parasitic wasp, is also used in controlling the silver leaf whitefly. The adult lay eggs inside the silver leaf whitefly larvae, and on hatching, the young *Encarsia* feed on the larvae from inside out.

On the other hand, predators are organisms that kill and eat other organisms (Roda *et al.* 2020). However, their

Table 1: Framework of the silver leaf whitefly Control Measures and their Applicability in Africa Farming context

Silver leaf whitefly control Method	Method Description	Applicability	Afford-ability	Reasons for method applicability
Biological Methods: Entomopathogens, parasitoids Predators EPNs	Specific to target pest. Work better in screen houses/greenhouses Less developed in Africa Great opportunity to reduce synthetic pesticide uses	** *	*	Expensive and not common in the Africa farming context African small-scale farmers do not manage greenhouse production (dominate the sector) Require non-contaminated environment for their survival Lack of identity, selection, preparation and application knowledge Rectifies synthetic pesticides problems
Management of fertilizer and irrigation	Crops become succulent and prone to silver leaf whitefly attack	*	*	Africa soils are fertile and with farm yard manure (FYM) from domestic animals, farms have enough Nitrogen. Agriculture in Africa is mostly rain-fed or conducted near water bodies and so a possibility of crops becoming succulent and so attractive to silver leaf whitefly
Fallowing/host free period	The period between successive planting is left in-between seasons- break the host life cycle	**	*	Require communal efforts to create host free period-Require big land and there is a scarcity of fertile land Require knowledge to know alternative host plants to the pest to avoid them
Trap crops	Alternative host crops that attract and hold the pest to reduce the attack to the main crop	***	***	Available and affordable in the Africa farming context Planted as border rows-attract the pest Gives soil organic matter/ animal feed
Reflectance mulch	reflectance created by coloured mulch scares silver leaf whitefly from the host plant	***	*	Poor availability and affordable of mulch materials by most African farmers Suitable for greenhouse crop production as it covers a small area Crops with reflectance characteristics that can be used as mulch are not known to farmers.
Intercropping and companion farming	Different crops planted on the field at the same time pests move from crop to crop-spend less time/ crop	***	***	Applicable in Africa where farmers grow different crops in their fields as a means of diversification unlike mono-cropping in the developed world Farmers have narrow selection of the best plants to be intercropped to reduce the silver leaf whitefly population due to lack of knowledge.
Sticky traps	They are coloured cards with glue. The colour attracts insect pest and the glue stick them on the card to death	***	*	Used for monitoring pest presence and population to alert the farmer before the pest population reaches the economic threshold level Useful in all farm settings.
Screen houses/greenhouses	Constructed by nets or glasses are used to exclude silver leaf whitefly from the crops	***	*	Construction materials are expensive and so not affordable to resource-poor small-scale farmers
Use of resistant crop varieties	Few resistant crop varieties are available and their development requires high investment in terms of funds and knowledge	***	*	Lack of resistant crop varieties There is a lack of knowledge and fund in their development
Chemical Synthetic	Their use is increasing year after year Adopted as first option control means	*	*	Contaminate the environment, non-target organisms, producers and consumers Pesticides residues in crop produces-trade barrier especially in the EU markets Higher production costs-regular purchase of pesticides Silver leaf whitefly develop pesticides resistance Lack of spraying equipment's and techniques burdened by silver leaf whitefly hiding under the leaf surface Poor instruction as most pesticides' container labels is in a foreign language Less available in rural areas Disposing empty containers and remaining pesticides is tricky

*= Less applicable; **= Applicable; ***= Very applicable

effectiveness is influenced by predators and parasitoids number, release rate, intra- and inter-specific competition, stage, size and density of silver leaf whitefly nymph, environmental factors and host plants that influence the performance of silver leaf whitefly pest (Shah *et al.* 2015). For example, releasing *C. carnea* at different rates leads to a substantial reduction of aphids and silver leaf whiteflies in various vegetable crops. Also, the release of *Chrysoperla carnea* at the rate of five larvae per plant effectively controlled the silver leaf whiteflies and aphids in sweet pepper. In comparison, the release of ten larvae per squash plant was adequate in managing the two pests in Saudi Arabia (Alghamdi *et al.* 2018).

The fact that natural enemies such as predators are affected by the environmental conditions in the season

challenge the use of these organisms to control silver leaf whiteflies (Pérez-Hedo *et al.* 2021). Another limitation of using natural enemies is that they can become herbivores and feed on the target crop especially when the target insect pest (prey) population is reduced (scares) or limited by confinement (Urbaneja-Bernat *et al.* 2019; Roda *et al.* 2020). Also, in most cases, natural enemies like parasitoids perform better when applied in greenhouse conditions and combination with other methods. Consequently, selecting suitable individuals for successful silver leaf whitefly control require knowledge that is lacking in most small-scale African farmers. Also, most small-scale African farmers carry tomato production in the open fields. These attributes challenge African crop producers making this method of pest control to have low applicability in Africa (Lenteren 2012). In addition,

Table 2: Key Tomato Production stakeholders and their Inclusive Action in Silver leaf whitefly Management for Increased Tomato Production in Africa

Factor	Challenge	Inclusive action needed
Host plants	Lack of resistant cultivars and wide host range (Gill <i>et al.</i> 2015)	<p> Policymakers: Making policies that emphasize on investing in researching resistant plant materials against insect pests</p> <p> Encourage nations to pool resources and efforts to invest on development of resistant plant materials against silver leaf whitefly to safeguard the crops and the environment.</p> <p> Formulating policies that emphasize on extension services provision to educate farmers on selecting planting materials that withstand pests and disease attacks.</p> <p> Researchers: Developing resistant cultivars through researching on wild plant relatives with resistant genes</p> <p> Growers: Use seeds and planting materials from reliable sources, practising good agricultural practices</p> <p> Practice mixed planting that and intercropping increases biodiversity along the field margins to raise the number of natural enemies (Andrew and Hill 2017)</p>
Environmental	The increased drought accelerates silver leaf whitefly invasiveness (Xiao <i>et al.</i> 2016).	<p> Policymakers: Formulating policies that advocate the effects of climate change and support research on amelioration/restoration of the ecosystem</p> <p> Researchers: Researching on crops that suit new agro-ecological zones</p> <p> Research and promotion of good agricultural practices to increase resilience to climate change</p> <p> Carrying pest risk surveillance frequently to monitor the climate, pest appearance and abundance and keep up to date pest list</p> <p> Developing and using modelling prediction tools to forecast short- and long-term pest populations</p> <p> Researching on appropriate production, processing, storage and distribution technologies that reduce crop pest infestation (Crawford and Tertton 2016).</p> <p> The most commonly mentioned strategies are modified IPM practices, monitoring climate and insect pest populations and the use of modelling predictions tools (Raza <i>et al.</i> 2015)</p> <p> Growers: Using production technologies that are produced and approved by research to be appropriate for a specific agro-ecological zone.</p>
Insect pest level	<p> Tricky infestation strategies -hiding under leaf surface (Kumar <i>et al.</i> 2017).</p> <p> Pesticide resistance development (Moodley <i>et al.</i> 2019)</p> <p> Resisting climate variations (Xiao <i>et al.</i> 2016).</p>	<p> Policymakers To advise governments to invest in research on the development of new strategies for pest management such as the development of new pesticides formulations, repellents and attractants (Gomez-Zavaglia <i>et al.</i> 2020)</p> <p> Advice governments across Africa to invest in research to develop more non-chemicals tomato pest control means to reduce the use of synthetic pesticides to protect the environment and the people involved in agricultural production.</p> <p> Researchers: Development of new pest pesticides formulations that will reach the pest (stick on the pest for contact pesticides).</p> <p> Research on best pesticides application techniques for effective results</p> <p> Research on newer and efficient biological control agents</p> <p> Research on conventional breeding methods like genetic engineering to come up with resistant plant varieties (Gomez-Zavaglia <i>et al.</i> 2020)</p> <p> Growers: Use the approved pesticides and best application methods for best pest control</p> <p> Rely on non-chemical pest control methods and use the chemical pest control method as the last option while well informed on the details of the particular pesticide underuse (Gollin 2018).</p>
Pest control methods	Some pest management options exist although the pest continues to dominate agricultural production systems.	<p> Policymakers: Making policies that put in place pesticides management systems that promote safe, efficient and responsible pesticides use for sustainable agricultural development.</p> <p> Formulating policies that advocate soil conservation, especially in uplands, to encourage the construction of structures such as ridges to reduce runoff speed and silver leaf whitefly spread</p> <p> Researchers: Researching on new and effective pest control methods such as lowering the number of insects to be tolerated before economic yield losses (economic intervention thresholds) are attained</p> <p> Research on simple, cheap, and locally available materials to construct screen houses to allow small resource-poor farmers to practice protected agriculture to minimize silver leaf whitefly attacks on their crops.</p> <p> Isolation and characterization of new predators and parasitoids and find the proper combination for effective pest control.</p> <p> Means of environmental restoration to harbor predators and parasitoids that can kill silver leaf whiteflies.</p> <p> Researching on means of increasing entomopathogenic fungi virulence, stability, and shelf life</p> <p> Research on cheap and safe irrigation methods for small-scale farmers to reduce the spread of silver leaf whitefly through conventional irrigation methods.</p> <p> Establish soil specific tomato plant mineral requirements for improved African tomato production.</p> <p> Growers</p> <p> Prepare their fields to direct excess water to the water bodies and dig dams to receive and store excess water to avoid carrying silver leaf whiteflies from one area to another.</p> <p> Add organic matter to their tomato fields to improve the soil water holding capacity.</p> <p> Applying soil specific tomato plant mineral fertilizers for improved African tomato production.</p> <p> To research the best combination of the IPM methods to come up with a combination that is effective and efficient in reducing insect population to a manageable level.</p> <p> Growers: Adopt pest control methods developed and approved by researchers to be the best n reducing the pest population.</p>
Production cost	<p> Increased pesticide application rates and frequency (Satar <i>et al.</i> 2018)</p> <p> Increased viral diseases (Ochilo <i>et al.</i> 2019)</p>	<p> Policymakers: Addressing proper pesticides usage for safer food production and environmental protection</p> <p> Researchers: More research on pesticides with different modes of action and alternatives to pesticides pest control methods</p> <p> Growers: Seeking education and knowledge on pests and their proper control as they lack such knowledge and application of pesticides with different modes of action to break down pest resistance to pesticides (Matowo <i>et al.</i> 2020)</p>

there are limited number of known predators and parasitoids for controlling silver leaf whitefly pests.

Use of entomopathogens, particularly entomopathogenic fungi (EPF)

Microorganisms such as bacteria, fungus, and viruses can

kill arthropods (Gonzalez *et al.* 2016). Out of all insect-killing entomopathogens, EPF are the most effective against sap-sucking insect pests such as Silver leaf whiteflies worldwide (BuGtl *et al.* 2018), as the fungi directly rupture the host cuticle and enter the silver leaf whitefly haemocoel in the process of sap-sucking unlike bacteria and viruses that need to be ingested by the host (Lacey *et al.* 2015; Dong *et*

al. 2016). Entomopathogens can exhibit a multi-site action, which prevents silver leaf whitefly from resistance development and if rationally used, they aid in managing resistance (Ruiu 2018).

EPF are pathogenic species to insects such as silver leaf whitefly (Maina *et al.* 2018), taking the lead of biopesticides in the world market in controlling silver leaf whitefly, where about 100 mycoinsecticides are commercially available (Jaronski and Mascarin 2017).

Most EPF are from from the phylum entomophthoromycota and Ascomycota in the order Hypocreales, although they are less aggressive when compared with those from the order entomophthorales (Humber 2012). Those from the order entomophthorales can cause dramatic epizootics that rapidly reduce the silver leaf whitefly population. However, their mass production in the laboratory, storage, and formulation is complex, limiting their use as a biological control (Pell *et al.* 2010).

Commonly commercial and mainly used EPF are *Metarhizium anisopliae*, *Lecanicillium or Verticillium*, *Beauveria bassiana*, *Isaria fumosorosea* and *Ashersonia* spp. that cause natural mortality to silver leaf whiteflies at all life stages (Dong *et al.* 2016; Gao *et al.* 2017; Hatting *et al.* 2019). Nevertheless, EPF have a limited shelf life, less stable and slow action, contributing to their slow development as mycoinsecticides (Dong *et al.* 2016). Also, some EPF are less aggressive example Hypocreales (Humber 2012), while mass production in the laboratory, storage and formulation of some EPF like entomophthorales is complex, limiting their use as a biological control (Pell *et al.* 2010). Also, pathogenic insect fungi work better in a controlled environment such as greenhouses, which are not affordable by African farmers who work under open fields, limiting the application of EPF in the African farming context.

Cultural method

The cultural silver leaf whitefly control method is an approach that involves careful management of environmental factors (Spatial and temporal) and production practices to limit silver leaf whitefly damage (Perring *et al.* 2018). Cultural methods, especially those involving several plant species and silver leaf whitefly, may be complex to implement. Still, they are more common in African small-scale diversified farming than in large-scale monoculture production (Walgenbach 2018). Some standard silver leaf whitefly cultural control methods are reviewed hereunder.

Crop management

Crop management involves good agricultural practices, including fertilizers and irrigation methods. The fertilizer and irrigation methods can play a role in controlling silver leaf whiteflies if used judiciously. Nitrogenous fertilizers are applied to supply nitrogen, an essential plant nutrient that promotes growth and determines the ultimate yield and

quality of vegetables through the synthesis and accumulation of free amino acids, proteins, and sugars (Ddamulira *et al.* 2019). Tomato farmers in Africa commonly use nitrogenous fertilizers due to their importance (Ortas 2013). However, nitrogen accumulation in plants directly attracts more Silver leaf whitefly to feed and promote their growth (Walgenbach 2018), oviposition with less feeding (Park *et al.* 2009), and affect silver leaf whitefly's optimal growth in case it is limited. Further, Nitrogen fertilizers increase Silver leaf whiteflies' feeding preference, food consumption, growth, survival, reproduction and population density, thereby increasing the susceptibility of crop plants to sucking pests (Hosseini-Gharalari *et al.* 2015; Bala *et al.* 2018).

On the other hand, the method of irrigation used has a relationship with silver leaf whitefly infestation and the virus occurrence, where drip irrigation is associated with lower silver leaf whitefly densities and virus incidence (Abd-Rabou and Simmons 2012) as the irrigation feeds water at the plant root zone and reduces the possibility of splashing silver leaf whiteflies and the virus to the nearby host. In Africa, about 80 per cent of farmers are small scale who carry out rain-fed agriculture where runoff can push silver leaf whiteflies and spread them widely to nearby farms (Lowder *et al.* 2016).

Fallowing and host-free period

Fallowing and host-free periods are periods created between successive cropping seasons by removing alternative hosts to deprive silver leaf whitefly and the virus from a food source as they can survive all year round with the availability of host plants (Chandel *et al.* 2012). The practice can be altering planting dates to provide as much time as possible between successive crops to reduce silver leaf whitefly and virus density (Perring *et al.* 2018). Nevertheless, the practice can be difficult as silver leaf whitefly has many host plants, including weeds, as detected in over 460 samples of 50 different weed species from 15 plant families (Papayiannis *et al.* 2011). Therefore, in Africa, where the climate permits a wide range of host plants, silver leaf whitefly management requires efforts from all stakeholders.

A host-free period can be created by uprooting old hibernated and hidden host plants at a distance of 10 km² as done in Cyprus to deprive silver leaf whitefly of a food source (Walgenbach 2018). The host-free method is affordable to farmers due to its little cost, making it applicable in Africa. However, farmers lack knowledge of all silver leaf whitefly host plants.

The use of trap crops

Trap crops are alternative hosts to insect pests such as Silver leaf whiteflies that attract, intercept and retain the target pest and reduce their damage to the main crop (Deletre *et al.*

2016). These crops produce volatiles that influences the insect's selection and preference to a particular host plant based on their suitability as the substrate for egg-laying and development (Smith *et al.* 2014). This volatiles produced by the host plants attracts and influence the insect's host selection before the insect lands on the main crop (Luan *et al.* 2013). Examples of trap crops are squash when planted with tomatoes, eggplant planted with maize or eggplant planted with tomatoes (Choi *et al.* 2016). Also *Solanum viarum* growing with tomatoes acted as a trap crop for *Helicoverpa armigera*, prohibited larval growth and survival (Gyawali *et al.* 2021).

The method is of high potential in controlling invasive insects such as Silver leaf whiteflies in crops grown outside greenhouses, particularly in Africa, where farmers grow several crops simultaneously on the same field (Mercader *et al.* 2011). Therefore, there is a need for intercropping repellent plants (push) and attractive plants (pull) to influence pest population and distribution (Khan *et al.* 2008). In the African farming context, this silver leaf whitefly control method is less costly and suitable.

Reflectance mulches

Reflectance mulches are plastic metalized Ultra Violet materials that are used as mulches (Perring *et al.* 2018). These materials interfere with the radiation necessary for the insect's ecological behavior adaptation, by providing surface area for reflection of some light wavelength into the sky, affecting silver leaf whitefly landing behavior and deterring them from the crop (Doukas and Payne 2014; Ojiako *et al.* 2018). The materials can be used in making UV-blocking nets and silvery/white ground coverings that also control silver leaf whiteflies. Nevertheless, living mulches, including other plants, can surround the protected crops to mask them from silver leaf whitefly.

The effectiveness of reflectance mulches depends much on the mulch colour, whereby materials painted with aluminium and aluminium foil, grey and silver are the most effective in controlling Silver leaf whiteflies (Patel *et al.* 2021). These mulches work best during the early crop growth stages, especially in the first 5 weeks after planting, when the virus carried by Silver leaf whiteflies is likely to invade the plant. With time the quality of the mulches decreases due to decreasing of the UV reflectance ability as the mulch gets contaminated with the soil and shaded as the target plant grow and subsequently needs to be changed over time (Smith *et al.* 2000). Unfortunately, this silver leaf whitefly control technology is not reported in Africa, although it is very suitable. Accordingly, African nations can transfer such technology to Africa to help deal with silver leaf whiteflies.

The use of sticky insect traps

Insect traps are used to monitor the silver leaf whitefly

population changes in the greenhouse and conventional cultivation to help determine the insect's entrance site, infestation spots, insect presence and quantity, distribution pattern and species density (Hosseini-Gharalari *et al.* 2015). The traps provide farmers with important information that guides their decisions on when to get the maximum benefits and reduce pesticides usage. The traps may have an adhesive inner layer or adhesive layer mixed with food bait to attract and kill insect pests such as silver leaf whitefly. Furthermore, the traps are of various colours depending on the target insect, although most insects are attracted to yellow, making yellow sticky traps dominant (Pedigo and Rice 2014). However, a combination of colours and food baits containing food material as an attractant and an insecticide might increase the trap's effectiveness against silver leaf whitefly.

Intercropping and companion farming

Intercropping is a practice where multiple crops are planted simultaneously on the same field (Lulie 2017). Intercropping makes efficient labour usage, increases income and diet diversity, stabilizes production, maximizes return under low technology levels, and, most importantly, reduces diseases and pests such as silver leaf whitefly. Furthermore, intercropping allow insect pest such as silver leaf whitefly to feed for a shorter period than when only one host is available as they move from one host plant to the next (Mutisya *et al.* 2016). In the African farming context, many crops that can be intercropped as means to control silver leaf whitefly are available. Some of them are intercropping tomatoes with tubers, cereals and other vegetables (Umeh *et al.* 2002), intercropping tomatoes with onion. Tomatoes grown in combination with basil had the lowest silver leaf whitefly infestation (Son *et al.* 2018). Intercropping tomato and coriander increase the diversity of predatory arthropods and, in turn, reduces the population of tomato leaf miner (Medeiros *et al.* 2009). Also, intercropping maize and leguminous crops showed a significant reduction of Fall Armyworm (FAW) and maize stem borer at the early stages of the crop growth to tasseling in Uganda (Hailu *et al.* 2018). Thus, intercropping allow African farmers to control silver leaf whitefly as most African farmers are familiar with the method.

Using greenhouses and screen houses

Greenhouses and screen houses are constructed by nets and plastic films to provide physical barriers that restrict silver leaf whitefly from accessing the crops and protect the crop from extreme weather conditions leading to improved crop yield (Mutisya *et al.* 2016; Sotelo-Cardona *et al.* 2021). For example, the use of insect nets in the production of cabbage seedlings modified the nursery microclimate, reduced insect pests, and improved cabbage production in Kenya (Muleke *et al.* 2013) while in Tanzania insect nets reduced insect

pests such as silver leaf whitefly in tomatoes and increased yields (Nordey *et al.* 2020). Additionally, the insect nets can be treated with insecticides to prohibit insects pests. An example was using nets treated with alpha-cypermethrin that successfully reduced the population of silver leaf whitefly and black bean aphids in French beans (Martin *et al.* 2013; Gogo *et al.* 2014). The mesh sizes of 230 × 900 µm or less is required to exclude silver leaf whitefly while keeping the structure ventilated (Bethke and Paine 1991). The use of nets with a mesh size smaller than the mentioned will challenge the ventilation needed to manage relative humidity and temperature in the system (Stansly and Naranjo 2010). Also, electrically charged screens effectively control Silver leaf whiteflies, although their application in the African farming context may be complex as the method is expensive (Nonomura *et al.* 2014; Takikawa *et al.* 2016).

The use of resistant plant materials

Resistant plant materials are materials that can produce several secondary chemicals that are either anti-nutritional or toxic, which enable them to survive insect pests such as Silver leaf whitefly (Ndakidemi and Dakora 2003; Vosman *et al.* 2018). Such plants have defense mechanisms that interfere with the insect (silver leaf whitefly) behavioral and or physiological activities (Vosman *et al.* 2018) and directly cause toxicity to the insect or immobilize them on the leaf due to their sticky nature (Rakha *et al.* 2017). The plant defense can be specific to one insect species or attack multiple insect pests to give plant-wide protection. The type and presence of glandular trichomes determine the plant defense. Glandular trichomes are hair-like structures that produce and store plant compounds (metabolites) responsible for plant resistance against enemies such as Silver leaf whiteflies and they are types I, IV, VI and VII (Bleeker *et al.* 2012; Perring *et al.* 2018).

Therefore the abundance of the metabolites produced by a certain tomato species depends on the type of glandular gland present and they correlate with its resistance against a particular insect pest (Firdaus *et al.* 2012). In most cases, metabolites responsible for tomato resistance are produced by wild tomato relatives such as *Solanum galapagense*, *S. pimpinellifolium*, *S. habrochaites* and *Solanum hirsutum* (Lima *et al.* 2016; Ben-Mahmoud *et al.* 2018). As such continuous screening of the existing tomato varieties and their related species for reduced pests (silver leaf whitefly), survival and fitness are essential (Curry and Pimentel 1971); thus, the involvement of crop production stakeholders in the continent is necessary. The use of pest-resistant tomato cultivars is an advantageous method in all farming conditions in Africa.

Chemical control method

The chemical pest control method uses synthetic pesticides to control pests. The method is considered to be highly

effective, convenient and kills a mass of Silver leaf whiteflies within a short period after application (Jiu *et al.* 2017; Naveen *et al.* 2017). Due to these qualities, the chemical pest control method is chosen as the first method of silver leaf whitefly control and it is used by most tomato farmers in tomato growing areas in the world (Laizer *et al.* 2019; Melo *et al.* 2019; Tambe *et al.* 2019; Dube *et al.* 2020). As a result, there is an increase in the use of synthetic pesticides, where from 1990 to 2010 about 342,000 tons of pesticides were used with 25% used in the developing countries and mostly applied in vegetables (Bon *et al.* 2014).

However, a lack of knowledge on pesticides selection and use by farmers leads to increased use of pesticides with a single mode of action to fight silver leaf whiteflies (Laizer *et al.* 2019). As a result, Silver leaf whiteflies developed pesticides resistance, making this control method ineffective (Legg *et al.* 2014). Thus, tomato farmers increased pesticide spraying frequency to control silver leaf whitefly, which accelerated the resistance of silver leaf whitefly to pesticides (Satar *et al.* 2018). As a result, tomato growers shifted to more toxic and banned pesticides, including organochlorines, such as Dichlorodiphenyltrichloroethane (DDT) for silver leaf whitefly control (Dari *et al.* 2016). Due to its long persistence in the environment and residue in crop products, DDT was banned and replaced by pyrethroids in the late 1970 and 1980 (Naveen *et al.* 2017). Farmers also mixed pesticides which increased their synergies in controlling Silver leaf whiteflies. For instance, pyrethroids and a moderate amount of compounds from organophosphates and carbamates were mixed (Castle *et al.* 2014). However, the mixture lost its efficiency due to improper and uncontrolled use, and at the same time, the silver leaf whitefly showed reduced susceptibility to this mixture. Subsequently, newer insecticides entered the market for controlling Silver leaf whiteflies. Among them were the Insect Growth Regulators (IGR), pyriproxyfen, buprofezin and neonicotinoids (Horowitz *et al.* 2018). The use of these newer insecticides increased the threat to the environment, especially the non-target organisms and the consumers' health (Antwi and Reddy 2015; Baffour-Awuah *et al.* 2016).

Recommendation for a way forward about silver leaf whitefly in Africa

From the reviewed materials on the mechanisms for survival of silver leaf whitefly, the study recommends the provision of quality and up-to-date extension services to the tomato producers to equip them with the required knowledge for improved tomato production in Africa. Also, there is a need for joint action of all stakeholders involved in the tomato production value chain in addressing the problems due to silver leaf whitefly as summarized in Table 2, where each stakeholder has a role to play and in totality farmer's production problems are settled. In terms of silver

leaf whitefly control measure used and their suitability in the African context, tomato producers should select the control method that is applicable and affordable to the particular farming context to trap the merits of the method selected as summarized in Table 1.

Conclusion

Silver leaf whitefly threatens tomatoes and other crops of economic importance worldwide, causing substantial financial losses regardless of the available control options. Crop producers employed various silver leaf whitefly management options to keep the population of this pest at a manageable level. The most common silver leaf whitefly control options are chemical pesticides and cultural methods, minimal resistant plant cultivars and biological silver leaf whitefly control. Most cultural silver leaf whitefly control methods such as trap crops, intercropping and companion farming seem highly applicable in Africa. In contrast, some other techniques such as screens and greenhouses, reflectance mulches and resistant plant materials have low applicability among African small scale farmers because they are not affordable by these small and resource-poor farmers. Despite the efforts made to control silver leaf whitefly, the pest is still a big problem in tomato production in Africa. Therefore, there is a need to research more effective ways used elsewhere to control this pest.

Acknowledgements

We acknowledge the Centre for Research, Agricultural Advancement, Teaching Excellence and Sustainability in Food and Nutrition Security (CREATES) through Nelson Mandela African Institution of Science and Technology (P151847) for financial support to my studies that make the production of this review article possible.

Author Contributions

SEM planned the study and write the 1st original draft, and PAN and ERM reviewed and edit the final draft of the manuscript. All authors contributed to finalizing this manuscript.

Conflicts Interests

The authors have declared that no competing interest exists.

References

Abd-Rabou S, AM Simmons (2012). Effect of three irrigation methods on incidences of *Bemisia tabaci* (Hemiptera: Aleyrodidae) and some whitefly-transmitted viruses in four vegetable crops. *CABI Direct* 8:21–26

Acharya R, YK Shrestha, SR Sharma, KY Lee (2020). Genetic diversity and geographic distribution of *Bemisia tabaci* species complex in Nepal. *J Asia Pac Entomol* 23:509–515

Alam MM, MN Islam, MZ Haque, R Humayun, KM Khalequzzaman (2016). Bio-rational management of whitefly (*Bemisia tabaci*) for suppressing tomato yellow leaf curl virus. *Bang J Agric Res* 41:583–597

Alghamdi A, S Al-Otaibi, S Sayed (2018). Field evaluation of indigenous predacious insect, *Chrysoperla carnea* (Steph.) (Neuroptera: Chrysopidae), fitness in controlling aphids and whiteflies in two vegetable crops. *Egypt J Biol Pest Cont* 28:1–8

Andrew NR, SJ Hill (2017). Effect of climate change on insect pest management. *Environ Pest Manage* 197:195–223

Antwi FB, G Reddy (2015). Toxicological effects of pyrethroids on non-target aquatic insects. *Environ Toxicol Pharmacol* 40:915–923

Baffour-Awuah S, AA Annan, O Maiga-Ascofare, SD Dieudonné, P Adjei-Kusi, E Owusu-Dabo, KJP Obiri-Danso (2016). Insecticide resistance in malaria vectors in Kumasi, Ghana. *Parasit Vect* 9:1–8

Bala K, AK Sood, VS Pathania, S (2018). Effect of plant nutrition in insect pest management: A review. *J Pharmacogn Phytochem* 7:2737–2742

Ben-Mahmoud S, JR Smeda, TM Chappell, C Stafford-Banks, CH Kaplinsky, T Anderson, MA Mutschler, GG Kennedy, DE Ullman (2018). Acylsugar amount and fatty acid profile differentially suppress oviposition by western flower thrips, *Frankliniella occidentalis*, on tomato and interspecific hybrid flowers. *PLoS One* 13:1–20

Bethke JA, TD Paine (1991). Screen hole size and barriers for exclusion of insect pests of glasshouse crops. *J Entomol Sci* 26:169–177

Bleeker PM, R Mirabella, PJ Diergaarde, A VanDoorn, A Tissier, MR Kant, M Prins, MD Vos, MA Haring, RC Schuurink (2012). Improved herbivore resistance in cultivated tomato with the sesquiterpene biosynthetic pathway from a wild relative. *Proc Nat Acad Sci* 109:20124–20129

Bon HD, J Huat, L Parrot, A Sinzogan, T Martin, E Malezieux, JF Vayssières (2014). Pesticide risks from fruit and vegetable pest management by small farmers in sub-Saharan Africa. A review. *Agron Sustain Dev* 34:723–736

Boykin LM, PJD Barro (2014). A practical guide to identifying members of the *Bemisia tabaci* species complex and other morphologically identical species. *Front Ecol Evol* 2:45–49

BuGti GA, C Na, W Bin, LH FeNG (2018). Control of plant sap-sucking insects using entomopathogenic fungi *Isaria fumosorosea* strain (Ifu13a). *Plant Prot Sci* 54:258–264

Castle SJ, P Merten, NJP Prabhaker (2014). Comparative susceptibility of *Bemisia tabaci* to imidacloprid in field-and laboratory-based bioassays. *Pest Manage Sci* 70:1538–1546

Chandel R, V Chandla, K Verma, M Pathania (2012). Insect pests of potato in India: Biology and management. In: *Insect Pests of Potato Global Perspectives on Biology and Management*, pp:227–268. Giordanengo P, C Vincent, A Alyokhin (Eds.). Academic Press Elsevier Incorporation

Choi YS, IS Hwang, GJ Lee, GJ Kim (2016). Control of *Bemisia tabaci* Genn.(Hemiptera: Aleyrodidae) adults on tomato plants using trap plants with systemic insecticide. *Kor J Appl Entomol* 55:109–117

Cock MJ, JCV Lenteren, J Brodeur, BI Barratt, F Bigler, K Bolckmans, FL Cónsoli, F Haas, PG Mason, JRP Parra (2010). Do new access and benefit sharing procedures under the convention on biological diversity threaten the future of biological control? *Biocontrol* 55:199–218

Crawford A, A Terton (2016). *Review of current and planned adaptation action in Tanzania*, pp:1–60. CARIAA working paper No. 14.

Curry JP, D Pimentel (1971). Evaluation of tomato varieties for resistance to greenhouse whitefly. *J Econ Entomol* 64:1333–1334

Dari L, A Addo, KA Dzisi (2016). Pesticide use in the production of Tomato (*Solanum lycopersicum* L.) in some areas of Northern Ghana. *Afr J Agric Res* 11:352–355

Ddamulira G, R Idd, S Namazzi, F Kalali, J Mundingotto, M Maphosa (2019). Nitrogen and potassium fertilizers increase cherry tomato height and yield. *J Agric Sci* 11:1–8

Deletre E, F Chandre, B Barkman, C Menut, TJP Martin (2016). Naturally occurring bioactive compounds from four repellent essential oils against *Bemisia tabaci* whiteflies. *Pest Manage Sci* 72:179–189

- Dong T, B Zhang, Y Jiang, Q Hu (2016). Isolation and classification of fungal whitefly entomopathogens from soils of Qinghai-Tibet Plateau and Gansu Corridor in China. *PLoS One* 11:1–12
- Doukas D, CC Payne (2014). Greenhouse whitefly (Homoptera: Aleyrodidae) dispersal under different UV-light environments. *J Econ Entomol* 100:389–397
- Dube J, G Ddamulira, MJA Maphosa (2020). Tomato breeding in sub-Saharan Africa-Challenges and opportunities: A review. *Afr Crop Sci J* 28:131–140
- Ferrari J, F Vavre (2011). Bacterial symbionts in insects or the story of communities affecting communities. *Biol Sci* 366:1389–1400
- Firdaus S, AWV Heusden, N Hidayati, EDJ Supena, R Mumm, RCD Vos, RG Visser, BJT Vosman (2013). Identification and QTL mapping of whitefly resistance components in *Solanum galapagense*. *Theor Appl Genet* 126:1487–1501
- Firdaus S, AWV Heusden, N Hidayati, EDJ Supena, RG Visser, BJE Vosman (2012). Resistance to *Bemisia tabaci* in tomato wild relatives. *Euphytica* 187:31–45
- Flint ML, RVD Bosch (2012). *Introduction to Integrated Pest Management*. Springer Science & Business Media, Berlin, Germany
- Gao T, Z Wang, Y Huang, NO Keyhani, ZIS Huang (2017). Lack of resistance development in *Bemisia tabaci* to *Isaria fumosorosea* after multiple generations of selection. *Sci Rep* 7:1–11
- Ghelani M, B Kabaria, Y Ghelani, K Shah, M Acharya (2020). Biology of whitefly, *Bemisia tabaci* (Gennadius) on tomato. *J Entomol Zool Stud* 8:1596–1599
- Gill HK, H Garg, AK Gill, JL Gillett-Kaufman, BA Nault (2015). Onion thrips (Thysanoptera: Thripidae) biology, ecology, and management in onion production systems. *J Integr Pest Manage* 6:6–14
- Gogo EO, M Saidi, JM Ochieng, T Martin, V Baird, MJH Ngouajio (2014). Microclimate modification and insect pest exclusion using agronet improve pod yield and quality of French bean. *HortScience* 49:1298–1304
- Gollin D (2018). Smallholder agriculture in Africa: An overview and implications for policy IIED 780 working paper. IIED, London. <http://pubs.iied.org/14640IIED/> Accessed: 27 October 2021
- Gomez-Zavaglia A, JC Mejuto, J Simal-Gandara (2020). Mitigation of emerging implications of climate change on food production systems. *Intl Food Res J* 134:1–12
- Gonzalez F, C Tkaczuk, MM Dinu, Ž Fiedler, S Vidal, E Zchori-Fein, GJ Messelink (2016). New opportunities for the integration of microorganisms into biological pest control systems in greenhouse crops. *J Pest Sci* 89:295–311
- Gyawali P, SY Hwang, P Sotelo-Cardona, R Srinivasan (2021). Elucidating the fitness of a dead-end trap crop strategy against the tomato fruitworm, *Helicoverpa armigera*. *Insects* 12:506–524
- Hadjistrylli M, GK Roderick, JK Brown (2016). Global population structure of a worldwide pest and virus vector: Genetic diversity and population history of the *Bemisia tabaci* sibling species group. *PLoS One* 11:1–32
- Hailu G, S Niassy, KR Zeyaur, N Ochatum, SJAJ Subramanian (2018). Maize-legume intercropping and push-pull for management of fall armyworm, stemborers, and striga in Uganda. *Agron J* 110:2513–2522
- Hatting JL, SD Moore, AP Malan (2019). Microbial control of phytophagous invertebrate pests in South Africa: Current status and future prospects. *J Invertebr Pathol* 165:54–66
- Horowitz AR, PC Ellsworth, R Mensah, I Ishaaya (2018). Integrated management of whiteflies in cotton. In: *Compendium of lead invited papers*. pp:155–168. International congress on cotton and other fiber crops. 20–23 February, 2018. Meghalaya, India
- Hosseini-Gharalari A, A Mohammadipour, N Koupi (2015). A new method for analysing sticky-card data in entomology. *Weta* 50:48–54
- Humber RAJM (2012). Entomophthoromycota: A new phylum and reclassification for entomophthoroid fungi. *Mycotaxon* 120:477–492
- Jaronski S, G Mascarin (2017). Mass production of fungal entomopathogens. In: *Microbial Control of Insects Mite Pests*, Vol. 9, pp:141–155. Lacey LA (Ed). Academic Press, London
- Jiu M, J Hu, LJ Wang, JF Dong, YQ Song, HZ Sun (2017). Cryptic species identification and composition of *Bemisia tabaci* (Hemiptera: Aleyrodidae) complex in Henan province, China. *Jns Sci* 17:78–84
- Khan ZR, DG James, CA Midega, JA Pickett (2008). Chemical ecology and conservation biological control. *Biocontrol* 45:210–224
- Kliot A, S Kontsedalov, G Lebedev, M Ghanim (2016). Advances in whiteflies and thrips management. In: *Advances in Insect Control and Resistance Management*, pp:205–218. Horowitz AR, I Ishaaya (Eds). Springer International Publishing, Dordrecht, Germany
- Kriticos D, PD Barro, T Yonow, N Ota, R Sutherst (2020). The potential geographical distribution and phenology of *Bemisia tabaci* Middle East/Asia Minor 1, considering irrigation and glasshouse production. *Bull Entomol Res* 110:567–576
- Kumar A, S Sachan, S Kumar, P Kumar (2017). Efficacy of some novel insecticides against whitefly (*Bemisia tabaci* Gennadius) in Brinjal. *J Entomol Zool Stud* 5:424–427
- Lacey L, D Grzywacz, D Shapiro-Ilan, R Frutos, M Brownbridge, M Goettel (2015). Insect pathogens as biological control agents: Back to the future. *J Invertebr Pathol* 132:1–41
- Lahey Z, PJ Stansly (2015). An updated list of parasitoid Hymenoptera reared from the *Bemisia tabaci* species complex (Hemiptera: Aleyrodidae). *Fla Entomol* 98:456–463
- Laizer HC, MN Chacha, PA Ndakidemi (2019). Farmers' knowledge, perceptions and practices in managing weeds and insect pests of common bean in northern Tanzania. *Sustainability* 11:4076–4086
- Lamichhane JR, S Dachbrodt-Saaydeh, P Kudsk, A Messéan (2016). Toward a reduced reliance on conventional pesticides in European agriculture. *Plant Dis* 100:10–24
- Legg JP, R Shirima, LS Tajebe, D Guastella, S Boniface, S Jeremiah, E Nsami, P Chikoti, C Rapisarda (2014). Biology and management of *Bemisia* whitefly vectors of cassava virus pandemics in Africa. *Pest Manage Sci* 70:1446–1453
- Lenteren JCV (2012). The state of commercial augmentative biological control: Plenty of natural enemies, but a frustrating lack of uptake. *BioControl* 57:1–20
- Lenteren JCV, K Bolckmans, J Köhl, WJ Ravensberg, A Urbaneja (2018). Biological control using invertebrates and microorganisms: Plenty of new opportunities. *BioControl* 63:39–59
- Lima IP, JT Resende, JR Oliveira, MV Faria, DM Dias, NC Resende (2016). Selection of tomato genotypes for processing with high zingiberene content, resistant to pests. *Hortic Bras* 34:387–391
- Liu TX, PA Stansly, D Gerling (2015). Whitefly parasitoids: Distribution, life history, bionomics, and utilization. *Annu Rev Entomol* 60:273–292
- Lowder SK, J Skoet, T Raney (2016). The number, size, and distribution of farms, smallholder farms, and family farms worldwide. *World Dev* 87:16–29
- Luan JB, DM Yao, T Zhang, LL Walling, M Yang, YJ Wang, SS Liu (2013). Suppression of terpenoid synthesis in plants by a virus promotes its mutualism with vectors. *Ecol Lett* 16:390–398
- Lulie B (2017). Intercropping practice as an alternative pathway for sustainable agriculture: A review. *J Agric Sci Res* 5:440–452
- Lv N, L Wang, W Sang, CZ Liu, BL Qiu (2018). Effects of endosymbiont disruption on the nutritional dynamics of the pea aphid *Acyrtosiphon pisum*. *Insects* 9:161–170
- Maina U, I Galadima, F Gambo, DJJOE Zakaria, Z Studies (2018). A review on the use of entomopathogenic fungi in the management of insect pests of field crops. *J Entomol Zool* 6:27–32
- Martin T, R Palix, A Kamal, E Deletre, R Bonafos, S Simon, M Ngouajio (2013). A repellent net as a new technology to protect cabbage crops. *J Econ Entomol* 106:1699–1706
- Matowo NS, M Tanner, G Munhenga, SA Mapua, M Finda, J Utzinger, V Ngowi, FO Okumu (2020). Patterns of pesticide usage in agriculture in rural Tanzania call for integrating agricultural and public health practices in managing insecticide-resistance in malaria vectors. *Malaria J* 19:1–16
- McKenzie CL, V Kumar, CL Palmer, RD Oetting, LS Osborne (2014). Chemical class rotations for control of *Bemisia tabaci* (Hemiptera: Aleyrodidae) on poinsettia and their effect on cryptic species population composition. *Pest Manage Sci* 70:1573–1587
- Medeiros MA, ER Sujii, HC Morais (2009). Effect of plant diversification on abundance of South American tomato pinworm and predators in two cropping systems. *Hortic Bras* 27:300–306

- Melo ADP, INR Zandamela, MD Siteo, CDELD Melo, NM Candido (2019). The use of pesticides in tomato production: Exposition of chokwe farmers-mozambique. *Intl J Res Agric Sci* 6:1–10
- Mendes R, M Kruijt, ID Bruijn, E Dekkers, MVD Voort, JH Schneider, YM Piceno, TZ DeSantis, GL Andersen, PA Bakker (2011). Deciphering the rhizosphere microbiome for disease-suppressive bacteria. *Science* 332:1097–1100
- Mercader RJ, NW Siegart, AM Liebhold, DG McCullough (2011). Simulating the effectiveness of three potential management options to slow the spread of emerald ash borer (*Agrilus planipennis*) populations in localized outlier sites. *Can J For Res* 41:254–264
- Moodley V, A Gubba, PL Mafongoya (2019). A survey of whitefly-transmitted viruses on tomato crops in South Africa. *Crop Prot* 123:21–29
- Mugerwa H, J Colvin, T Alicai, CA Omongo, R Kabaalu, P Visendi, P Sseruwagi, SE Seal (2021). Genetic diversity of whitefly (*Bemisia* spp.) on crop and uncultivated plants in Uganda: Implications for the control of this devastating pest species complex in Africa. *J Pest Sci* 94:1307–1330
- Mugerwa H, S Seal, HL Wang, MV Patel, R Kabaalu, CA Omongo, T Alicai, F Tairo, J Ndunguru, P Sseruwagi (2018). African ancestry of New World, *Bemisia tabaci*-whitefly species. *Sci Rep* 8:1–11
- Muleke E, M Saidi, F Itulya, T Martin, M Ngouajio (2013). The assessment of the use of eco-friendly nets to ensure sustainable cabbage seedling production in Africa. *Agronomy* 3:1–12
- Mutisya S, M Saidi, A Opiyo, M Ngouajio, T Martin (2016). Synergistic effects of agronet covers and companion cropping on reducing whitefly infestation and improving yield of open field-grown tomatoes. *Agronomy* 6:42–55
- Navas-Castillo J, E Fiallo-Olivé, S Sánchez-Campos (2011). Emerging virus diseases transmitted by whiteflies. *BioControl* 49:219–248
- Naveen N, R Chaubey, D Kumar, K Rebijith, R Rajagopal, B Subrahmanyam, S Subramanian (2017). Insecticide resistance status in the whitefly, *Bemisia tabaci* genetic groups Asia-I, Asia-II-1 and Asia-II-7 on the Indian subcontinent. *Sci Rep* 7:1–15
- Ndakidemi PA, FD Dakora (2003). Legume seed flavonoids and nitrogenous metabolites as signals and protectants in early seedling development. *Funct Plant Biol* 30:729–745
- Nonomura T, Y Matsuda, K Kakutani, Y Takikawa, J Kimbara, K Osamura, SI Kusakari, H Toyoda (2014). Prevention of whitefly entry from a greenhouse entrance by furnishing an airflow-oriented pre-entrance room guarded with electric field screens. *J Agric Sci* 6:172–184
- Nordey T, E Deletre, N Mlowe, TJ Martin (2020). Small mesh nets protect tomato plants from insect pests and increase yields in eastern Africa. *J Horticult Sci Biotechnol* 95:222–228
- Nzanza B, P Mashela (2012). Control of whiteflies and aphids in tomato (*Solanum lycopersicum* L.) by fermented plant extracts of neem leaf and wild garlic. *Afr J Biotechnol* 11:16077–16082
- Ochilo WN, GN Nyamasyo, D Kilalo, W Otieno, M Otipa, F Chege, T Karanja, EK Lingeera (2019). Ecological limits and management practices of major arthropod pests of tomato in Kenya. *J Agric Sci* 4:29–42
- Ojiako F, A Ibe, E Ogu, C Okonkwo (2018). Effect of varieties and mulch types on foliar insect pests of okra [*Abelmoschus esculentus* L. (Moench)] in a humid tropical environment. *Agrorsearch* 18:38–56
- Ortas IJ (2013). Influences of nitrogen and potassium fertilizer rates on pepper and tomato yield and nutrient uptake under field conditions. *Sci Res Essays* 7:1048–1055
- Papayiannis L, N Katis, A Idris, JK Brown (2011). Identification of weed hosts of tomato yellow leaf curl virus in Cyprus. *Plant Dis* 95:120–125
- Park MK, JG Kim, YH Song, JH Lee, KI Shin, K Cho (2009). Effect of nitrogen levels of two cherry tomato cultivars on development, preference and honeydew production of *Trialeurodes vaporariorum* (Hemiptera: Aleyrodidae). *J Asia Pac Entomol* 12:227–232
- Patel C, R Srivastava, A Rana, R Kunwar, J Tiwari, K Dudpuri (2021). Evaluation of reflective silver plastic mulch on controlling whitefly and associated disease incidence on tomato crop. *J Entomol Zool Stud* 9:1608–1611
- Pedigo LP, ME Rice (2014). *Entomology and Pest Management*: Waveland Press, Long Grove, Illinois, USA
- Pell J, J Hannam, D Steinkraus (2010). Conservation biological control using fungal entomopathogens. *BioControl* 55:187–198
- Pérez-Hedo M, C Riahi, A Urbaneja (2021). Use of zoophytophagous mirid bugs in horticultural crops: Current challenges and future perspectives. *Pest Manage Sci* 77:33–42
- Perring TM, PA Stansly, T Liu, HA Smith, SA Andreason (2018). Whiteflies: Biology, ecology, and management. In: *Sustainable Management of Arthropod Pests of Tomato*, pp: 73–110. Wakil W, GE Brust, TM Perring (Eds). Academic Press, Elsevier, London
- Primack RB, RT Corlett (2011). *Tropical Rain Forests: An Ecological and Biogeographical Comparison*. John Wiley & Sons, New York, USA
- Rakha M, N Bouba, S Ramasamy, JL Regnard, P Hanson (2017). Evaluation of wild tomato accessions (*Solanum* spp.) for resistance to two-spotted spider mite (*Tetranychus urticae* Koch) based on trichome type and acylsugar content. *Genet Res Crop Evol* 64:1011–1022
- Ramos AS, RNSD Lemos, VA Costa, ALBG Peronti, EAD Silva, JM Mondego, AA Moreira (2018). Hymenopteran parasitoids associated with scale insects (Hemiptera: Coccoidea) in tropical fruit trees in the eastern Amazon, Brazil. *Fla Entomol* 101:273–279
- Rana VS, ST Singh, NG Priya, J Kumar, R Rajagopal (2012). *Arsenophonus* GroEL interacts with CLCuV and is localized in midgut and salivary gland of whitefly *B. tabaci*. *PLoS One* 7:1–13
- Raza MM, MA Khan, M Arshad, M Sagheer, Z Sattar, J Shafi, Eu Haq, A Ali, U Aslam, A Mushtaq (2015). Impact of global warming on insects. *Arch Phytopathol Taylor Franc* 48:84–94
- Roda A, J Castillo, C Allen, A Urbaneja, M Pérez-Hedo, S Weihman, PA Stansly (2020). Biological control potential and drawbacks of three zoophytophagous mirid predators against *Bemisia tabaci* in the United States. *Insects* 11:670–686
- Rui L (2018). Microbial biopesticides in agroecosystems. *Agronomy* 8:235–246
- Satar G, MR Ulusoy, R Nauen, K Dong (2018). Neonicotinoid insecticide resistance among populations of *Bemisia tabaci* in the Mediterranean region of Turkey. *Bull Insectol* 71:171–177
- Schoeller EN, M Yassin, RA Redak (2018). Host-produced wax affects the searching behavior and efficacy of parasitoids of the giant whitefly *Aleurodicus dugesii* (Hemiptera: Aleyrodidae). *BioControl* 121:74–79
- Shadmany M, LM Boykin, R Muhamad, D Omar (2019). Genetic diversity of *Bemisia tabaci* (Hemiptera: Aleyrodidae) species complex across Malaysia. *J Econ Entomol* 112:75–84
- Shah MMR, SZ Zhang, TX Liu (2015). Whitefly, host plant and parasitoid: A review on their interactions. *Asian J Appl Sci* 4:47–60
- Simmons AM, DG Riley (2021). *Improving Whitefly Management*, Vol. 12, p:470. *Insects*: Multidisciplinary Digital Publishing Institute, Basel, Switzerland
- Skaljic M, S Kanakala, K Zanic, J Puizina, IL Pleic, M Ghanim (2017). Diversity and phylogenetic analyses of bacterial symbionts in three whitefly species from Southeast Europe. *Insects* 8:113–131
- Smith HA, CA Nagle, GA Evans (2014). Densities of eggs and nymphs and percent parasitism of *Bemisia tabaci* (Hemiptera: Aleyrodidae) on common weeds in west central Florida. *Insects* 5:860–876
- Smith HA, RL Koenig, HJ McAuslane, R McSorley (2000). Effect of silver reflective mulch and a summer squash trap crop on densities of immature *Bemisia argentifolii* (Homoptera: Aleyrodidae) on organic bean. *J Econ Entomol* 93:726–731
- Son D, I Somda, A Legreve, B Schiffers (2018). Effect of plant diversification on pest abundance and tomato yields in two cropping systems in Burkina Faso: Farmer practices and integrated pest management. *Intl J Biol Chem* 12:101–119
- Sotelo-Cardona P, MY Lin, RJC Srinivasan (2021). Growing tomato under protected cultivation conditions: Overall effects on productivity, nutritional yield, and pest incidences. *Crops* 1:97–110
- Speyer ER (1927). An important parasite of the greenhouse white-fly (*Trialeurodes vaporariorum*, Westwood). *Bull Entomol Res* 17:301–308
- Sri NR, S Jha (2018). Whitefly biology and morphometry on tomato plants. *J Entomol Zool Stud* 107:496–507

- Stansly PA, SE Naranjo (2010). *Bemisia: Bionomics and Management of a Global Pest*. Springer, Dordrecht, The Netherlands
- Takikawa Y, Y Matsuda, T Nonomura, K Kakutani, K Okada, S Morikawa, M Shibao, SI Kusakari, H Toyoda (2016). An electrostatic nursery shelter for raising pest and pathogen free tomato seedlings in an open-window greenhouse environment. *J Agric Sci* 8:13–25
- Tambe AB, BM Mbanga, DL Nzefa, MG Nama (2019). Pesticide usage and occupational hazards among farmers working in small-scale tomato farms in Cameroon. *J Egypt Publ Health Assoc* 94:1–7
- Umeh V, FO Kuku, E Nwanguma, OS Adebayo, A Manga (2002). A survey of the insect pests and farmers' practices in the cropping of tomato in Nigeria. *Tropicultrae* 20:181–186
- Urbaneja-Bernat P, P Bru, J González-Cabrera, A Urbaneja, A Tena (2019). Reduced phytophagy in sugar-provisioned mirids. *J Pest Sci* 92:1139–1148
- Vashisth S, Y Chandel, P Sharma (2013). Entomopathogenic nematodes – a review. *Agric Rev* 34:163–175
- Vosman B, WPV Westende, B Henken, HDV Eekelen, RCD Vos, RE Voorrips (2018). Broad spectrum insect resistance and metabolites in close relatives of the cultivated tomato. *Euphytica* 214:1–14
- Walgenbach JF (2018). Integrated Pest Management Strategies for Field-Grown Tomatoes. In: *Sustainable Management of Arthropod Pests of Tomato*, pp:323–339. Wakil W, GE Brust, TM Perring (Eds). Academic Press, Elsevier, London
- Walker GP, TM Perring, TP Freeman (2009). Life history, functional anatomy, feeding and mating behavior. In: *Bemisia Bionomics and Management of a Global Pest*, pp:109–160. Springer, Dordrecht, The Netherlands
- Wilson H, KM Daane (2017). Review of ecologically-based pest management in California vineyards. *Insects* 8:108–120
- Xiao N, LL Pan, CR Zhang, HW Shan, SS Liu (2016). Differential tolerance capacity to unfavourable low and high temperatures between two invasive whiteflies. *Sci Rep* 6:1–10
- Zhang X, N Yang, F Wan (2014). Population density of *Bemisia tabaci* (Gennadius) Hemiptera: Aleyrodidae) on different plants in the field. *Acta Ecol Sin* 34:4652–4661